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RESEARCH

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Species turnover within cystic fibrosis lung microbiota is indicative of acute pulmonary exacerbation onset

Leah Cuthbertson¹, Lauren Hatfield², Helen Gavillet³, Michelle Hardman⁴, Ryan Marsh³, Damian W. Rivett⁵ and Christopher van der Gast^{3,6*}

Abstract

Background Acute pulmonary exacerbations (PEX) are associated with increased morbidity and earlier mortality for people living with cystic fibrosis (pwCF). The most common causes of PEX in CF are by bacterial infection and concomitant inflammation leading to progressive airway damage. To draw attention to the seriousness of PEX they have been labelled as 'lung attacks', much like a 'heart attack' for acute myocardial infarction. Treatment typically starts when a pwCF presents with worsening respiratory symptoms. Hence, there is a pressing need to identify indicative biomarkers of PEX onset to allow more timely intervention. Set within an ecological framework, we investigated temporal microbiota dynamics to connect changes in the lung microbiota of pwCF to changes in disease states across a PEX event.

Results Species-time relationships (STR) describe how the richness of a community changes with time, here STRs were used to assess temporal turnover (w) within the lung microbiota of each pwCF ($n=12$, mean sample duration 315.9 ± 42.7 days). STRs were characterised by high interpatient variability, indicating that turnover and hence temporal organization are a personalized feature of the CF lung microbiota. Greater turnover was found to be significantly associated with greater change in lung function with time. When microbiota turnover was examined at a finer scale across each pwCF time series, w -values could clearly be observed to increase in the exacerbation period, then peaking within the treatment period, demonstrating that increases in turnover were not solely a result of perturbations caused by PEX antibiotic interventions. STR w -values have been found to have a remarkable degree of similarity for different organisms, in a variety of habitats and ecosystems, and time lengths (typically not exceeding $w=0.5$). Here, we found w -values soon increased beyond that. It was therefore possible to use the departure from that expected norm up to start of treatment to approximate onset of PEX in days (21.2 ± 8.9 days across the study participants).

Conclusions Here, we illustrate that changes in turnover of the lung microbiota of pwCF can be indicative of PEX onset in considerable advance of when treatment would normally be initiated. This offers translational potential to enable early detection of PEX and consequent timely intervention.

Keywords Cystic fibrosis, Lung microbiome, Temporal dynamics, Island biogeography, Species-time relationships, Microbiome ecology, Species turnover, Lung function, Pulmonary exacerbation

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Background

Cystic fibrosis (CF) is characterised by a vicious cycle of recurring respiratory infection and consequent inflammation that gradually damages lung tissue and degrades lung function throughout the life of a person with CF (pwCF) [1, 2]. Additionally, that cycle is further intensified by the frequency and cumulation of acute pulmonary exacerbations (PE_x) experienced through the life of a pwCF [1, 3, 4]. PE_x are associated with increased morbidity, reduced quality of life, and earlier mortality for people living with chronic lung diseases, including cystic fibrosis (CF), non-CF bronchiectasis, chronic obstructive pulmonary disease (COPD), and asthma [2, 5, 6]. As PE_x represent significant life events for people living with such chronic lung diseases they have been labelled as ‘lung attacks’ to call attention to their seriousness, much like a ‘heart attack’ for acute myocardial infarction [1–3]. Although the specific aetiology and underpinning mechanisms that lead to PE_x are poorly understood, the most common causes of PE_x in CF are by bacterial infections triggering an increased inflammatory response, leading to progressive airway damage [1, 2, 7].

While, there is no universally agreed clinical definition of PE_x [1, 8], a useful simple description of PE_x in CF is an acute worsening of respiratory symptoms severe enough to warrant oral or intravenous antibiotics [3]. Whereby initiation of antibiotic intervention in CF is typically based on worsening of lung function [2]; as measured by forced expiratory volume in 1 s (FEV₁) and expressed as a normalized percent of predicted value (%FEV₁) in a pwCF to indicate disease severity [9, 10]. Moreover, antibiotic intervention of PE_x is directed against bacterial pathogens chronically infecting a pwCF [1, 2]. Prevention of PE_x is one of the main goals in CF disease management [11]. Treatment of PE_x typically starts when a pwCF presents as they experience increased respiratory symptoms and an acute decrease in lung function [1, 11]. However, this can lead to subsequent delays in starting antibiotic intervention and in turn could lead to more severe impact of a PE_x [3, 11]. Therefore, early PE_x detection and consequent timely intervention is highly desirable. Microbiological surveillance of respiratory secretions throughout the life of a pwCF is central to clinical care in CF [12, 13]. In a PE_x context, culture-based surveillance is used to inform chronic infection status of canonical CF pathogens, which in turn guides choice of antimicrobial treatment [13, 14]. Plus, following PE_x diagnosis, increased surveillance for the duration of treatment is recommended to further direct and assess efficacy of antibiotic therapy [12, 13]. While culture-based surveillance of targeted canonical pathogens has been useful for PE_x management, it is not predictive of PE_x onset and

has been shown to be both limited and biased for pathogen detection in CF, e.g. [12, 15, 16].

Lower airway infections do not adhere to classical ‘one microbe, one disease’ concepts of infection pathogenesis born out of Koch’s postulates [12, 17, 18]. Instead, pulmonary infections in CF and across a range of chronic lung diseases are comprised of diverse and temporally dynamic lung infection microbiota [17–21]. Moreover, it is the case that ecological communities, whether animal, plant, or microbial, are inherently dynamic through time [22]. Wherein species constantly turnover within years, months, weeks, or even days and hours in the case of microbial species within a microbiome [22]. Yet, our understanding of human microbiota has relied heavily on spatial organization of microbial species and less on how those species are organized in time [17, 22, 23]. As such, it has been advocated that temporal dynamics set within an ecological framework need to be considered when attempting to connect change in human microbiota to change in disease status [17, 20, 24]. For example, the theory of island biogeography has proven to be a useful framework to apply to the respiratory microbiome, as the lungs can be considered as dynamic ‘island’ habitats subject to immigration and extinction of species through time [17, 25, 26]. Crucially, for understanding temporal microbiota dynamics it has temporal turnover, defined as “the number of species eliminated and replaced per unit of time”, as its central concept [17, 23].

Set within that framework and utilising species-time relationships (STRs) to measure temporal turnover, we recently determined the ecological patterns and processes underpinning temporal turnover of the chronic and intermittently infecting members of dynamic lung infection microbiota from individual children and adults with CF [17]. CF lung microbiota have high interpatient variability and are highly personalized to the individual pwCF [9], expanding upon that spatial principle we found that temporal organization, as measured with STRs, is also a highly personalized feature of the CF lung microbiome [17, 24]. We concluded that this approach incorporating a personalized patient focus has translational potential to guide and improve therapeutic targeting of lung microbiota in chronic lung diseases, including in CF.

In the current study, we applied this ecological approach to gain novel understanding of the longitudinal lung microbiota dynamics across a PE_x event within individual adults with CF (awCF). Specifically, we sought to determine whether changes in bacterial species turnover within the lung microbiota of individual awCF could be indicative of PE_x onset. We achieved this by retrospectively analysing longitudinal microbiota and clinical data collected from individual awCF over the course of a PE_x [27]. To differentiate between disease status leading up to

and after PEx in individual patients, we applied the useful 'BETR' disease state classifications to each individual awCF longitudinal dataset; (B) baseline, (E) exacerbation, (T) treatment, and (R) recovery [27, 28].

Methods

Study and patient sampling

This retrospective study utilises longitudinal data derived from a prospective study that focused on combined cohort-wide changes in lung microbiology through BETR disease periods [27]. The study was reviewed and approved by the Southampton and Southwest Hampshire Research Ethics Committee, UK (06/Q1704/24). All patients provided written informed consent. Sputum samples were collected from 12 awCF at Southampton General Hospital, Southampton, UK. Participating awCF were selected based on having experienced at least one PEx in the 12 months prior to the start of the study and were persistently sputum productive. Diagnostic culture-based microbiology indicated that all participating awCF were chronically colonised with *Pseudomonas aeruginosa* within 12 months prior to start of study, as such all participants were all receiving antibiotics with known activity against this pathogen. Patients were assessed at their usual clinic appointments by their regular clinical team and included routine and emergency visits. Spirometry was performed by their usual clinical team. The clinical characteristics of patients included in the current study are summarised in Table 1.

All sputum samples were stored at 4 °C immediately after spontaneous expectoration and transferred to –80 °C storage within 12 h of production [29, 30]. Decisions to start and end treatment for PEx were directed by treating clinicians based on worsening and subsequent stabilisation or improvement of clinical symptoms, respectively. Samples and data were retrospectively partitioned into five disease periods: (B₀) baseline pre-PEx; (E) PEx, within 30 days prior to start of antibiotic treatment; (T) period of time patients were receiving treatment for pulmonary exacerbation; (R) recovery, within 30 days post-treatment for PEx; and (B₁) baseline post-PEx [27, 28]. During the study period 10 of the 12 awCF were treated for PEx. The two awCF (P3 and P7) that remained at baseline (B₀) served as a useful comparison.

Sequencing

Sputum was washed with phosphate-buffered saline to remove saliva, as previously described [31]. DNA from dead or damaged cells, as well as extracellular DNA, was excluded from analysis via cross-linking with propidium monoazide prior to DNA extraction, as previously described [32]. Subsequent DNA extraction, 16S rRNA gene targeted amplicon sequencing via barcode

encoded FLX amplicon pyrosequencing, and sequence analyses within the mothur sequencing analysis platform are described in detail in Cuthbertson et al. (2016) [27]. Given the length of the ribosomal sequences analysed, species identities should be considered putative. The raw sequence data reported in this study have been deposited in the European Nucleotide Archive under accession numbers PRJEB5815 and PRJEB7346. The relevant barcode information for each sample along with anonymised clinical metadata, processed microbiota data have been deposited at figshare.com under <https://figshare.com/s/f34060344dba52278861>.

Statistical analyses

All regression analyses, coefficients of determination (R^2), degrees of freedom, F -statistics, and significance (P) were calculated using XLSTAT v2018.1 (Addinsoft, Paris, France). Kruskal–Wallis analyses in conjunction with the post-hoc Dunn test, were also performed in XLSTAT. Persistence-abundance relationships (PARs) were used to visualize the temporal distribution of all bacterial taxa, including chronic and intermittently colonizing taxa, within each awCF time series [17]. Using a modification of the Leeds criteria [12, 14], individual awCF were considered chronically or intermittently colonized with a given bacterial taxon if it had >50% or ≤50% temporal persistence, respectively [17].

Species-time relationships (STRs) are well established ecological methods for measuring temporal turnover in communities of animals, plants, and microorganisms [17, 18]. STRs have been used to describe how the observed taxon richness of a community in a habitat of fixed size increases with the length of time over which the community is monitored [33, 34]. The STR is modelled with the power regression equation, $S = cT^w$. Where S is the cumulative number of observed taxa over time T , c is the intercept, and w is the slope of the line or temporal scaling exponent, where increasing values of w can be taken as greater rates of turnover [35]. Here, STRs were used to assess species turnover within each awCF over each time series. STRs were constructed using the moving window method, as previously described [23]. Unlike other STR construction approaches, this method incorporates immigration of new species, local extinctions, and recolonizations of same taxa through time, which would be anticipated in a time series of this study's extent [23].

To examine changes in lung function, cumulative changes in individual awCF lung function with time were calculated by taking cumulative absolute change in %FEV₁ between consecutive timepoints across each time series. Plots of cumulative lung function change versus time were then fitted with a power regression model, $\Delta\%FEV_1 = aT^b$. Where $\Delta\%FEV_1$ is cumulative change in

Table 1 Clinical characteristics of individual patients

Patient	Age ^a	Sex ^b	CFTR genotype ^c		Genotype 2	%FEV ₁ ^d	Pancreatic Insufficient	CF-related diabetes	PEx antibiotics ^e		Intravenous	PEx frequency ^f
			Genotype 1						Oral			
1	30	M	F508 del	NK		57.8 ± 5.8	No	No	Ciprofloxacin			3
2	45	F	F508 del	F508 del		25.2 ± 5.6	Yes	Yes			Colistin + Tobramycin	5
3	47	M	F508 del	NK		36.5 ± 2.8	Yes	Yes				4
4	22	F	F508 del	F508 del		30.7 ± 4.9	Yes	No	Ciprofloxacin		Meropenem + Amikacin	5
5	55	M	F508 del	G58E		81.1 ± 5.0	Yes	No			Ceftazidime + Gentamicin	2
6	21	F	F508 del	F508 del		61.2 ± 6.9	Yes	No	Ciprofloxacin			3
7	40	M	F508 del	F508 del		60.8 ± 9.2	Yes	Yes				3
8	22	M	F508 del	F508 del		28.6 ± 5.6	Yes	Yes			Meropenem + Colistin	6
9	17	F	F508 del	F508 del		73.6 ± 6.5	Yes	No			Ceftazidime + Gentamicin	3
10	24	F	F508 del	G542X		64.4 ± 7.1	Yes	No	Clarithromycin			3
11	20	M	F508 del	F508 del		29.7 ± 7.8	Yes	No	Ciprofloxacin		Metronidazole	4
12	20	M	F508 del	F508 del		45.9 ± 3.3	Yes	No			Ceftazidime + Gentamicin	4

^a Age in years
^b Sex: *F* female and *M* male
^c Cystic fibrosis transmembrane conductance regulator (CFTR) genotype (NK = not known)
^d Lung function—Mean %FEV₁ and standard deviation of the mean (SD) over course of study
^e Antibiotic interventions for pulmonary exacerbation (PEx) by oral or intravenous administration
^f Number of PEx in 12 months prior to start of study

lung function, a is the intercept, T is time in days, and b is the rate of change in lung function over time.

Results

Here, we analysed respiratory samples from 12 awCF collected over the course of a PEx. The clinical characteristics of individual patients are summarised in Table 1. Two of the 12 awCF (P3 and P7) did not experience a PEx during the course of the study and hence were at baseline (B_0) throughout. For those that did experience a PEx, the mean ($\pm$ standard deviation of the mean [SD]) sample period was 315.9 ± 42.7 days (minimum = 221, maximum = 368 days). The mean ($\pm$ SD) number of samples for those awCF was 22.2 ± 5.1 (minimum = 16, maximum = 35 days). For P3 and P7 the duration was 313 and 300 days, and with 9 and 6 samples collected over those periods, respectively.

Chronic and intermittent colonization

To visualise the temporal distributions of every chronic and intermittently colonizing bacterial taxon within each patient, the longitudinal persistence of every bacterial taxon observed in the lung microbiota of an awCF was plotted against its mean relative abundance across the temporal samples it was detected in (Fig. 1). The resulting PARs for each patient were all positive and significant, in that chronic infecting taxa were persistent and common while the intermittent infecting taxa were infrequent and rare. The mean number ($\pm$ SD) of chronic and intermittent taxa across all patients was 12.6 ± 7.0 (minimum = 5, maximum = 31) and 49.6 ± 21.4 (minimum = 18, maximum = 90), respectively (Fig. 1). All patients were chronically infected with *Pseudomonas aeruginosa*. P1 was also chronically colonized with *Staphylococcus aureus* and *Stenotrophomonas maltophilia*. P5 and P7 were also chronically colonized with the latter. While P11 was also chronically colonized with *Achromobacter xylosoxidans*. Furthermore, PARs indicated all patients were all intermittently colonized with at least one and as many as four canonical CF pathogens (Fig. 1).

Species-time relationships and temporal turnover

Temporal turnover within the lung microbiota of each awCF was measured using the slope (w) values from STRs fitted to each whole-time series (Fig. 2). For each awCF, STRs for the whole microbiota as well as the chronic and intermittent colonizing taxa were plotted. The mean w ($\pm$ SD) for the whole microbiota was 0.421 ± 0.120 (minimum = 0.306, maximum = 0.647). The mean w ($\pm$ SD) for the chronic colonizing taxa was 0.255 ± 0.093 (minimum = 0.128, maximum = 0.438), where temporal turnover was significantly lower than observed in the whole microbiota (Kruskal–Wallis: $H=$

9.01, $P=0.003$). Conversely, turnover in the intermittent colonizing taxa group (mean $w=0.560 \pm 0.159$ [minimum = 0.352, maximum = 0.838]) was significantly higher than in the whole microbiota ($H=5.88$, $P=0.015$). Turnover in the intermittent colonizing taxa was also significantly higher than in the chronic colonizing taxa group ($H=14.10$, $P<0.0001$). When the BETR disease state framework was superimposed to each patient time series a notable increase in cumulative taxa richness was observed in the exacerbation and treatment states for the whole microbiota and intermittent colonizing taxa, and to a lesser extent for the chronic colonizing taxa (Fig. 2). Conversely, no notable increases in taxa accumulation were observed in the awCF that did not experience a PEx during the study (P3 and P7).

Changes in lung function with time

Next, lung function-time relationships were plotted to investigate the cumulative absolute change in %FEV₁ across each awCF time series (Fig. 3). Like the STRs, the cumulative absolute change in lung function ($\Delta\%$ FEV₁) over time was observed to notably increase within the exacerbation and treatment diseases states. However, notable increases in cumulative $\Delta\%$ FEV₁ were absent for the awCF that did not experience a PEx within the study (Fig. 3). Moreover, the rate of change in lung function over time (b) was typically higher for patients that experienced a PEx (mean $b=0.214 \pm 0.080$) when compared to those that did not (b for P3 and P7 was 0.100 and 0.143, respectively). Changes in %FEV₁ through time are presented in Fig. S1.

Relationships between changes in lung function and temporal turnover

When relationships between temporal turnover and lung function were explored, positive significant correlations were found between the rate of change in lung function (b) and STR derived turnover values (w) values from the whole microbiota, and the chronic and intermittent colonizing taxa groups (Fig. 4). In each instance, greater changes in lung function were accompanied with greater changes in temporal turnover (Fig. 4). Conversely, no significant relationships between mean %FEV₁ over the course of each awCF time series and turnover (w) were observed (Microbiota, $F_{1,10}=0.99$, $R^2=0.09$, $P=0.343$; Chronic, $F_{1,10}=0.99$, $R^2=0.17$, $P=0.188$; and intermittent, $F_{1,10}=0.05$, $R^2=0.05$, $P=0.825$).

Rates of turnover at finer temporal scales through PEx

To further elucidate relationships between turnover, lung function, and BETR disease states through time, the temporal scale examined was changed from the entire time course to a finer scale, incorporating a moving 5

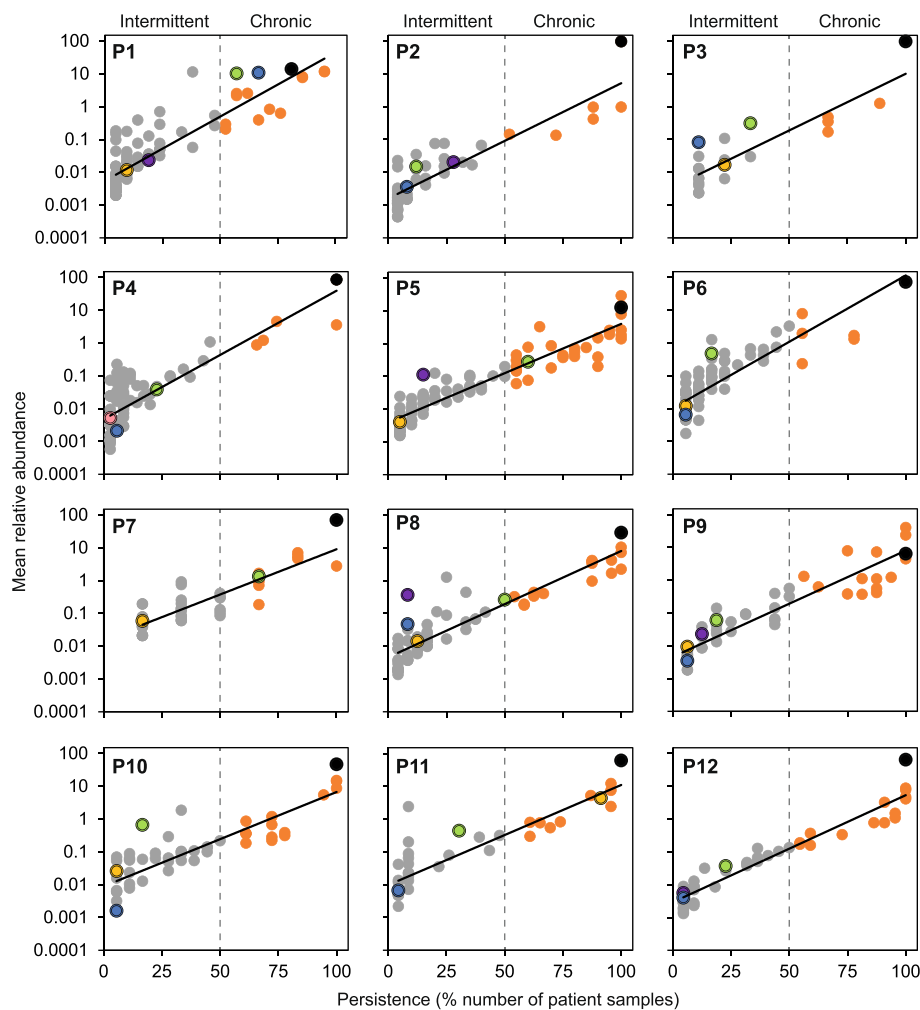


Fig. 1 Persistence and abundance of chronic and intermittent colonizing bacterial taxa within patients. Using a modification of the Leeds criteria of chronic infection, patients were deemed to be chronically (orange circles) or intermittently (grey) colonized with a given bacterial taxon if present in $>50\%$ or $\leq 50\%$ samples, respectively. Canonical pathogens are marked as follows: *Pseudomonas aeruginosa*, black circle; *Staphylococcus aureus*, blue; *Stenotrophomonas maltophilia*, green; *Burkholderia cepacia* complex, pink; *Haemophilus influenzae*, purple; and *Achromobacter xylosoxidans*, yellow. Chronic or intermittent colonization status for all bacterial taxa within the microbiota of each patient is highlighted in the supplemental microbiota data (see “Data availability” section). Persistence–abundance relationship regression statistics: (P1) $R^2 = 0.77$, $F_{1,88} = 290.1$; (P2) $R^2 = 0.77$, $F_{1,47} = 168.9$; (P3) $R^2 = 0.78$, $F_{1,21} = 76.2$; (P4) $R^2 = 0.64$, $F_{1,93} = 166.5$; (P5) $R^2 = 0.88$, $F_{1,92} = 642.1$; (P6) $R^2 = 0.72$, $F_{1,84} = 220.6$; (P7) $R^2 = 0.76$, $F_{1,40} = 125.5$; (P8) $R^2 = 0.84$, $F_{1,64} = 346.4$; (P9) $R^2 = 0.85$, $F_{1,49} = 284.8$; (P10) $R^2 = 0.74$, $F_{1,53} = 153.1$; (P11) $R^2 = 0.80$, $F_{1,42} = 170.1$; (P12) $R^2 = 0.93$, $F_{1,49} = 675.1$. All relationships were significant ($P < 0.0001$ in all instances)

timepoint ‘quadrat’ approach. Wherein, within each awCF time series, values of w and b were calculated respectively from STRs and lung function–time relationships from the first 5 timepoints and then calculated again after moving the temporal quadrat sequentially to the second time point, then the third, and so on across the remaining time series (Fig. 5). A benefit of this more granular approach is it allowed changes across BETR disease states to be more readily determined. For brevity, we focused this analysis on turnover within the whole microbiota only. For patients who experienced a

PEX, the general observed trend of microbiota turnover, albeit with substantial interpatient variation, was one of increasing values of w in pre-PEX baseline (B_0 , when data was available), then increasing further across exacerbation (E) and reaching a peak within treatment (T). After peaking, turnover then notably started to decrease within the treatment period, and then continued to decrease within the recovery (R), and post-PEX baseline (B_1) disease states (Fig. 5). In some of the PEX patients (e.g. P4, P6, P8, P10, and P11) turnover was then observed to start increasing again within the B_1 period. Within the

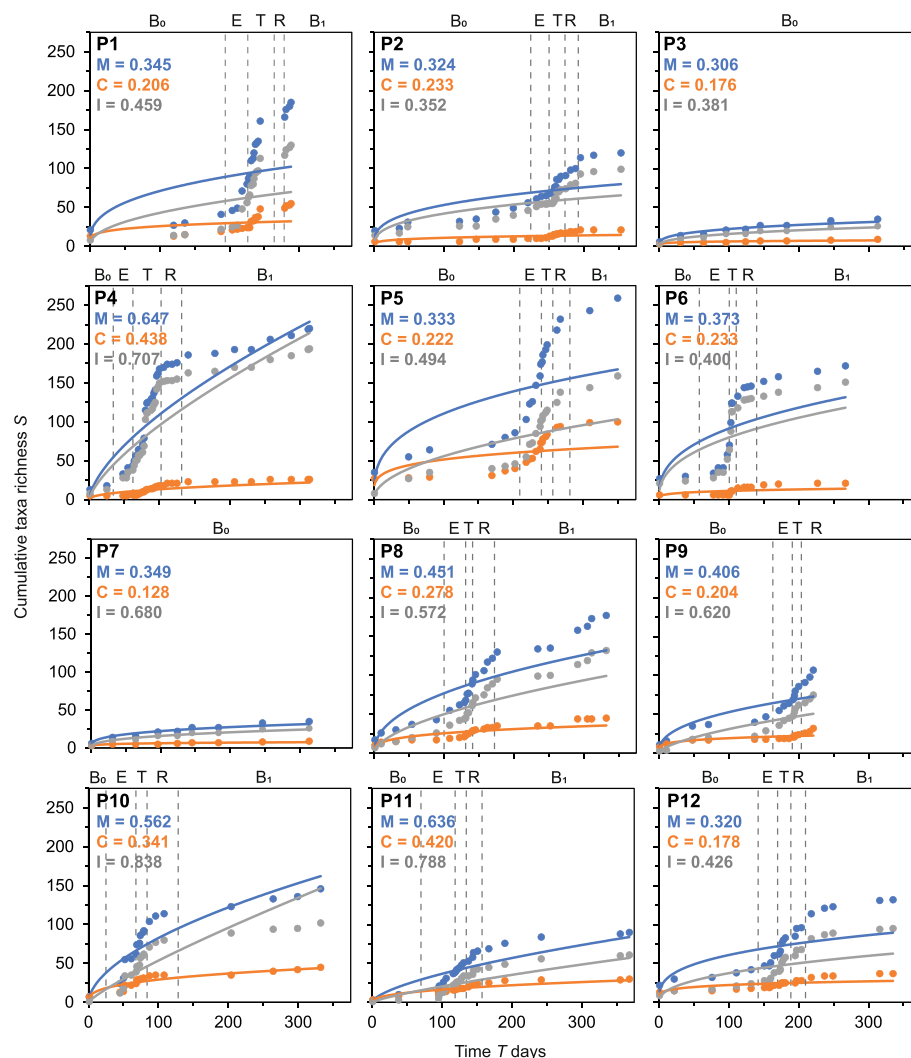


Fig. 2 Species-time relationships within patient lung microbiota. Disease states have been superimposed for each patient: B₀, baseline pre-exacerbation; E, exacerbation; T, treatment period; R, recovery period; and B₁, post-exacerbation baseline. Given are species-time relationships (STRs) for microbiota (blue), and the chronically (orange) and intermittently (grey) colonising taxa in each patient (P1 to P12). Also given in each instance is the STR slope value w for the microbiota (M), and the chronically (C) and intermittently (I) colonising taxa. All STRs were significant ($P < 0.05$). Full STR regression statistics are reported in Table S1

non-PEx patients (P3 and P7) an increase in turnover was observed across time in B₀. The rates of change in lung function (b) also broadly followed the observed general trend in microbiota turnover, usually with some lag and again with substantial interpatient variation (Fig. 5).

Discussion

Acute pulmonary exacerbations (PEx) are clinically significant events that are associated with increased morbidity and earlier mortality for pwCF [8, 36]. Furthermore, PEx can account for approximately half of the loss in lung function across a pwCF's lifetime [36]. Consequently, early detection and timely intervention is crucial [1, 2, 7].

However, PEx treatment is typically only initiated shortly after a pwCF presents with worsening respiratory symptoms [9, 17]. Hence, there is a pressing need to identify predictive biomarkers of PEx onset [2, 8, 36]. Here, we investigated temporal turnover to connect longitudinal changes within the lung microbiota of individual awCF to changes in disease states across a PEx event. Despite the highly personalized nature of CF lung microbiota, we found proof of concept that changes in temporal turnover could be indicative of the onset of PEx.

In agreement with our previous work, that first devised a modification of the Leeds criteria to define chronic infection status for all members of the microbiota within

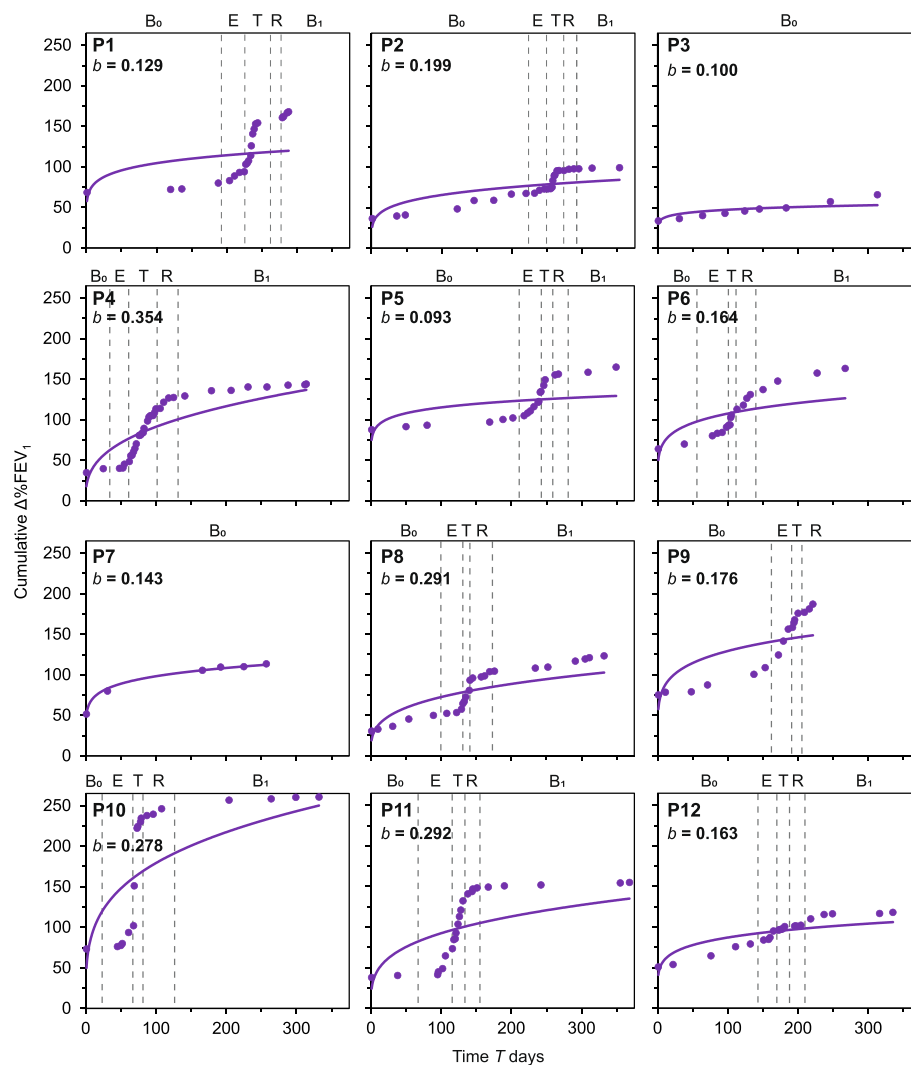


Fig. 3 Cumulative changes in patient lung function with time. Given in each instance is the cumulative absolute change in $\%FEV_1$ between consecutive timepoints. Each plot has been fitted with a power regression; $\Delta\%FEV_1 = at^b$. Where $\Delta\%FEV_1$ is cumulative change in lung function, a is the intercept, T is time in days, and b = slope/rate of change in lung function over time. Disease states have been superimposed for each patient: B_0 , baseline pre-exacerbation; E , exacerbation; T , treatment period; R , recovery period; and B_1 post-exacerbation baseline. Lung function-time relationship regression statistics: (P1) $R^2 = 0.25$, $F_{1,19} = 6.46$, $P = 0.02$; (P2) $R^2 = 0.61$, $F_{1,23} = 35.60$, $P < 0.0001$; (P3) $R^2 = 0.67$, $F_{1,7} = 14.19$, $P = 0.0007$; (P4) $R^2 = 0.60$, $F_{1,33} = 49.22$, $P < 0.0001$; (P5) $R^2 = 0.34$, $F_{1,18} = 9.10$, $P = 0.008$; (P6) $R^2 = 0.51$, $F_{1,16} = 16.87$, $P = 0.001$; (P7) $R^2 = 0.99$, $F_{1,4} = 920.53$, $P < 0.0001$; (P8) $R^2 = 0.70$, $F_{1,22} = 50.77$, $P < 0.0001$; (P9) $R^2 = 0.58$, $F_{1,14} = 19.25$, $P = 0.001$; (P10) $R^2 = 0.44$, $F_{1,16} = 12.66$, $P = 0.003$; (P11) $R^2 = 0.42$, $F_{1,21} = 15.38$, $P = 0.001$; (P12) $R^2 = 0.72$, $F_{1,20} = 51.99$, $P < 0.0001$. All relationships were significant ($P < 0.05$ in all instances). Changes in patient lung function with time are presented in Fig. S1

individual pwCF, we found that individual patients can be chronically colonized with not just a single pathogen but with multiple bacterial species, including multiple canonical CF pathogens (Fig. 1) [17]. In contrast, culture-based microbiological data for the study patients indicated that each patient was chronically colonized only with *Pseudomonas aeruginosa*. Culture offers a limited surveillance assessment that is constrained to detecting presence or absence of a handful of targeted bacteria within a respiratory sample [12]. Thus, not only does it

provide a blinkered view of the microbial species present, but it is also now recognized that culture can significantly underestimate presence of readily cultivable pathogens, including *P. aeruginosa* and *S. aureus* [12, 17]. Combined this has implications for choosing optimal antimicrobial interventions in general as well as to combat PEx [17, 30]. This adds further weight to recommending use of molecular approaches for microbiological surveillance in CF that define all microbial species present within an individual's lung infection microbiota through time [12, 17].

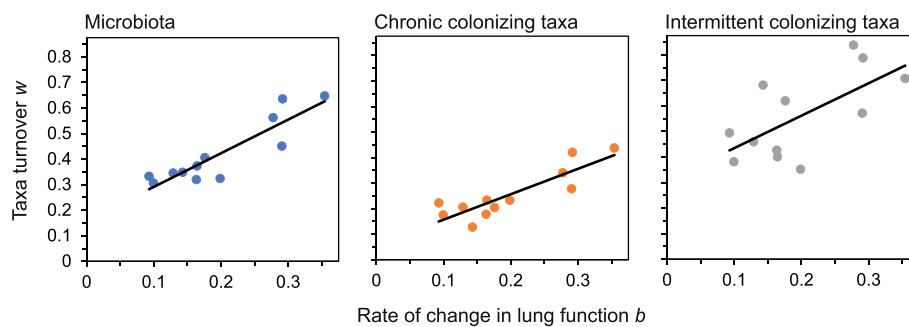


Fig. 4 Relationship between rate of change in lung function and taxa turnover. This relationship is plotted for the whole microbiota and the chronic and intermittent colonizing taxa groups. Taxa turnover w is derived from species-time relationships (Fig. 2). Rate of change in lung function b is derived from lung function-time relationships (Fig. 3). Regression statistics: microbiota $R^2 = 0.80$, $F_{1,10} = 40.08$, $P < 0.0001$; chronic taxa $R^2 = 0.76$, $F_{1,10} = 31.15$, $P < 0.0001$; and Intermittent taxa $R^2 = 0.43$, $F_{1,10} = 7.45$, $P = 0.022$. All relationships were significant at $P < 0.05$ level

For temporal turnover, high interpatient variability was evident in both the STR w -values and within the shapes of individual STRs for the whole microbiota and the chronic and intermittent colonizing groups (Fig. 2). This indicated that turnover and hence temporal organization are a personalized feature of the lung microbiota in CF, adding further support to the premise that CF lung microbiota through time and space are highly personalized to the individual pwCF [17, 19, 20, 37]. An expectation of STRs where no system perturbation occurs would be relatively low w -values and a steady cumulation of species richness through time [38]. This was observed for the STRs for the awCF that did not experience a PEx (Fig. 2). Conversely, those that did experience a PEx all demonstrated typically higher w -values in comparison along with notable increases in species richness cumulation through both the exacerbation and treatment periods (Fig. 2). Interestingly, the lung function-time relationships also exhibited similar patterns of increases in cumulative $\Delta\%FEV_1$ and hence that was also highly personalized to the individual pwCF (Fig. 3). Moreover, significant positive relationships were observed between values for rates of change in lung function (b) and turnover (w) within the microbiota, and chronic and intermittent colonizing taxa. This revealed that greater turnover was associated with greater change in lung function with time (Fig. 4). An explanation for the personalized nature of these observations could stem from that CF PEx events are likely not all the same between and even within pwCF [39]. Wherein, PEx are now thought to be caused by intricate and still not fully

understood interactions between the patient and bacteria [1], fungi [40], respiratory viruses [41], and air pollution [42] either singularly or in combination, further compounding the heterogeneous nature of PEx.

When microbiota turnover was examined at a finer scale across the time series for each awCF, values of w could clearly be observed to increase in the exacerbation period and often from within the pre-PEx baseline period and peaking within the treatment period (Fig. 5). This demonstrated that increases in turnover were not solely the result of the start of treatment and subsequent PEx antibiotic administration causing perturbations to the lung microbiome. Consequently, we posit that the increase in turnover ahead of treatment could be indicative of the onset of a PEx. Meta-analyses of STRs across different lengths of time and covering communities of animals, plants, and microbes across a wide variety of habitats and ecosystems have found STR derived rates of turnover (w) to have a remarkable degree of regularity that typically do not exceed more than 0.5 [43, 44]. Here, we found w -values soon increased beyond that, with a mean peak w ($\pm$ SD) across PEx patients of 22.74 ± 16.28 (minimum $w = 6.53$, maximum $w = 59.21$) (Fig. 5). As such, it was possible to use the departure from normal ranges of w up to the initiation of treatment to approximate onset of PEx in days across the awCF in the study. From w exceeding the typical maximum of $w = 0.5$ to start of treatment initiation the mean ($\pm$ SD) was 35.8 ± 20.1 days across PEx patients. When we take a more parsimonious approach and conservatively increase the

(See figure on next page.)

Fig. 5 Finer scale examination of temporal turnover (w) and changes in rate of lung function (b) within individual patients. To derive finer scale measures of w (columns) and b (circles), species-time relationships and cumulative changes in lung function were respectively plotted using windows of five sample time points, shifting by one time point for each new plot along the time series for each patient (P1 to P12). Values of w and b values are plotted at the middle time point of each subsequent five time point window. Disease states have been superimposed for each patient: B₀, baseline pre-exacerbation; E, exacerbation; T, treatment period; R, recovery period; and B₁ post-exacerbation baseline

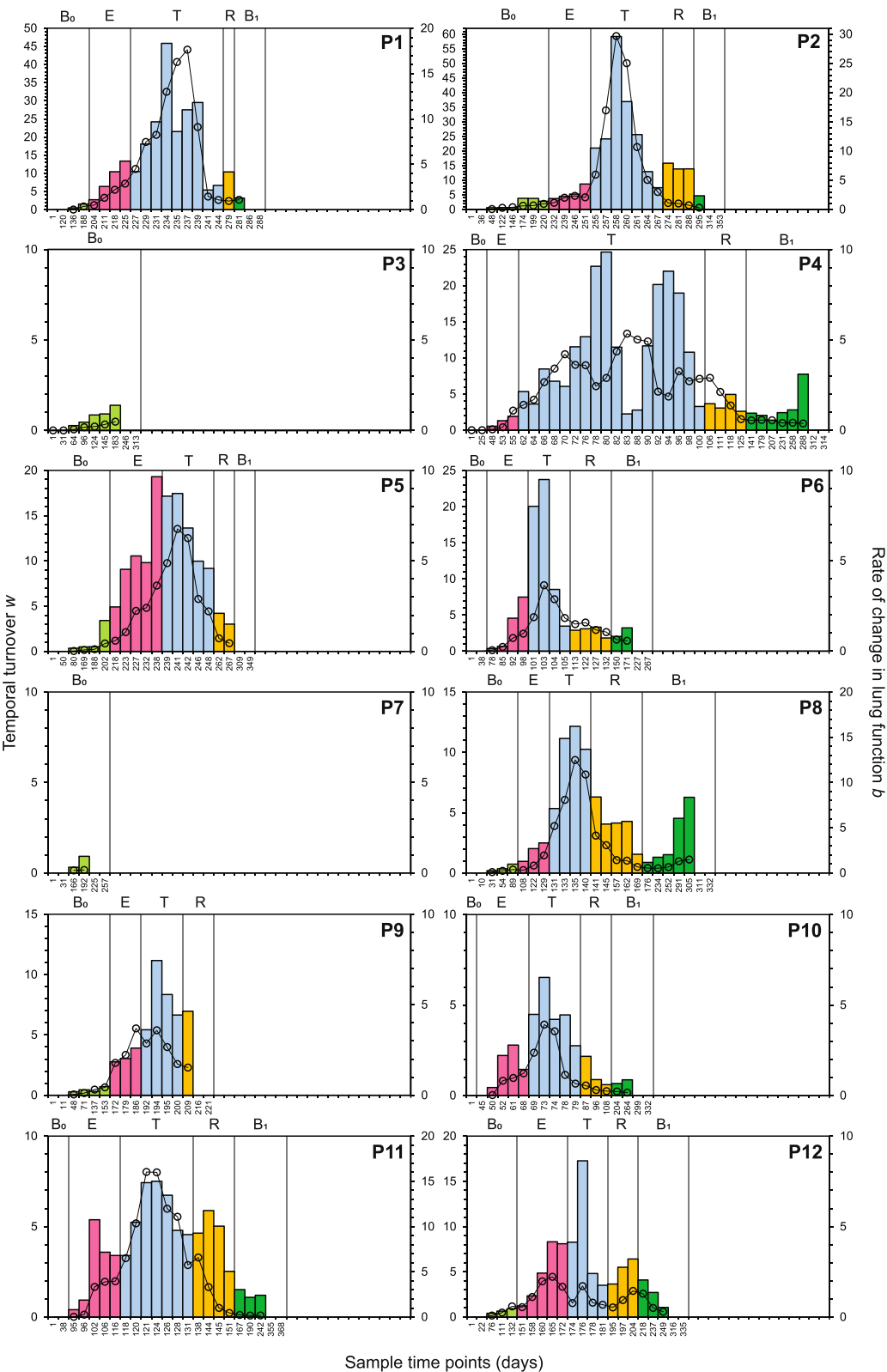


Fig. 5 (See legend on previous page.)

threshold to $w > 1.0$, to account for any negligible departure from the expected norm, the onset of exacerbation to start of treatment becomes 21.2 ± 8.9 days. Additionally, retrospective checking of clinical records, revealed for the PEx patients which had observed increases in turnover within the post-PEx baseline period (e.g. P4, P6, P8, P10, and P11) and the two non-PEx patients who both experienced increases in turnover across baseline (B_0), all went on to experience a PEx and intervention for PEx within weeks after the end of the study sampling. Regardless of which cutoff of turnover ($w > 0.5$ or $w > 1.0$) is used to indicate the onset of PEx, it is fair to state that both provide a considerable lead in before the start of antibiotic treatment. Therefore, our findings offer an exciting proof of concept which uniquely establishes that changes in microbiota turnover could potentially be used to indicate onset of an approaching PEx. Combined with incorporation of molecular approaches that define all microbial species present into microbiological surveillance of respiratory secretions throughout the life of a pwCF, this could enable an opportunity for pre-emptive intervention, which in turn could reduce the impacts of a PEx both in the short- and cumulatively in the long-term that result from delayed antibiotic interventions.

There are potential limitations to this study that deserve consideration. While our work offers an exciting prospect to identify onset of PEx, to further advance this work future clinical studies will need to have larger patient numbers representative of the wider CF population, including adults and children with CF. It is the case that more granular and longer longitudinal microbiota studies in CF are accompanied by an inevitable reduction in the amount of participating pwCF, as was the case here [17, 19, 20, 37]. While these ecology-based studies are providing previously unrealised and hard-won insight into the temporal dynamics of lung infection microbiota, planning of future clinical studies will need to carefully consider how to maximise patient participation and retention. Samples underpinning the data analysed in this retrospective study were collected before widespread availability of effective CF transmembrane conductance regulator (CFTR) modulators (e.g. elexacaftor/tezacaftor/ivacaftor [ETI]). How the temporal dynamics of lung infection microbiota will be affected within people on CFTR modulators is not well understood [17, 37]. Moreover, although ETI therapy has been shown to reduce the frequency of PEx, they still occur and are clinically impactful [45, 46]. Further, due to significant improvements in lung function experienced by pwCF on ETI therapy [46], alternatives to $\%FEV_1$ as a clinical indicator of health in CF are required. Additionally, in countries such as the

UK and USA, approximately 10% of the CF population either do not have eligible CFTR mutations or do not tolerate CFTR modulator therapy [47]. Likewise, there are many countries where there is limited or no access to CFTR modulator therapies [47]. Therefore, for those pwCF who are ineligible, intolerant, or lack access to modulators our findings remain directly relevant.

Conclusions

Setting microbiome research within a theoretical ecology framework is a parsimonious and pragmatic solution to understanding and predicting the complex ecology of respiratory microbiota related to clinical outcomes [25, 48]. The application of ecological theory to microbiome research is currently very limited [17, 18]. However, the power of these approaches are indeed becoming increasingly apparent in respiratory microbiome research, from using ecological modelling to define distinct PEx microbiota types and predicting treatment outcomes for PEx [20], to here where we illustrate that changes in temporal turnover are indicative of PEx onset in considerable advance of when antimicrobial treatment would normally be initiated. These theoretical based advances are all underpinned by considering temporal microbiome dynamics to connect changes in human microbiota to change in health state [17, 20, 37]. This goes beyond what traditional ‘one microbe, one disease’ approaches alone are capable of when combatting polymicrobial infections in chronic lung diseases [17]. Application of our work offers novel translational potential to enable early detection of PEx in CF and hence could allow consequent timely intervention and improve PEx management and even prevention. Moving on from basic science research, future clinical studies with greater patient numbers will be needed to realise this exciting potential. There is also real possibility to apply the temporal framework used here to PEx events across a range of chronic airway diseases.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40168-025-02143-5>.

Additional file 1: Supplementary Table S1. Species-time relationship (STR) regression statistics. Given are slope w and intercept c from the STR power function. Also given in each instance are degrees of freedom (df), F -statistic, coefficient of determination (R^2), and significance (P).

Additional file 2: Supplementary Figure S1. Changes in patient lung function with time. Given in each instance is the $\%FEV_1$ for consecutive timepoints. Disease states have been superimposed for each patient: B_0 , baseline pre-exacerbation; E, exacerbation; T, treatment period; R, recovery period; and B_1 post-exacerbation baseline.

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Authors' contribution

CvdG conceived the study. LC, DWR, and CvdG performed data and statistical analysis. CvdG, LC, and DWR verified the underlying data. CvdG, LC, and DWR were responsible for the original draft of the manuscript. All authors contributed to the development of the final manuscript. CvdG is the guarantor of this work. All authors read and approved the final manuscript.

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Data availability

The raw sequence data reported in this study have been deposited in the European Nucleotide Archive under accession numbers PRJEB581 and PRJEB7346. Anonymised clinical metadata, processed microbiota data have been deposited at figshare.com under <https://figshare.com/s/f34060344dba52278861>.

Declarations

Ethics approval and consent to participate

The study was reviewed and approved by the Southampton and Southwest Hampshire Research Ethics Committee, UK (06/Q1704/24). All patients provided written informed consent.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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