

Please cite the Published Version

Hawley, John ^(D), Forster, Samuel C. and Giles, Edward M. (2025) Exercise, Gut Microbiome, and Gastrointestinal Diseases: Therapeutic Impact and Molecular Mechanisms. Gastroenterology. ISSN 0016-5085

DOI: https://doi.org/10.1053/j.gastro.2025.01.224

Publisher: Elsevier

Version: Accepted Version

Downloaded from: https://e-space.mmu.ac.uk/638497/

Usage rights: (cc) BY

Creative Commons: Attribution 4.0

Additional Information: This is an open access article published in Gastroenterology, by Elsevier.

Enquiries:

If you have questions about this document, contact openresearch@mmu.ac.uk. Please include the URL of the record in e-space. If you believe that your, or a third party's rights have been compromised through this document please see our Take Down policy (available from https://www.mmu.ac.uk/library/using-the-library/policies-and-guidelines)

110

111

112

113

114

115

116

117

118

119

120

^{a1} ^{a2} Exercise, Gut Microbiome, and Gastrointestinal Diseases:
 ^{a2} Therapeutic Impact and Molecular Mechanisms

John A. Hawley,^{1,2} Samuel C. Forster,^{3,4} and Edward M. Giles^{3,5}

1

6

7

8

9

10

11

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

¹The Mary MacKillop Institute for Health Research, Australian Catholic University, Melbourne, Victoria, Australia; ²Department of Sport and Exercise Sciences, Manchester Metropolitan University Institute of Sport, Manchester, United Kingdom; ³Centre for Innate Immunity and Infectious Diseases, Hudson Institute of Medical Research, Clayton, Victoria, Australia; ⁴Department of Molecular and Translational Sciences, Monash University, Clayton, Victoria, Australia; and ⁵Department of Paediatrics, Monash University, Clayton, Victoria, Australia

The benefits of regular physical activity (PA) on disease prevention and treatment outcomes have been recognized for centuries. However, only recently has interorgan communication triggered by the release of "myokines" from contracting skeletal muscles emerged as a putative mechanism by which exercise confers protection against numerous disease states. Cross-talk between active skeletal muscles and the gut microbiota reveal how regular PA boosts host immunity, facilitates a more diverse gut microbiome and functional metabolome, and plays a positive role in energy homeostasis and metabolic regulation. In contrast, and despite the large interindividual variation in the human gut microbiome, reduced microbial diversity has been implicated in several diseases of the gastrointestinal (GI) tract, systemic immune diseases, and cancers. Although prolonged, intense, weight-bearing exercise conducted in extreme conditions can increase intestinal permeability, compromising gut-barrier function and resulting in both upper and lower GI symptoms, these are transient and benign. Accordingly, the gut microbiome has become an attractive target for modulating many of the positive effects of regular PA on GI health and disease, although the precise dose of exercise required to induce favourable changes in the microbiome and enhance host immunity is currently unknown. Future efforts should concentrate on gaining a deeper understanding of the factors involved in exercise-gut interactions through the generation of functional 'omics readouts (ie, metatranscriptomics, metaproteomics, and metabolomics) that have the potential to identify functional traits of the microbiome that are linked to host health and disease states, and validating these interactions in experimental and preclinical systems. A greater understanding of how PA interacts with the GI tract and the microbiome may enable targeted therapeutic strategies to be developed for individuals and populations at risk for a variety of GI diseases.

Keywords: AMPK; Cross-Talk; Fecal Microbiota Transfer; Gastrointestinal Tract; Immunity; Inflammation; Ischemia; mTOR; Myokines; Physical Activity; Probiotics; Skeletal Muscle.

E pidemiologic and cross-sectional data provide overwhelming evidence to demonstrate that lifelong physical activity (PA, defined as voluntary bodily movement produced by skeletal muscles that requires energy expenditure) postpones the onset of numerous chronic metabolic

conditions and noncommunicable diseases (NCDs) and is associated with an enhanced quality of life and extended health span.¹⁻⁶ In this regard, there has been belated recognition that "exercise is medicine" with the advancement and implementation of evidence-based approaches to elevate the status of PA in primary healthcare settings.⁷⁻¹¹ Many interdependent mechanisms underpin the healthpromoting effects of PA although the precise molecular bases by which lifelong PA (ie, exercise training) promotes human health and reduces disease risk remain poorly understood.¹² Although the biochemical and metabolic adaptations to exercise training have been extensively investigated in skeletal muscle, heart, adipose tissue, and the vasculature,¹³ only recently has there been appreciation that the gut microbiome plays a fundamental role in promoting some of the beneficial effects of regular PA on both general health, athletic performance, and anticancer immunity.¹⁴⁻¹⁶ Murine models using a variety of exercise interventions reveal consistent changes in the gut microbiome associated with improved health outcomes and longevity.¹⁷ Despite less than 5% overlap between human and mouse microbiota composition¹⁸ and several interspecies differences.¹⁹ such investigations provide opportunities for examining the mechanistic interactions between exercise and the gut microbiota that may be of direct translational value for discovering the functional processes underlying several human gastrointestinal (GI) disease states.^{20,21}

The gut microbiome constantly interacts with the host immune system, producing a range of metabolites that exert both local (cell/tissue/organ) and systemic (whole-body) effects.²² These interactions can trigger health or disease states by generating either harmful metabolites that

Abbreviations used in this paper: AMPK, AMP-activated protein kinase; BMI, body mass index; CI, confidence interval; CRC, colorectal cancer; FMT, fecal microbiota transplantation; GI, gastrointestinal; IBD, inflammatory bowel disease; IBS, irritable bowel syndrome; IL, interleukin; MASLD, metabolic dysfunction-associated steatotic liver disease; MET, metabolic equivalent; mTOR, Akt/mammalian target of rapamycin; NK, natural killer; NCD, noncommunicable disease; PA, physical activity; RR, relative risk; SCFA, short-chain fatty acids; TNF- α , tumor necrosis factor alpha; VO₂max, maximal oxygen consumption.

© 2025 The Author(s). Published by Elsevier Inc. on behalf of the AGA Institute. This is an open access article under the CC BY license (http:// creativecommons.org/licenses/by/4.0/). 0016-5085

https://doi.org/10.1053/j.gastro.2025.01.224

provoke disorders (eg, inflammatory bowel disease [IBD]) 121 or favorable metabolites (ie, short-chain fatty acids [SCFA]) 122 that are health-promoting. For this reason, the role of the 123 gut microbiome on its host has been the focus of intense 124 research, with the gut microbiota now established as a 125 powerful modulator of the efficacy of several treatments of 126 GI diseases. There has been an emergence of clinical in-127 terventions targeting the microbiome to enhance health 128 outcomes, particularly cancer,²³ from diagnosis and recur-129 rence to prognosis as well as treatment effectiveness.²⁴ Here 130 we discuss the role of PA and skeletal muscle metabolic 131 cross-talk with other tissues and organs, especially the GI 132 tract, with a focus on how exercise can modulate microbial 133 diversity. We describe the effects of exercise on the GI tract 134 concentrating on the role of bacteria and other micro-135 organisms in this process. Finally, the role of exercise and 136 the gut microbiome in the prevention of GI diseases will be 137 discussed in anticipation of emerging treatments that target 138 the microbiome to modulate these effects by understanding 139 the complex and interdependent interactions between PA, 140 diet, and GI function. 141

Metabolic Communication During Exercise

Skeletal Muscle Cross-Talk

142

143

144

145

146

147

179

180

The health-promoting effects of regular PA have tradi-148 tionally been attributed to exercise-induced increases in 149 whole-body cardiorespiratory fitness (VO_{2max}) evoked by 150 extensive remodelling of the vascular system, especially the 151 peripheral skeletal muscles subjected to exercise.¹³ How-152 ever, during the past 20 years, interorgan communication or 153 "cross-talk" initiated by contracting skeletal muscles has 154 emerged as a complementary mechanism through which PA 155 confers protection against an array of disease states.²⁵ 156 Although the hypothesis of a "general, humoral effect of 157 exercise" was proposed more than 60 years ago,²⁶ the 158 concept of skeletal muscle as an endocrine organ gained 159 credibility just 2 decades ago when it was shown that the 160 cytokine interleukin-6 (IL6) was released from skeletal 161 muscle during exercise.²⁷ Since that time, cytokines and Q3 162 other peptides, which are released by contracting skeletal 163 muscle fibers and exert autocrine, paracrine, or endocrine 164 effects, have been classified as "myokines."28,29 The list of 165 bona fide myokines includes IL6, IL8, IL15, decorin, 166 follistatin-like 1, fibroblast growth factor-21, irisin, chemo-167 kine CXC motif ligand-1 also known as keratinocyte-derived 168 chemokine, mitochondrially encoded peptide-c and 169 meteorin-like, although there are likely many more proteins 170 secreted by contracting skeletal muscles.^{25,29} Established 171 roles for "adipokines" and "hepatokines" as exercise-172 stimulated signals that coordinate systemic metabolism in 173 response to PA and inactivity have been reviewed previ-174 ously.^{25,30,31} The discovery of interorgan cross-talk offers a 175 framework for understanding how PA transmits many of its 176 beneficial effects on whole-body metabolic health (Figure 1). 177 178

Recently, important links between skeletal muscle and the gut microbiota have been uncovered, revealing how

Gastroenterology Vol. ■, Iss. ■

exercise facilitates a more diverse gut microbiome and functional metabolome, casting new light on the interconnectivity between these 2 organs in health and disease states.³² Exercise training alters both the bacterial community structure and numerous taxa that are associated with improved host health.³³ However, these exercise-induced changes are not universal with only moderate to highintensity exercise undertaken more than 3 times per week for longer than 8 weeks being consistently associated with either alterations in bacterial community and/or community structure.³⁴ In that analysis, the health status of the cohort under investigation created variability and potential confounders in results,³⁴ with a rather liberal definition of "healthy" that included individuals with no defined exclusion criteria in terms of chronic illness.

Contemporary investigations show that the gut microbiota exerts multiple effects on skeletal muscle bioenergetics³⁵ and have revealed how the gut bacteria respond to an exercise challenge with reciprocal roles in fuel availability, muscle function, and endurance capacity.^{16,36} The notion of cross-talk between the gut microbiota and contracting skeletal muscle emerged from studies in rodents showing increases in several species of SCFAs (acetate, butyrate, propionate, and conjugated linoleic acid) after endurance training.^{37–39} Although early studies demonstrated that SCFAs were absorbed in both the small and large intestine by similar mechanisms, it is now accepted that there exist species differences in SCFA producers as well as different transporter isoforms expressed in enterocytes along the intestine.⁴⁰ SCFAs have a protective effect on the host by reducing inflammation through transcriptional inhibition of cytokines and inflammatory proteins. Among the SCFAs, butyrate has received considerable attention for its beneficial effects on both cellular energy metabolism and intestinal homeostasis: butyrate is the primary fuel for colonocytes, increasing colonic epithelial cell proliferation, promoting gut barrier integrity, and regulating host gene expression and immunity.41

Compelling evidence for the existence of a "muscle-gut axis" came from the study of Lahiri et al⁴² who compared the skeletal muscle of germ-free mice that lacked a gut microbiota to the skeletal muscle of pathogen-free mice with an intact functional gut microbiota. In contrast to pathogenfree mice, skeletal muscle from the germ-free animals displayed significant atrophy and reduced muscle strength, underpinned by a decreased expression of insulin-like growth factor-1 along with reduced transcription of genes associated with skeletal muscle growth and mitochondrial function. Treating germ-free mice with a cocktail of SCFAs resulted in a reduced expression of Atroain-1 and an increased expression of myoblast determination protein-1, and partially reversed the functional impairments to muscle. These data support a role for the gut microbiota in regulating skeletal muscle mass and function, although whether this is a direct effect or immune-mediated remains to be determined. The influence of the microbiota-derived SCFA on hosts includes proliferation, differentiation, and aspects of metabolism, with most of these functions acting via gene expression, with butyrate modulating the expression of >20% of genes in humans.⁴³

181

182

183

184

185

186

187

188

189

190

191

ARTICLE IN PRESS

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

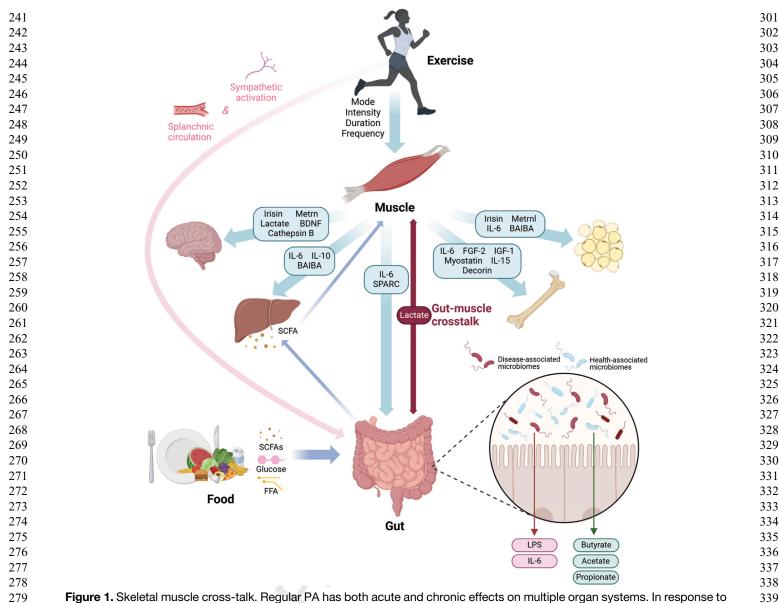
356

357

358

359

360



contractile stimuli, skeletal muscles secrete a range of molecules that "communicate" with other tissues and organs including adipose tissue, liver, brain, gut, bone, and the GI tract. While the majority of myokines are released from skeletal muscle in response to an acute bout of exercise, adipokines and hepatokines exert their biological function in response to repeated exercise stimuli through changes in whole-body metabolism. The release of cytokines and other mediators, alongside neurologic and vascular changes induced by exercise, modulate the intestinal barrier and gut function. Such changes are highly individual and are modified by the prevailing exercise challenge (mode, intensity, duration, and environmental condi-4 tions), an individual's habitual diet, and the microbiome. Exercise training is associated with increased microbial diversity and veb abundance of bacterial species, increasing SCFAs, whereas reduced microbial diversity in human intestines has been implicated in several GI diseases and CRC, with the microbiome exacerbating inflammation or other disease processes with øð immunomodulatory functions. BAIBA, β-aminoisobutyric acid; BDNF, brain-derived neurotrophic factor; FFA, free fatty acids; orint FGF-2, fibroblast growth factor-2; IGF-1, insulin-like growth factor 1; LPS, lipopolysaccardides; Metrnl, meteorin-like; SPARC, secreted protein acidic rich in cysteine. Created in BioRender. Belhaj M. (2025) https://BioRender.com/s64m052.

The first evidence demonstrating that exercise training in humans was associated with increased microbial diversity and abundance of bacterial species came from crosssectional studies on well-trained athletic populations. Clarke et al⁴⁴ reported that professional rugby players had higher alpha diversity and a greater relative abundance of the health-associated genus *Akkermansia* compared with highand low-body mass index (BMI) sedentary controls. Subsequent observational studies revealed that trained states were associated with a higher alpha diversity, an enrichment of beneficial taxa, and a higher abundance of fecal SCFAs.¹⁶ In these investigations, VO_{2max} accounted for up to a quarter of the variation in taxonomic richness after allowing for all other factors including diet,⁴⁵ with higher

299 300

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

Gastroenterology Vol. ■, Iss. ■

levels of fitness associated with a greater degree of both 361 alpha and beta diversity.44-47 These observations suggest 362 that a high level of cardiorespiratory fitness is related to 363 certain bacterial species and metabolite production, which 364 have been implicated in several diseases states including 365 reduced cancer risk.²³ Indeed, a recent study reported 366 greater gut microbiome diversity and differential abun-367 dances between colorectal cancer (CRC) survivors who were 368 physical active compared with those who were inactive,48 369 with active patients having enriched abundance of multi-370 ple dominant genera including Faecalibacterium and Blautia 371 and less dominant genera including Succiniclasticum and 372 Succinivibrio. At the phylum level, active patients had lower 373 Actinobacteria abundance. These findings are consistent 374 with studies showing higher abundances of Succinivibrio, 375 Faecalibacterium prausnitztii, Roseburia hominis, and 376 Akkermansia muciniphila in healthy individuals and ath-377 letes.^{44,45,47} Faecalibacterium and Succinivibrio have been 378 linked to health-promoting attributes including decreased 379 risk for CRC.49 380

Further evidence of a muscle-gut axis comes from the 381 work of Scheiman et al.⁵⁰ Using 16S rRNA profiling, these Q4 382 workers reported a higher abundance of Veillonella in 15 383 runners who completed the 2015 Boston marathon 384 compared with a group of 10 healthy but sedentary con-385 trols. Veillonella species metabolize lactate into the SCFAs 386 Q5 acetate and propionate via the methylmalonyl-CoA pathway 387 and, although many other microbes have the capacity to use 388 lactate, they do not possess the full pathway to convert 389 lactate to propionate. Using intracolonic infusion, a strain of 390 Veillonella atypica was isolated from stool samples of the 391 runners and when this strain was inoculated into mice, 392 treadmill running time to exhaustion was significantly 393 increased. These findings are intriguing as they highlight 394 that the microbiome may be a critical component of 395 endurance performance but raise the question of how this 396 performance-facilitating organism came to be more preva-397 lent among endurance-trained athletes in the first place. 398

In contrast, reduced microbial diversity has been impli-399 cated in many GI diseases^{51,52} although direct causation 400 remains to be determined. In severe liver disease and IBD, 401 antibiotics are frequently prescribed, leading to periods of 402 significantly reduced microbial diversity, potentially driving 403 ongoing disease pathogenesis. Such microbial effects are 404 also strongly associated with sarcopenia, defined as a loss of 405 skeletal muscle mass, which is often seen in parallel with 406 these diseases and is closely related to levels of PA.^{53,54} This 407 "dysbiosis" affects host metabolism and the functionality 408 and pathophysiology of several peripheral organs with ex-409 ercise a potential intervention to perturb gut microbiota 410 composition and re-establish gut symbiosis. There is a large 411 variation in the outcomes of exercise on the microbiome 412 across studies^{38,55,56} due to the complexity of the interac-413 tion with individual microbiome variation, different partic-414 ipant cohorts, the exercise dose (ie, the intensity, duration, 415 and frequency) and other variables (diet, cultural, and 416 geographical demographics).57 For example, the relative 417 abundance of Prevotella is associated with endurance-based 418 exercise programs,58 whereas in athletes who undertake 419 420

more intense power-based training, there is marked intestinal inflammation and greater abundance of bacteria involved in inflammatory processes such as Haemophilus, Rothia, Mucispirillum, and Ruminococcus gnavus.^{59,60} Although the optimal dose of exercise to induce favourable changes in the gut microbiota is unknown, it is likely to be specific to an individual's prevailing health status and background diet. Results from a recent investigation demonstrate that exercise training-induced alterations to gut microbiota composition and function were dependent on the body composition of participants. Allen et al⁶¹ reported that short-term endurance exercise training increased fecal concentrations of SCFAs in lean individuals $(BMI < 25 \text{ kg/m}^2)$ but not those with obesity (BMI > 30 25) kg/m^2), an observation that was independent of diet. Of Q_6 note, the exercise-induced changes to the microbiota, VO_{2max}, and body composition were rapidly lost in both cohorts when the training stimulus was terminated, suggesting that shifts in the metabolic capacity of the gut microbiota were strongly associated with the prevailing fitness levels, were transient, and were highly dependent on a regular exercise stimulus. Clearly, regular PA is necessary to produce favorable long-term modifications in the gut microbiota composition, largely through immune-metabolic pathways associated with anti-inflammatory effects.^{61–63} Further studies are required to understand the mechanisms that regulate changes in the composition and functions of the microbiome caused by PA along with their related effects.33

Diseases of the GI Tract and Effects of Exercise on GI Symptoms

Acute exercise has diverse effects on the GI tract (Figure 2) that can broadly be divided into upper (ie, heartburn, belching, regurgitation, and epigastric pain) or lower GI symptoms (ie, nausea, diarrhea, blood loss, flatulence, urgency, and abdominal pain). Exercise affects both motility and absorptive properties of the GI tract, with acute exercise slowing transit times in the upper GI tract due to increased sympathetic nervous activity.64 There is also a reduced absorptive capacity during exercise,65,66 with increased nutrient load to the caecum and colon driving symptoms in the lower GI tract.⁶⁷ The altered nutrient environment also interacts with the resident microbiome as a potential modifier of such symptoms. Upper GI symptoms are more common than lower GI symptoms with prolonged (>90 min) weight-bearing exercise (ie, running) associated with a greater number and severity of symptoms.^{68,69} GI problems are especially common in athletes training for and competing in endurance events with 30%-50% of athletes reporting 1 or more GI symptoms.⁷⁰ However, most GI symptoms are mild, Q7 transient, and of no risk to long-term health.

Ischemia and inflammation

Changes in splanchnic blood flow underpin many of the adverse clinical effects of exercise on GI function^{71,72} during which splanchnic ischemia leads to gut barrier 421

422

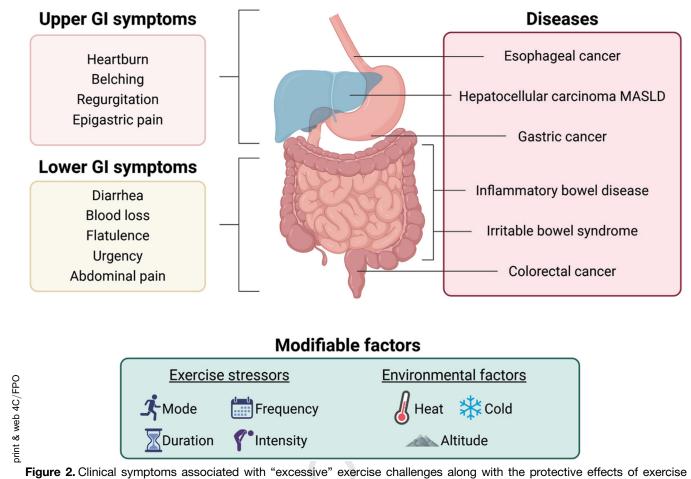


Figure 2. Clinical symptoms associated with "excessive" exercise challenges along with the protective effects of exercise training on GI diseases. Acute bouts of exercise can induce both upper and lower GI symptoms (*left*) depending on the prevailing exercise and environmental factors (*middle, lower*). Exercise training is associated with favorable outcomes for a number of GI diseases (*right*). Created in BioRender. Belhaj M. (2025) https://BioRender.com/s64m052.

Endotoxemia is suggestive of barrier dysfunction or

damage, with either bacteria or their products translocating

across the epithelial lining of the GI tract. Evidence of such

dysfunction during PA is confounded by the different modes

and intensities of exercise, the fitness level of subjects

studied, and the various measures of gut damage used in

investigations. This, in part, explains the wide range of

outcomes reported, although one should also consider that

the resident microbiome is another variable in exercise-

induced barrier stress. For example, a high protein diet

(1.5 g/kg body mass), commonly consumed by athletes

training for strength/power events, may contribute to dys-

amplify intestinal damage and symptoms. As exercise in-

tensity and duration increases, there is considerable evi-

dence for increases in indices of intestinal injury,

permeability, and endotoxemia, together with impairment of

gastric emptying, slowing of small intestinal transit, and

malabsorption.⁸⁶ The addition of heat stress and running

mode exacerbates markers of GI disturbance, with an ex-

ercise duration of ~ 2 hours at a moderate (60% of VO_{2max})

intensity seemingly the "threshold" at which significant GI

biosis (eg, increased *Escherichia coli* lipopolysaccharide) and Q8

disorders.⁷³⁻⁷⁵ Such exercise-induced ischemia leads to systemic endotoxemia and an associated inflammatory response.⁷⁶⁻⁷⁹ Historically intestinal damage has been measured after exercise through fecal blood loss⁸⁰ and more recently with intestinal-fatty acid binding protein^{75,81} and calprotectin.⁷⁵ In healthy individuals intestinal permeability is related to the prevailing exercise-intensity with intense exercise (>80% of VO_{2max}) resulting in greater increases in small intestinal permeability compared with exercise un-dertaken at 40%-60% of VO_{2max}. However, increases in permeability do not always result in a higher prevalence of GI symptoms and increased intestinal permeability.⁸² A study of endurance-trained athletes performing 1 hour of intense cycling at 80% of VO_{2max} reported little effect on barrier function or self-reported GI symptoms,⁸³ suggesting there is considerable variability in the effect of both the relative exercise intensity and mode on gut damage, with well-trained individuals experiencing fewer GI disturbances compared with exercise-naïve subjects. Indeed, the GI tract is highly adaptable: gastric emptying as well as stomach comfort can be "trained" and perceptions of fullness de-creases in athletes practicing a variety of nutritional stra-tegies before and during exercise.^{84,85}

perturbations occur.86

The Potential Role of the Gut Microbiome as a Mechanism to Reduce GI Symptoms During Exercise

604 A potential role for the microbiome in modulating the 605 effects of PA is through dietary interventions. Carbohydrate 606 ingestion immediately before or during exercise attenuates 607 GI symptoms and markers of intestinal damage⁸⁷⁻⁸⁹ and, 608 while this effect may be independent of the microbiome, 609 increased carbohydrate availability may temper the dys-610 biosis reported by many athletes.^{86,90} Other diets, including 611 a gluten-free diet, have been investigated in an attempt to 612 reduce GI symptoms, but appear ineffective.⁹¹ To support 613 the splanchnic circulation and reduce intestinal damage 614 during exercise, the nitric oxide precursors and amino acid 615 supplements L-arginine and L-citrulline supplements have 616 been investigated, although their efficacy is equivocal.^{92,93} 617 Although these dietary interventions are likely to have 618 moderate effects on the microbiome, it remains speculative 619 whether they would exert any effect on exercise capacity, or 620 the mechanism of such an effect.

621 Direct attempts to modulate the microbiome to preserve 622 permeability and attenuate other exercise-induced barrier 623 damage have been largely ineffective, including combina-624 tions comprising 6 (Bifidobacterium bifidum, Bifidobacte-625 rium lactis, Enterococcus faecium, Lactobacillus acidophilus, 626 Q9 L. brevis, and Lactococcus lactis) and 9 probiotic strains (L 627 acidophilus, Lactobacillus rhamnosus, Lactobacillus planta-628 rum, Lactobacillus fermentum, Lactobacillus casei, Bifido-629 bacterium breve, B lactis, B bifidum, and Streptococcus 630 thermophilus).94,95 In a recent double-blind placebo-631 controlled trial, probiotic supplementation (L acidophilus 632 and Bifidobacterium longum) administered for 5 weeks to 633 amateur runners reduced GI symptoms and was associated 634 with less dysbiosis.⁹⁶ In a double-blind, crossover study 16 635 runners were randomized to 4 weeks of daily supplemen-636 tation with a probiotic cocktail containing Pediococcus 637 acidilactici bacteria and L plantarum or placebo. Treadmill 638 running tests (90 minutes at 65%-70% of VO_{2max} or until 639 fatigue/GI symptoms developed, undertaken in 27°C) were 640 performed before and after supplementation. GI symptoms, 641 gut permeability-associated parameters (intestinal-fatty 642 acid binding protein, lipopolysaccharide-binding protein, 643 zonulin, and cytokines), and intestinal microbial content 644 were not altered by the probiotic supplementation.⁹⁷ L casei 645 supplementation for 7 days before 2 hours of running at 646 60% of VO_{2max} in extreme (34°C) heat did not alter resting 647 circulatory endotoxin concentration or plasma cytokine 648 profile compared with a placebo, although there was a trend 649 for higher plasma endotoxin and tumor necrosis factor 650 alpha (TNF- α) concentrations after exercise in the probiotic-651 supplemented group,98 suggesting that changes to the 652 microbiome may influence barrier function during PA. 653 although not always favorably. Although probiotic supple-654 mentation has traditionally focused on gut health, in recent 655 years, the clinical applications of probiotics have broadened 656 to allergic, metabolic, inflammatory, GI, and respiratory 657 conditions. Probiotic supplementation could yield small 658 beneficial effects in promoting health in trained individuals, 659

likely by reducing the risk of respiratory and GI illness during intensified periods of training and competition but precise compositions and dosage regimes remain to be determined. Such benefits would most likely be mediated by changes in gut microbiota and enhanced mucosal barrier integrity in the GI and respiratory tracts.⁹⁹

The Potential Role of Exercise-Microbiome Cross-Talk in GI Diseases

Exercise undertaken according to the American College of Sports Medicine guidelines¹⁰⁰ is associated with improved health outcomes in numerous NCDs, including many nonmalignant conditions of the GI tract and also several cancers. Perhaps the most obvious GI condition linked with regular PA is metabolic dysfunction-associated steatotic liver disease (MASLD), with exercise recommended as a primary treatment across many international guidelines.¹⁰¹ Although MASLD is intimately related to lack of PA, obesity, and a poor diet, not all sedentary people with obesity develop fatty infiltration of the liver. The gut microbiome is related to this complex interplay with a "gutliver axis" of diet, genetics, and inflammation being potential drivers of the fibrosis and risk of hepatocellular carcinoma. Treatment strategies for these conditions should consider combining regular exercise with microbiome modulation to impact MASLD¹⁰² because exercise can reverse gut "dysbiosis" in MASLD patients.¹⁰³

IBD has 2 major phenotypic forms: Crohn's disease and ulcerative colitis, with rising worldwide incidence. Although the precise etiology of IBD is unknown, several factors arising from adipose tissue and skeletal muscle have been implicated including cytokines, adipokines, and myokines.¹⁰⁴ In a high-fat-fed murine model of colitis, moderate-intensity exercise (voluntary wheel running) significantly decreased macroscopic and microscopic colitis, increased colonic blood flow, and attenuated plasma TNF- α , IL6, monocyte chemoattractant protein-1, IL1 β , and leptin levels. In contrast, in sedentary mice fed the same diet, colonic lesions were aggravated, colonic tissue weight increased, and the plasma TNF- α , IL6, monocyte chemoattractant protein-1, IL1 β and leptin levels significantly increased.¹⁰⁵ In children with IBD, intense exercise did not alter symptomatic outcomes or changes in serum inflammatory cytokines when compared with age-matched controls.¹⁰⁶ However, a recent systematic review of 637 patients (34% males) and pooled evidence from 6 randomized clinical trials found exercise improved disease activity but not disease-specific quality of life compared with controls.¹⁰⁷ Given that IBD is triggered by an abnormal immune response to the resident microbiota,¹⁰⁸ any effects of exercise on IBD likely impact the microbiome, although a systematic review of the systemic inflammatory response in exercise-based intervention studies in IBD patients reported no consistent effect.¹⁰⁹ A combination of exercise and psychological interventions in a group of patients with Crohn's disease led to positive changes in their intestinal microbiome with accompanying reductions in systemic markers of inflammation.¹¹⁰

717

718

719

720

781

782

783

784

785

786

787

788

789

790

791

792

793

794

795

796

797

798

799

800

801

802

803

804

805

806

807

808

809

810

811

812

813

814

815

816

817

818

819

820

821

822

823

824

825

826

827

828

829

830

831

832

833

834

835

836

837

838

839

840

A recent meta-analysis concluded there was a reduced 721 risk of developing IBD in individuals who undertook regular 722 high- vs low-intensity PA.¹¹¹ However, early results from 723 the IBD-FITT study (ClinicalTrials.gov NCT04816812), a 724 RCT of 12 weeks of aerobic exercise in people with IBD,¹¹² Q11 725 show improvements in quality of life and other health-726 related outcomes, without any effect on disease activity.¹¹³ 727 These contrasting results suggest strong individual varia-728 tion, with differences in the timing of the commencement of 729 exercise training in relation to disease activity likely to be 730 important. Fatigue, a major symptom in IBD patients, is 731 associated with muscle deconditioning, with a small study 732 showing patients with Crohn's disease exhibit impaired 733 Q12 postexercise recovery, measured based on slower rates of 734 muscle phosphocreatine resynthesis.¹¹⁴ Although the links 735 between PA, the microbiome, and disease outcomes in IBD 736 remain unclear, sarcopenia is associated with poor clinical 737 outcomes independent of IBD activity and, therefore, muscle 738 health should be assessed in all IBD patients at routine 739 intervals.54 740

In patients with irritable bowel syndrome (IBS), a dis-741 ease without an easily measurable inflammatory response, 742 symptoms are reduced after exercise training^{115,116} and 743 associated with exercise-induced improvements in gut 744 motility along with favorable psychological changes. The 745 first-line therapy for IBS is the low Fermentable Oligosac-746 charides, Disaccharides, Monosaccharides, and Polyols 747 (FODMAP) diet,¹¹⁷ which modulates its effect through direct 748 dietary impact on the microbiome. As such, the benefit of 749 exercise on IBS and other motility-related GI conditions may 750 be mediated, in part, through the microbiome. A recent trial 751 reporting fewer IBS symptoms after exercise training¹¹⁸ 752 Q13 also found that exercise reduced cell-free mitochondrial 753 DNA in the plasma of the control (non-IBS) group.^{118,119} 754 Because mitochondrial DNA is a damage-associated molec-755 ular pattern that is bidirectionally associated with dysbio-756 sis,¹²⁰ the results from that study suggest that PA can 757 modify cellular damage-associated molecular patterns in 758 healthy patients but not in IBS patients, which may be due 759 to different microbial ecologies in those with IBS. A recent 760 Cochrane review reported moderate positive outcomes of 761 regular PA on IBS symptoms without clear improvement in 762 quality of life,¹²¹ again suggesting strong individual varia-763 tion in such effects. Putative mechanisms that link the 764 microbiome to effects of PA on IBS patients include changes 765 to transit time, greater SCFA production, improvement in 766 the intestinal barrier, as well as modulation of the immune 767 response.122 768

Exercise and GI Cancer Prevention: Molecular Mechanisms Related to the Microbiome

The protective effect of regular, moderate to vigorous PA against cancer risk is well established with evidence from epidemiologic studies, randomized controlled trials, and meta-analyses from cohort investigations showing that regular exercise reduces the risk of between 7 and 13 different cancers and cancer mortality, including less frequent recurrence and fewer/less severe adverse effects.¹²³⁻¹²⁸ Regular PA is inversely related to the risk of cancer at the proximal (relative risk [RR], 0.76; 95% confidence intervals [CI], 0.70-0.83) and distal colon (RR, 0.77; 95% CI, 0.71-0.83), although there is limited evidence for any positive effects of PA on reducing risk of rectal cancers (RR, 0.98; 95% CI, 0.88-1.08), indicating that different mechanisms are operating in the development of colon and rectal cancer.¹²⁹ The RR for gastroesophageal cancer is 0.82 (CI, 0.74–0.90).¹³⁰ The cancer for which regular PA confers the greatest protective effect is CRC,¹³¹⁻¹³³ with moderate levels of PA associated with a 25% reduction of CRCs and greater amounts of PA linked to a greater (40%) reduction in risk.^{134,135} Even colorectal polyps can be reduced in number by increasing levels of PA.^{132,136} After a year-long exercise training intervention in previously sedentary men who completed a minimum of 250 min/week of moderateto vigorous-intensity exercise and whose VO_{2max} increased by >5%, there was a significant decrease in colon crypt proliferation and associated biomarkers (crypt height, number, and relative position of Ki67+ cells in stained cells in colon mucosal crypts) compared with nonexercising control subjects.¹³² In an analyses of 48 studies that included 40,674 colon cancer/CRC cases, a graded inverse Q14 dose-response association was observed between PA and colon cancer for both sexes. This dose-response effect of physical activity on colon cancer risk was especially strong when patients performed activities of >4.5 metabolic equivalents (MET) vs < 4.5 MET (where 1 MET is the resting metabolic rate equivalent to 3.5 mL $O_2/kg/min$).¹³⁵ Of note, adjusting for potential cofounding factors including age, diet, and obesity does not diminish the observed associations between levels of PA and colon cancer occurrence.¹³² Exercise training has also been shown to reduce the risk of gastric cancer with individuals who are more physically active having a 19% lower incidence.¹³⁷ A similar prognostic effect was seen for esophageal cancer.^{138,139} Given the range of similar risk reduction it appears likely that there is at least a degree of mechanistic crossover in the benefits in PA across GI cancers, with the mechanisms underpinning such effects being multifactorial. Here we focus Q15 on those mechanisms potentially related to musclemicrobiome cross-talk.

Over recent years, the use of immunotherapy in cancer treatment has highlighted the critical role of the immune system in cancer pathogenesis. The microbiome is intimately involved in the development of the immune system, especially in the GI tract.^{140,141} Exercise induces a biphasic response in lymphocytes and mobilizes vascular, pulmonary, hepatic, and splenic white blood cells into the peripheral circulation. This initial lymphocytosis affects mainly natural killer (NK) cells, which increase several-fold above resting levels, with their mobilization proportional to the relative intensity of exercise. This response underpins adrenaline-stimulation of β 2-adrenergic receptors on the surface of lymphocytes leading to endothelial detachment and the subsequent recirculation of lymphocytes into the bloodstream. Blood lymphocyte count decrease 1-2 hours after exercise cessation with transient lymphopenia not

769

770

771

772

773

774

775

776

777

Gastroenterology Vol. ■, Iss. ■

901

902

903

904

905

906

907

908

909

910

911

912

913

914

915

916

917

918

919

920

921

922

923

924

925

926

927

928

929

930

931

932

933

934

935

936

937

938

939

940

941

942

943

944

945

946

947

948

949

950

951

952

953

954

955

956

957

958

959

960

uncommon, although NK and T-cell counts return to baseline within 24 hours.¹⁴² The precise mechanisms by which
exercise-stimulated catecholamine signalling drive immune
cell mobilization, redistribution, and function are not
known, nor is the relationship by which this could be
modified by changes in the microbiome.¹⁴³

The immune-stimulating effects resulting from an acute 847 bout of exercise persist for several hours and the use of 848 exercise-stimulated T cells to augment the volume of cells 849 that could be used for adoptive transfer into lymphopenic 850 patients has been discussed elsewhere.¹⁵ However, it is the 851 chronic effects of the progressive accumulation of frequent 852 acute challenges that are likely to play regulatory roles in 853 tumor growth kinetics and tumor metabolism, through their 854 effects of immune function in general, and on antitumor 855 immune function specifically.¹⁵ In rodents, a program of 856 voluntary wheel running for 2 months was linked to 857 improved antitumorigenic function through an increase in 858 splenic NK cell cytoxic function, an effect that persisted 3 859 weeks after the training intervention ceased.¹⁴⁴ A similar 860 immune response has been reported in humans with Lynch 861 syndrome (a hereditary condition with a high lifetime risk of 862 CRCs and endometrial cancers), where chronic (12 months) 863 exercise training decreased inflammatory markers (prosta-864 glandin E) in colon and blood and increased the colonic 865 mucosa levels of NK and CD8+ T cells in the exercise-866 trained patients.¹⁴⁵ These findings have important implica-867 tions for cancer interventions in patients with Lynch syn-868 drome because they demonstrate the beneficial biological 869 effects of exercise in the immune system of a target organ in 870 patients at risk for cancer. Given the relationship between 871 mucosal immunity and the microbiome,¹⁴⁶ it is conceivable 872 that such an effect may be mediated through intestinal 873 microbiota. 874

While PA modulates pathways regulating local and sys-875 temic inflammation and oxidative stress, 126 exercise-876 induced activation/suppression of pathways that sense 877 and regulate energy availability, cellular metabolism, tumor 878 development, proliferation, metastasis, and cytoskeleton 879 organization are also important in conferring the protective 880 Q16 effect of PA. These pathways include the AMP-activated 881 protein kinase (AMPK) and the Akt/mammalian target of 882 rapamycin (mTOR) pathways. The targets of these signalling 883 nodes include many transcription factors, coactivators, and 884 repressors¹³ with all of these pathways potentially modified 885 by alternations in the gut microbiome. Indeed, a growing 886 body of evidence supports the beneficial effects of AMPK on 887 gut health, such as enhancing intestinal absorption, 888 improving barrier function, suppressing colorectal carcino-889 genesis, and reducing intestinal inflammation and 890 metabolic-related disease states. Conversely, AMPK is 891 inhibited under conditions of obesity and diabetes, both of 892 which are correlated with impaired intestinal barrier 893 function.147 894

The AMPK pathway is activated in tumors in response to an acute bout of PA, although exercise training may inhibit carcinogenesis by suppressing the activation of the mTOR signaling network in carcinomas, an effect mediated through effects of PA on circulating growth factors and hormones

that regulate the mTOR network that are distinct from those affecting mTOR activity in contracting skeletal muscle.¹⁴⁸ Although this premise has some mechanistic basis, no studies have fully elucidated the direct effects of mTOR signalling on tumor growth in humans.¹⁴⁹ Tumors have altered cellular metabolism that favors aerobic glycolysis to support the high energy turnover and rapid cell proliferation. Accordingly, intratumoral metabolism will be impacted by whole-body PA, with results from preclinical studies suggesting that tumors with intrinsically high metabolism are susceptible to exercise-induced energy depletion. Such shifts in metabolism induced by exercise modulate the metabolic reprogramming that occurs during carcinogenesis to support cell growth and proliferation, and suggest that carcinogenesis can be inhibited or enhanced by effects on intermediary metabolism linked to PA and dependent on exercise intensity and duration. A recent example in CRC patients provided mechanistic insights into PA impact on tumor growth. In large CRC datasets, RPS4X expression (related to "stemness"), which is associated with metastases and poor outcomes, is down-regulated with exercise.¹⁵⁰ Given the relationship between the microbiome and stem cell regulation in the intestine, it is feasible that exercise may influence CRC risk via this pathway. This makes CRC a potential early candidate for combining microbiome modulation with PA in improving clinical outcomes.

Another potential mechanism for the beneficial effects of regular exercise on CRC risk and tumor growth is the training-induced increase in SCFA production. CRC patients typically present with a compromised gut microbiota characterized by a reduced abundance of butyrate-producing taxa, including Roseburia and Lachnospiraceae. Results from in vitro studies demonstrate that butyrate differentially regulates gene expression in healthy vs cancerous cells.¹⁵¹ In healthy epithelial cells, butyrate is rapidly metabolized via the mitochondrial TCA cycle, causing in the accumula- Q17 tion of cytosolic citrate and acetyl CoA, thus increasing the acetylation of histones by histone acetyltransferases. This epigenetic modification increases expression of genes involved in cell proliferation and cell turnover, strengthening the intestinal barrier. However, in CRC cells, mitochondrial dysfunction results in the accumulation of butyrate in the cytosol. This free butyrate inhibits histone deacetylases, resulting in the epigenetic suppression of proliferation and promotion of cell death pathways that may lead to a reduction in tumor size and reduces the chance of metastasis.¹⁵¹

Human Microbiome Research: Current Perspectives and Directions for Future Research

The benefits of regular PA in preventing and/or treating numerous NCDs have been known for centuries. However, the precise mechanisms by which exercise training defends and protects the body against a range of lifestyle-induced diseases have not been elucidated. The discovery that contracting skeletal muscle is an endocrine organ acting as the primary metabolic communicator for interorgan

1021

1022

1023

1024

1025

1026

1027

1028

1029

1030

1031

1032

1033

1034

1035

1036

1037

1038

1039

1040

1041

1042

1043

1044

1045

1046

1047

1048

1049

1050

1051

1052

1053

1054

1055

1056

1057

1058

1059

1060

1061

1062

1063

1064

1065

1066

1067

1068

1069

1070

1071

1072

1073

1074

1075

1076

1077

1078

communication provides a plausible mechanism by which 961 exercise training boosts immunity at both the whole-body 962 and local (tissue) level. Although regular PA facilitates a 963 more diverse gut microbiome and functional metabolome 964 with direct and variable effects on GI disease outcomes, the 965 precise dose of exercise necessary to induce favorable 966 changes in the gut microbiome and enhance host immunity 967 is unknown. Although "excessive" PA can have detrimental 968 effects on intestinal barrier integrity, these are transient and 969 benign. The gut microbiome, therefore, is an attractive 970 target for modulating many of the positive effects of exercise 971 on GI health and disease with the links between certain GI 972 microbiota and the disease states of peripheral tissues/or-973 gans making microbial modulation a potentially potent 974 immunotherapeutic therapy. 975

Current evidence indicates that a certain level of horm-976 esis is necessary to perturb the human gut microbiota, with 977 exercise interventions lasting a minimum of 2-3 months 978 needed to produce consistent changes across alpha and beta 979 diversity and most genera,³⁴ although some of the observ-980 able modifications in gut microbiota occur 2-4 weeks after 981 the onset of an appropriate exercise stimulus.³⁴ As noted 982 previously,⁶¹ the beneficial effects of exercise training on 983 the gut microbiota are rapidly reversed after several weeks 984 of inactivity in both lean and obese individuals, a similar 985 time-course of loss of adaptation observed in patients with 986 celiac disease.¹⁵² In athletes with a prolonged history of 987 exercise training, the alpha-diversity and global composition 988 of the gut microbiome remain unaffected by 3 weeks of 989 reduced exercise,¹⁵³ suggesting that changes in exercise 990 volume have less of an impact on individuals with a back-991 ground of regular PA. However, the duration (months or 992 years) of regular PA required to establish "steady-state" 993 homeostasis in the gut microbiome that persists in the face 994 of the removal of an exercise stimulus is unknown. 995

There remain many challenges facing microbiome sci-996 ence with several fundamental questions still remaining. For 997 example, much of the observed interindividual differences 998 in the gut microbiome can be attributed to the fact that 999 some parts of the total microbiome remain poorly charac-1000 terized, with their biological significance largely unknown. 1001 Nevertheless, an emerging area with potential clinical sig-1002 nificance to improve host health are a number of microbial 1003 interventions, including fecal microbiota transfer (FMT). A 1004 recent study examined the effects of FMT from exercised-1005 trained mice to germ-free animals.¹⁵⁴ Fecal samples from 1006 sedentary and trained animals were gavaged into germ-free 1007 mice. After receiving fecal samples from trained donor mice, 1008 recipient mice had elevated levels of AMPK and insulin-like 1009 growth factor-1 in skeletal muscle along with improved 1010 whole-body glycemia and insulin sensitivity, effects that 1011 were mediated, in part, by the anti-inflammatory properties 1012 of bile acid deconjugation. This study concluded that FMT 1013 mimics the health-promoting effects of exercise by inducing 1014 critical exercise-inducible signal transduction pathways in 1015 skeletal muscle, thereby improving metabolic health 1016 through the "muscle and gut axis."¹⁵⁴ Other studies in mu-1017 rine models of obesity also demonstrate that the beneficial 1018 effects of both exercise and diet are transmissible via 1019 1020

FMT.¹⁵⁵ In humans, FMT involves the infusion of feces from a healthy donor to the GI tract of a recipient patient with the aim of treating diseases associated with alteration of gut microbiota. Although the results from several rodent studies reveal positive outcomes of FMT from both exercise- or dietmanipulated donor animals to germ-free mice on several health markers,^{154,155} data from FMT in humans is lacking for the majority of GI-related disease states. Indeed, consistent with a European consensus conference on FMT in clinical practice,¹⁵⁶ *Clostridium difficile* infection is the only indication where market authorization for FMT has been achieved.

There are several exciting clinical opportunities in microbiome science with the recent application of a host of molecular techniques offering the potential for a greater understanding of the multiplicity and complexity of networks involved in exercise responses and on the mechanisms by which muscle "communicates" with the GI tract to mediate many of the beneficial effects of PA. While the structural characteristics of the gut microbiota have been described through the application of 16S rRNA amplicon sequencing,¹⁵⁷ next-generation sequencing techniques and the creation and integration of functional 'omics readouts (ie, metatranscriptomics, metaproteomics, and metabolomics) will provide more accurate assessment of health and disease states and provide a basis for functional and experimental validation. This is essential as up to 85% of the variance within the human microbiome from population-based studies is still unaccounted.⁵⁷ For example, in a longitudinal study of Q18 patients with Crohn's disease (n = 303) and ulcerative colitis (n = 228), most of the compositional variance of the gut microbiota remained unexplained.¹⁵⁸

Resetting gut microbiome-derived signals of "unhealthy" ageing through personalized or subpopulation-level microbiome-associated interventions is a promising area of research that will be informed by large shotgun metagenomics-based studies and data analytics coupled experimental with preclinical models. Resulting microbiome-based therapeutics for the elderly will need a combination of therapies, including dietary intervention with microbial restoration of lost strains.¹⁵⁹ The identification of microbial profiles in middle-age that confer increased disease risk later in life (eg, biomarkers for colon cancer) present a formidable clinical challenge that will be confounded by the loss of dietary diversity with age, along with the associated decrease of microbiota diversity accompanied by increased risk of inflammation.¹⁶⁰ Levels of habitual PA also decrease with age¹⁶¹ and are associated with sarcopenia and loss of function.¹⁶⁰ The influence of these lifestyle factors on modifying microbiome composition and controlling for them in large-scale human studies presents a huge challenge when trying to unravel the mechanisms underpinning the therapeutic effects of PA and diet interventions on the gut microbiota, and will require integration of large multicenter cohorts, advanced 'omics technologies and sophisticated experimental validations of the mechanistic associations identified.

Increasing evidence demonstrates the association of specific taxa in the development of certain GI diseases

including many cancers; altering the composition of the gut 1081 microbiota to improve the efficacy of anticancer drugs may 1082 be a feasible intervention in the future. Interindividual dif-1083 ferences in patients' treatment-naïve gut microbiome may 1084 play a role in the efficacy of the hosts response to certain 1085 therapies, analogous to "low vs high responders" to exercise 1086 training interventions. Another exciting area for future 1087 research is gaining a better understanding of the relation-1088 ship between gut microbiota, skeletal muscle mass, and 1089 physical function¹⁶² particularly during ageing, which is 1090 associated with reduced microbiota biodiversity, increased 1091 interindividual variability, lower representation of butyrate-1092 producing bacteria, and over-representation of patho-1093 bionts.¹⁶³ Reduced muscle mass has been associated with 1094 distinct microbiota composition and reduced fermentive 1095 capacity in mice, with the administration of probiotics and 1096 butyrate to mouse models of muscle wasting, associated 1097 with improved muscle mass.¹⁶² Especially promising are the 1098 results from preclinical data showing that colonization of 1099 mice with gut microbiota from exercise-trained mice 1100 attenuated the response to chemically induced colitis, 1101 reduced colon shortening, attenuated mucus depletion, and 1102 augmented expression of cytokines involved in tissue 1103 regeneration.¹⁶⁴ As such, future efforts should concentrate 1104 on gaining a deeper understanding of the factors involved in 1105 exercise-gut interactions through the application of 1106 advanced techniques to measure both the microbiome and 1107 the systemic effects of exercise in a variety of diseased 1108 populations. It is hoped that future therapies to treat a range 1109 of GI-related disorders, including cancers, will be based on a 1110 growing recognition that regular PA can positively modify 1111 the human gut microbiome, boost immunity, and decrease 1112 the incidence, progression, and personal burden of these 1113 NCDs. As such, it is recommended that regular PA be 1114 incorporated into standard clinical treatment protocols for 1115 individuals with several GI-related diseases. 1116

References

- 1. Blair SN, Kohl HW 3rd, Paffenbarger RS Jr, et al. Physical fitness and all-cause mortality. A prospective study of healthy men and women. JAMA 1989;262:2395–2401.
- Booth FW, Chakravarthy MV, Gordon SE, et al. Waging war on physical inactivity: using modern molecular ammunition against an ancient enemy. J Appl Physiol {1985} 2002;93:3–30.
- 3. Kodama S, Saito K, Tanaka S, et al. Cardiorespiratory fitness as a quantitative predictor of all-cause mortality and cardiovascular events in healthy men and women: a meta-analysis. JAMA 2009;301:2024–2035.
- 4. Pedersen BK, Saltin B. Evidence for prescribing exercise as therapy in chronic disease. Scand J Med Sci Sports 2006;16(Suppl 1):3–63.
- Pedersen BK, Saltin B. Exercise as medicine evidence for prescribing exercise as therapy in 26 different chronic diseases. Scand J Med Sci Sports 2015;25(Suppl 3):1–72.
- 6. Ruegsegger GN, Booth FW. Health benefits of exercise. Cold Spring Harb Perspect Med 2018;8.

Gastroenterology Vol. ■, Iss. ■

1144

1145

1146

1147

1148

1149

1150

1151

1152

1153

1154

1155

1156

1157

1158

1159

1160

1161

1162

1163

1164

1165

1166

1167

1168

1169

1170

1171

1172

1173

1174

1175

1176

1177

1178

1179

1180

1181

1182

1183

1184

1185

1186

1187

1188

1189

1190

1191

1192

1193

1194

1195

1196

1197

1198

1199

1200

- 7. Cormie P, Trevaskis M, Thornton-Benko E, et al. Exercise medicine in cancer care. Aust J Gen Pract 2020; 49:169–174.
 8. Crospek L, Bapkin NM, Cormia P, et al. "New is the time.
- 8. Czosnek L, Rankin NM, Cormie P, et al. "Now is the time for institutions to be investing in growing exercise programs as part of standard of care": a multiple case study examining the implementation of exercise oncology interventions. Support Care Cancer 2023;31:422.
- 9. Ross R, Blair SN, Arena R, et al. Importance of assessing cardiorespiratory fitness in clinical practice: a case for fitness as a clinical vital sign: a scientific statement from the American Heart Association. Circulation 2016; 134:e653–e699.
- 10. Sallis RE. Exercise is medicine and physicians need to prescribe it. Br J Sports Med 2009;43:3–4.
- Schmitz KH, Campbell AM, Stuiver MM, et al. Exercise is medicine in oncology: engaging clinicians to help patients move through cancer. CA Cancer J Clin 2019; 69:468–484.
- 12. Neufer PD, Bamman MM, Muoio DM, et al. Understanding the cellular and molecular mechanisms of physical activity-induced health benefits. Cell Metab 2015;22:4–11.
- 13. Hawley JA, Hargreaves M, Joyner MJ, et al. Integrative biology of exercise. Cell 2014;159:738–749.
- 14. Cerda B, Perez M, Perez-Santiago JD, et al. Gut microbiota modification: another piece in the puzzle of the benefits of physical exercise in health? Front Physiol 2016;7:51.
- 15. Fiuza-Luces C, Valenzuela PL, Gálvez BG, et al. The effect of physical exercise on anticancer immunity. Nat Rev Immunol 2024;24:282–293.
- 16. Sales KM, Reimer RA. Unlocking a novel determinant of athletic performance: the role of the gut microbiota, short-chain fatty acids, and "biotics" in exercise. J Sport Health Sci 2023;12:36–44.
- 17. Mailing LJ, Allen JM, Buford TW, et al. Exercise and the gut microbiome: a review of the evidence, potential mechanisms, and implications for human health. Exerc Sport Sci Rev 2019;47:75–85.
- Beresford-Jones BS, Forster SC, Stares MD, et al. The mouse gastrointestinal bacteria catalogue enables translation between the mouse and human gut microbiotas via functional mapping. Cell Host Microbe 2022; 30:124–138.e8.
- Hawley JA, Sassone-Corsi P, Zierath JR. Chrono-nutrition for the prevention and treatment of obesity and type 2 diabetes: from mice to men. Diabetologia 2020; 63:2253–2259.
- Boytar AN, Nitert MD, Morrision M, et al. Exerciseinduced changes to the human gut microbiota and implications for colorectal cancer: a narrative review. J Physiol 2022;600:5189–5201.
- 21. Rosshart SP, Herz J, Vassallo BG, et al. Laboratory mice born to wild mice have natural microbiota and model human immune responses. Science 2019;365: eaaw4361.
- 22. Mirzaei R, Afaghi A, Babakhani S, et al. Role of microbiota-derived short-chain fatty acids in cancer development and prevention. Biomed Pharmacother 2021;139:111619.

1139 1140

1117

1118

1119

1120

1121

1122

1123

1124

1125

1126

1127

1128

1129

1130

1131

1132

1133

1134

1135

1136

1137

1138_{Q19}

Exercise and the Gut 11

1201

1202

1203

1204

1205

1206

1207

1208

1209

1210

1211

1212

1213

1214

1215

1216

1217

1218

1219

1220

1221

1222

1223

1224

1225

1226

1244

1245

1247

1248

1254

1255

1256

1257

1258

1259

1260

- 23. Hart NH, Wallen MP, Farley MJ, et al. Exercise and the gut microbiome: implications for supportive care in cancer. Support Care Cancer 2023;31:724.
- 24. Sepich-Poore GD, Zitvogel L, Straussman R, et al. The microbiome and human cancer. Science 2021;371.
- 25. Murphy RM, Watt MJ, Febbraio MA. Metabolic communication during exercise. Nat Metab 2020; 2:805-816.
- 26. Goldstein MS. Humoral nature of the hypoglycemic factor of muscular work. Diabetes 1961;10:232-234.
 - 27. Febbraio MA, Pedersen BK. Muscle-derived interleukin-6: mechanisms for activation and possible biological roles. FASEB J 2002;16:1335-1347.
 - 28. Pedersen BK, Febbraio MA. Muscle as an endocrine organ: focus on muscle-derived interleukin-6. Physiol Rev 2008.
 - 29. Whitham M, Febbraio MA. The ever-expanding myokinome: discovery challenges and therapeutic implications. Nat Rev Drug Discov 2016;15:719-729.
- 30. Weigert C, Hoene M, Plomgaard P. Hepatokines-a novel group of exercise factors. Pflugers Arch 2019; 471:383-396.
- 31. Roca-Rivada A, Al-Massadi O, Castelao C, et al. Muscle tissue as an endocrine organ: comparative secretome profiling of slow-oxidative and fast-glycolytic rat muscle explants and its variation with exercise. J Proteom 2012; 75:5414-5425.
- 1227 32. Cook MD, Allen JM, Pence BD, et al. Exercise and gut 1228 immune function: evidence of alterations in colon im-1229 mune cell homeostasis and microbiome characteristics 1230 with exercise training. Immunol Cell Biol 2016; 1231 94:158-163.
- 1232 33. Monda V, Villano I, Messina A, et al. Exercise modifies 1233 the gut microbiota with positive health effects. Oxidative 1234 Med Cellular Longevity 2017;2017:3831972.
- 1235 34. Boytar AN, Skinner TL, Wallen RE, et al. The effect of 1236 exercise prescription on the human gut microbiota and 1237 comparison between clinical and apparently healthy populations: a systematic review. Nutrients 2023; 1238 15:1534. 1239
- 35. Okamoto T, Morino K, Ugi S, et al. Microbiome poten-1240 tiates endurance exercise through intestinal acetate 1241 production. Am J Physiol Endocrinol Metab 2019; 1242 316:E956-E966. 1243
 - 36. Hawley JA. Microbiota and muscle highway two way traffic. Nat Rev Endocrinol 2020;16:71-72.
- 37. Allen JM, Berg Miller ME, Pence BD, et al. Voluntary and 1246 forced exercise differentially alters the gut microbiome in c57bl/6j mice. J Appl Physiol (1985) 2015; 118:1059-1066.
- 1249 38. Matsumoto M, Inoue R, Tsukahara T, et al. Voluntary 1250 running exercise alters microbiota composition and in-1251 creases n-butyrate concentration in the rat cecum. Bio-1252 sci Biotechnol Biochem 2008;72:572-576. 1253
 - 39. Lambert JE, Myslicki JP, Bomhof MR, et al. Exercise training modifies gut microbiota in normal and diabetic mice. Appl Physiol Nutr Metab 2015; 40:749-752.
 - 40. Gill RK, Saksena S, Alrefai WA, et al. Expression and membrane localization of mct isoforms along the length

of the human intestine. Am J Physiol Cell Physiol 2005: 289:C846-C852.

- 41. Peng L, Li ZR, Green RS, et al. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of amp-activated protein kinase in caco-2 cell monolayers. J Nutr 2009;139:1619-1625.
- 42. Lahiri S, Kim H, Garcia-Perez I, et al. The gut microbiota influences skeletal muscle mass and function in mice. Sci Transl Med 2019;11:eaan5662.
- 43. Mirzaei R, Dehkhodaie E, Bouzari B, et al. Dual role of microbiota-derived short-chain fatty acids on host and pathogen. Biomed Pharmacother 2022;145:112352.
- 44. Clarke SF, Murphy EF, O'Sullivan O, et al. Exercise and associated dietary extremes impact on gut microbial diversity. Gut 2014;63:1913-1920.
- 45. Estaki M. Pither J. Baumeister P. et al. Cardiorespiratory fitness as a predictor of intestinal microbial diversity and distinct metagenomic functions. Microbiome 2016;4:42.
- 46. Barton W, Penney NC, Cronin O, et al. The microbiome of professional athletes differs from that of more sedentary subjects in composition and particularly at the functional metabolic level. Gut 2018;67:625-633.
- 47. Kulecka M, Fraczek B, Mikula M, et al. The composition and richness of the gut microbiota differentiate the top polish endurance athletes from sedentary controls. Gut Microbes 2020;11:1374-1384.
- 48. Himbert C, Stephens WZ, Gigic B, et al. Differences in the gut microbiome by physical activity and BMI among colorectal cancer patients. Am J Cancer Res 2022; 12:4789-4801.
- 49. Wirbel J, Pyl PT, Kartal E, et al. Meta-analysis of fecal metagenomes reveals global microbial signatures that are specific for colorectal cancer. Nat Med 2019; 25:679-689.
- 50. Scheiman J, Luber JM, Chavkin TA, et al. Meta-omics analysis of elite athletes identifies a performanceenhancing microbe that functions via lactate metabolism. Nat Med 2019;25:1104-1109.
- 51. Ahn J, Sinha R, Pei Z, et al. Human gut microbiome and risk for colorectal cancer. J Natl Cancer Inst 2013; 105:1907-1911.
- 52. Ott SJ, Musfeldt M, Wenderoth DF, et al. Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. Gut 2004;53:685-693.
- 53. Dasarathy S. Merli M. Sarcopenia from mechanism to diagnosis and treatment in liver disease. J Hepatol 2016; 65:1232-1244.
- 54. Gold SL, Raman M, Sands BE, et al. Putting some muscle into sarcopenia-the pathogenesis, assessment and clinical impact of muscle loss in patients with inflammatory bowel disease. Aliment Pharmacol Ther 2023;57:1216-1230.
- 55. Aragon-Vela J, Solis-Urra P, Ruiz-Ojeda FJ, et al. Impact of exercise on gut microbiota in obesity. Nutrients 2021;13.
- 56. Mohr AE, Jager R, Carpenter KC, et al. The athletic gut microbiota. J Int Soc Sports Nutr 2020;17:24.
- 57. Shanahan F, Ghosh TS, O'Toole PW. The healthy microbiome-what is the definition of a healthy gut microbiome? Gastroenterology 2021;160:483-494.

1316

1317

1318

1319

Gastroenterology Vol. ■, Iss. ■

1381

1382

1383

1384

1385

1386

1387

1388

1389

1390

1391

1392

1393

1394

1395

1396

1397

1398

1399

1400

1401

1402

1403

1404

1405

1406

1407

1408

1409

1410

1411

1412

1413

1414

1415

1416

1417

1418

1419

1420

1421

1422

1423

1424

1425

1426

1427

1428

1429

1430

1431

1432

1433

1434

1435

1436

1437

1438

- 1321 1322 1323 1324 1325 1326 1327 1328 1329 1330 1331 1332 1333 1334 1335 1336 1337 1338 1339 1340 1341 1342 1343 1344 1345 1346 1347 1348 1349 1350 1351 1352 1353 1354 1355 1356 1357 1358 1359 1360 1361 1362 1363 1364 1365 1366 1367 1368 1369 1370 1371 1372 1373 1374 1375 1376 1377
- 58. Cataldi S, Bonavolontà V, Poli L, et al. The relationship between physical activity, physical exercise, and human gut microbiota in healthy and unhealthy subjects: a systematic review. Biology 2022;11:479.
- 59. Clark A, Mach N. Exercise-induced stress behavior, gutmicrobiota-brain axis and diet: a systematic review for athletes. J Int Soc Sports Nutr 2016;13:43.
- 60. Bonomini-Gnutzmann R, Plaza-Diaz J, Jorquera-Aguilera C, et al. Effect of intensity and duration of exercise on gut microbiota in humans: a systematic review. Int J Environ Res Public Health 2022;19:9518.
- 61. Allen JM, Mailing LJ, Niemiro GM, et al. Exercise alters gut microbiota composition and function in lean and obese humans. Med Sci Sports Exerc 2018;50:747–757.
- 62. Mc Gettigan N, Allen K, Saeidi R, et al. A systematic review of the effect of structured exercise on inflammation and body composition in inflammatory bowel disease. Int J Colorectal Dis 2023;38:143.
- 63. Mc Gettigan N, O'Toole A, Boland K. "Role of exercise in preventing and restoring gut dysbiosis in patients with inflammatory bowel disease": a letter to the editor. World J Gastroenterol 2022;28:878–880.
- 64. Horner KM, Schubert MM, Desbrow B, et al. Acute exercise and gastric emptying: a meta-analysis and implications for appetite control. Sports Med 2015; 45:659–678.
- 65. van Wijck K, Pennings B, van Bijnen AA, et al. Dietary protein digestion and absorption are impaired during acute postexercise recovery in young men. Am J Physiol Regul Integr Comp Physiol 2013;304:R356–R361.
- 66. Lang JA, Gisolfi CV, Lambert GP. Effect of exercise in tensity on active and passive glucose absorption. Int J
 Sport Nutr Exerc Metab 2006;16:485–493.
 - 67. Schmitz L, Ferrari N, Schwiertz A, et al. Impact of
 endurance exercise and probiotic supplementation on
 the intestinal microbiota: a cross-over pilot study. Pilot
 Feasibility Stud 2019;5:76.
 - 68. Costa RJ, Snipe R, Camoes-Costa V, et al. The impact of
 gastrointestinal symptoms and dermatological injuries
 on nutritional intake and hydration status during ultra marathon events. Sports Med Open 2016;2:16.
 - 69. ter Steege RW, Van der Palen J, Kolkman JJ. Prevalence of gastrointestinal complaints in runners competing in a long-distance run: an internet-based observational study in 1281 subjects. Scand J Gastroenterol 2008; 43:1477–1482.
 - De Oliveira EP, Burini RC. Carbohydrate-dependent, exercise-induced gastrointestinal distress. Nutrients 2014;6:4191–4199.
 - 71. Grootjans J, Lenaerts K, Buurman WA, et al. Life and death at the mucosal-luminal interface: new perspectives on human intestinal ischemia-reperfusion. World J Gastroenterol 2016;22:2760–2770.
 - 72. ter Steege RW, Kolkman JJ. Review article: the pathophysiology and management of gastrointestinal symptoms during physical exercise, and the role of splanchnic blood flow. Aliment Pharmacol Ther 2012;35:516–528.
 - Dokladny K, Zuhl MN, Moseley PL. Intestinal epithelial barrier function and tight junction proteins with heat and exercise. J Appl Physiol (1985) 2016;120:692–701.
- 1378 1379 1380

- 74. Zuhl M, Schneider S, Lanphere K, et al. Exercise regulation of intestinal tight junction proteins. Br J Sports Med 2014;48:980–986.
- 75. van Wijck K, Lenaerts K, van Loon LJ, et al. Exerciseinduced splanchnic hypoperfusion results in gut dysfunction in healthy men. PLoS One 2011;6:e22366.
- Bosenberg AT, Brock-Utne JG, Gaffin SL, et al. Strenuous exercise causes systemic endotoxemia. J Appl Physiol (1985) 1988;65:106–108.
- 77. Camus G, Poortmans J, Nys M, et al. Mild endotoxaemia and the inflammatory response induced by a marathon race. Clin Sci (Lond) 1997;92:415–422.
- 78. Gill SK, Teixeira A, Rama L, et al. Circulatory endotoxin concentration and cytokine profile in response to exertional-heat stress during a multi-stage ultra-mara-thon competition. Exerc Immunol Rev 2015;21:114–128.
- 79. Gill SK, Hankey J, Wright A, et al. The impact of a 24-h ultra-marathon on circulatory endotoxin and cytokine profile. Int J Sports Med 2015;36:688–695.
- Robertson JD, Maughan RJ, Davidson RJ. Faecal blood loss in response to exercise. Br Med J (Clin Res Ed) 1987;295:303–305.
- 81. Tota L, Piotrowska A, Palka T, et al. Muscle and intestinal damage in triathletes. PLoS One 2019;14:e0210651.
- Pals KL, Chang RT, Ryan AJ, et al. Effect of running intensity on intestinal permeability. J Appl Physiol (1985) 1997;82:571–576.
- 83. Roca Rubio MF, Folkesson M, Kremp C, et al. Associations between various markers of intestinal barrier and immune function after a high-intensity exercise challenge. Physiol Rep 2024;12:e16087.
- 84. Jeukendrup AE. Training the gut for athletes. Sports Med 2017;47:101–110.
- 85. Burke LM, Hawley JA. Swifter, higher, stronger: what's on the menu? Science 2018;362:781–787.
- Costa RJS, Snipe RMJ, Kitic CM, et al. Systematic review: exercise-induced gastrointestinal syndromeimplications for health and intestinal disease. Aliment Pharmacol Ther 2017;46:246–265.
- Rehrer NJ, Goes E, DuGardeyn C, et al. Effect of carbohydrate on portal vein blood flow during exercise. Int J Sports Med 2005;26:171–176.
- 88. Costa RJS, Miall A, Khoo A, et al. Gut-training: the impact of two weeks repetitive gut-challenge during exercise on gastrointestinal status, glucose availability, fuel kinetics, and running performance. Appl Physiol Nutr Metab 2017;42:547–557.
- 89. Houghton MJ, Snipe RMJ, Williamson G, et al. Plasma measurements of the dual sugar test reveal carbohydrate immediately alleviates intestinal permeability caused by exertional heat stress. J Physiol 2023; 601:4573–4589.
- 90. Scrivin R, Slater G, Mika A, et al. The impact of 48 h high carbohydrate diets with high and low FODMAP content on gastrointestinal status and symptoms in response to endurance exercise, and subsequent endurance performance. Appl Physiol Nutr Metab 2024;49:773–791.
- 91. Lis D, Stellingwerff T, Kitic CM, et al. No effects of a short-term gluten-free diet on performance in nonceliac athletes. Med Sci Sports Exerc 2015;47:2563–2570.

1501

1502

1503

1504

1505

1506

1507

1508

1509

1510

1511

1512

1513

1514

1515

1516

1517

1518

1519

1520

1521

1522

1523

1524

1525

1526

1527

1528

1529

1530

1531

1532

1533

1534

1535

1536

1537

1538

1539

1540

1541

1542

1543

1544

1545

1546

1547

1548

1549

1550

1551

1552

1553

1554

1555

1556

1557

- 1441
 1442
 1443
 1443
 1444
 1444
 1445
 92. Buchman AL, O'Brien W, Ou CN, et al. The effect of arginine or glycine supplementation on gastrointestinal function, muscle injury, serum amino acid concentrations and performance during a marathon run. Int J Sports Med 1999;20:315–321.
- 1445
 1446
 1447
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448
 1448</l
- 1449
 1450
 1451
 1452
 94. Shing CM, Peake JM, Lim CL, et al. Effects of probiotics supplementation on gastrointestinal permeability, inflammation and exercise performance in the heat. Eur J Appl Physiol 2014;114:93–103.
- 1453
 1454
 1454
 1455
 1455
 1456
 1456
 1457
 95. Lamprecht M, Bogner S, Schippinger G, et al. Probiotic supplementation affects markers of intestinal barrier, oxidation, and inflammation in trained men; a randomized, double-blinded, placebo-controlled trial. J Int Soc Sports Nutr 2012;9:45.
- 145896. Wang L, Meng FJ, Jin YH, et al. Effects of probiotic1459supplementation on 12 min run performance, mood1460management, body composition and gut microbiota in1461amateur marathon runners: a double-blind controlled1462trial. J Exerc Sci Fit 2024;22:297–304.
- 146397. Lennon S, Lackie T, Miltko A, et al. Safety and efficacy of1464a probiotic cocktail containing *P. acidilactici* and *L.*1465plantarum for gastrointestinal discomfort in endurance1466runners: randomized double-blinded crossover clinical1467trial. Appl Physiol Nutr Metab 2024;49:890–903.
- 146898. Gill SK, Allerton DM, Ansley-Robson P, et al. Does short-
term high dose probiotic supplementation containing
Lactobacillus casei attenuate exertional-heat stress
induced endotoxaemia and cytokinaemia? Int J Sport
Nutr Exerc Metab 2016;26:268–275.
- 99. Pyne DB, West NP, Cox AJ, et al. Probiotics supplementation for athletes–clinical and physiological effects.
 Eur J Sport Sci 2015;15:63–72.
- 1476100. American College of Sports Medicine. Physical activity
guidelines, 2019.
- 101. Younossi ZM, Corey KE, Lim JK. Aga clinical practice update on lifestyle modification using diet and exercise to achieve weight loss in the management of nonalco-holic fatty liver disease: expert review. Gastroenterology 2021;160:912–918.
- 102. Benede-Ubieto R, Cubero FJ, Nevzorova YA. Breaking
 1484
 1485
 1486
 1486
 102. Benede-Ubieto R, Cubero FJ, Nevzorova YA. Breaking
 the barriers: the role of gut homeostasis in metabolicassociated steatotic liver disease (masld). Gut Microbes 2024;16:2331460.
- 103. Hughes A, Dahmus J, Rivas G, et al. Exercise training reverses gut dysbiosis in patients with biopsy-proven nonalcoholic steatohepatitis: a proof of concept study. Clin Gastroenterol Hepatol 2021;19:1723–1725.
 104. Hughes A, Dahmus J, Rivas G, et al. Exercise training reverses gut dysbiosis in patients with biopsy-proven nonalcoholic steatohepatitis: a proof of concept study.
- 1490104. Kaplan GG. The global burden of ibd: from 2015 to 2025.1491Nat Rev Gastroenterol Hepatol 2015;12:720–727.
- 1492105. Bilski J, Mazur-Bialy A, Brzozowski B, et al. Can exercise
affect the course of inflammatory bowel disease?
Experimental and clinical evidence. Pharmacol Rep
2016;68:827–836.
- 1496
 1497
 106. Ploeger H, Obeid J, Nguyen T, et al. Exercise and inflammation in pediatric Crohn's disease. Int J Sports Med 2012;33:671–679.
- 1499 1500

- 107. Jones K, Kimble R, Baker K, et al. Effects of structured exercise programmes on physiological and psychological outcomes in adults with inflammatory bowel disease (ibd): a systematic review and meta-analysis. PLoS One 2022;17:e0278480.
- 108. Manichanh C, Borruel N, Casellas F, et al. The gut microbiota in ibd. Nat Rev Gastroenterol Hepatol 2012; 9:599–608.
- 109. Baker KA, Miller TD, Marino FE, et al. The exerciseinduced inflammatory response in inflammatory bowel disease: a systematic review and meta-analysis. PLoS One 2022;17:e0262534.
- 110. Ilan K, Motro Y, Nemirovsky A, et al. Cognitive behavioral and mindfulness with daily exercise intervention is associated with changes in intestinal microbial taxa and systemic inflammation in patients with Crohn's disease. Gut Microbes 2024;16:2337269.
- 111. Tiong HT, Fan D, Frampton C, et al. Physical activity is associated with a decreased risk of developing inflammatory bowel disease: a systematic review and metaanalysis. J Crohn's Colitis 2024:jjae053.
- 112. Lund K, Knudsen T, Kjeldsen J, et al. The ibd-fitt study moderate-intensity exercise for patients with inflammatory bowel disease with moderate disease activity: an openlabel randomized controlled trial. Trials 2023;24:742.
- 113. Van De Pol N, Visser E, van der Woude C, et al. P744 evaluation of a lifestyle program based on physical activity on quality of life and fatigue in patients with inflammatory bowel disease: a pilot study. J Crohn's Colitis 2024;18:i1393–i1394.
- 114. McGing JJ, Serres S, Nicholas R, et al. Deconditioning in quiescent Crohn's disease patients with heightened fatigue perception. J Crohn's Colitis 2025:jjae194.
- 115. Johannesson E, Simren M, Strid H, et al. Physical activity improves symptoms in irritable bowel syndrome: a randomized controlled trial. Am J Gastroenterol 2011; 106:915–922.
- 116. Johannesson E, Ringstrom G, Abrahamsson H, et al. Intervention to increase physical activity in irritable bowel syndrome shows long-term positive effects. World J Gastroenterol 2015;21:600–608.
- 117. Lacy BE, Pimentel M, Brenner DM, et al. Acg clinical guideline: management of irritable bowel syndrome. Am J Gastroenterol 2021;116:17–44.
- 118. Bianco A, Russo F, Franco I, et al. Enhanced physical capacity and gastrointestinal symptom improvement in southern italian ibs patients following three months of moderate aerobic exercise. J Clin Med 2023;12.
- 119. Chimienti G, Russo F, Bianco A, et al. Effect of a 12week walking program monitored by global physical capacity score (gpcs) on circulating cell-free mtdna and dnase activity in patients with irritable bowel syndrome. Int J Mol Sci 2024;25.
- 120. Mazumder S, Bindu S, De R, et al. Emerging role of mitochondrial damps, aberrant mitochondrial dynamics and anomalous mitophagy in gut mucosal pathogenesis. Life Sci 2022;305:120753.
- 121. Nunan D, Cai T, Gardener AD, et al. Physical activity for treatment of irritable bowel syndrome. Cochrane Database of Systematic Reviews 2022.

1561

1562

1563

1564

1565

1566

Gastroenterology Vol. ■, Iss. ■

1679

1680

- 122. Li C, Li J, Zhou Q, et al. Effects of physical exercise on the microbiota in irritable bowel syndrome. Nutrients 2024;16:2657.
 138
 123. Arem H, Moore SC, Patel A, et al. Leisure time physical activity and mortality: a detailed pooled analysis of the dose-response relationship. JAMA Intern Med 2015;
- 175:959–967.
 124. McTiernan A, Friedenreich CM, Katzmarzyk PT, et al. Physical activity in cancer prevention and survival: a systematic review. Med Sci Sports Exerc 2019; 51:1252–1261.
- 1571
 125. Moore SC, Lee IM, Weiderpass E, et al. Association of leisure-time physical activity with risk of 26 types of cancer in 1.44 million adults. JAMA Intern Med 2016; 176:816–825.
- 1575
 1576
 1577
 126. Wang Q, Zhou W. Roles and molecular mechanisms of physical exercise in cancer prevention and treatment. J Sport Health Sci 2021;10:201–210.
- 1578
 1579
 1580
 127. Matthews CE, Moore SC, Arem H, et al. Amount and intensity of leisure-time physical activity and lower cancer risk. J Clin Oncol 2020;38:686–697.
- 1581128. Cormie P, Zopf EM, Zhang X, et al. The impact of ex-
ercise on cancer mortality, recurrence, and treatment-
related adverse effects. Epidemiol Rev 2017;39:71–92.
- 1584129. Robsahm TE, Aagnes B, Hjartåker A, et al. Body mass1585index, physical activity, and colorectal cancer by1586anatomical subsites: a systematic review and meta-1587analysis of cohort studies. Eur J Cancer Prevent 2013;158822:492–505.
- 1589 130. Ruiz-Casado A, Martín-Ruiz A, Pérez LM, et al. Exercise
 1590 and the hallmarks of cancer. Trends Cancer 2017;
 1591 3:423–441.
- 1592131. Smit KC, Derksen JWG, Beets GLO, et al. Physical ac-
tivity is associated with improved overall survival among
patients with metastatic colorectal cancer. Cancers
(Basel) 2022;14.
- 132. McTiernan A, Yasui Y, Sorensen B, et al. Effect of a 12month exercise intervention on patterns of cellular proliferation in colonic crypts: a randomized controlled trial.
 Cancer Epidemiol Biomarkers Prevent 2006; 15:1588–1597.
- 1601133. Conti L, Del Cornò M, Gessani S. Revisiting the impact1602of lifestyle on colorectal cancer risk in a gender1603perspective. Crit Rev Oncol/Hematol 2020;145:102834.
- 1604
 134. Chao A, Connell CJ, Jacobs EJ, et al. Amount, type, and timing of recreational physical activity in relation to colon and rectal cancer in older adults: the cancer prevention study ii nutrition cohort. Cancer Epidemiol Biomarkers Prev 2004;13:2187–2195.
- 135. Thune I, Furberg AS. Physical activity and cancer risk: dose-response and cancer, all sites and site-specific. Med Sci Sports Exerc 2001;33:S530–S550; discussion S609–S610.
- 136. Sanchez NF, Stierman B, Saab S, et al. Physical activity reduces risk for colon polyps in a multiethnic colorectal cancer screening population. BMC Res Notes 2012; 5:312.
 137. Deateneulou T. Ntenenio Statheneulou I. Tranninio IC.
- 1010137. Psaltopoulou T, Ntanasis-Stathopoulos I, Tzanninis IG,
et al. Physical activity and gastric cancer risk: a

systematic review and meta-analysis. Clin J Sport Med 2016;26:445–464.

- 138. Behrens G, Jochem C, Keimling M, et al. The association between physical activity and gastroesophageal cancer: systematic review and meta-analysis. Eur J Epidemiol 2014;29:151–170.
- 139. Fang P, Zhou J, Xiao X, et al. The prognostic value of sarcopenia in oesophageal cancer: a systematic review and meta-analysis. J Cachexia Sarcopenia Muscle 2023; 14:3–16.
- 140. Belkaid Y, Hand TW. Role of the microbiota in immunity and inflammation. Cell 2014;157:121–141.
- 141. Ansaldo E, Farley TK, Belkaid Y. Control of immunity by the microbiota. Ann Rev Immunol 2021;39:449–479.
- 142. Rosa-Neto JC, Lira FS, Little JP, et al. Immunometabolism-fit: how exercise and training can modify T cell and macrophage metabolism in health and disease. Exerc Immunol Rev 2022;28.
- 143. Nieman DC, Pence BD. Exercise immunology: future directions. J Sport Health Sci 2020;9:432–445.
- 144. MacNeil B, Hoffman-Goetz L. Effect of exercise on natural cytotoxicity and pulmonary tumor metastases in mice. Med Sci Sports Exerc 1993;25:922–928.
- 145. Deng N, Reyes-Uribe L, Fahrmann JF, et al. Exercise training reduces the inflammatory response and promotes intestinal mucosa-associated immunity in lynch syndrome. Clin Cancer Res 2023;29:4361–4372.
- 146. Neish AS. Mucosal immunity and the microbiome. Ann Am Thor Soc 2014;11:S28–S32.
- 147. Sun X, Zhu M-J. Amp-activated protein kinase: a therapeutic target in intestinal diseases. Open Biol 2017;7: 170104.
- 148. Thompson HJ, Jiang W, Zhu Z. Candidate mechanisms accounting for effects of physical activity on breast carcinogenesis. IUBMB Life 2009;61:895–901.
- 149. Hojman P, Gehl J, Christensen JF, et al. Molecular mechanisms linking exercise to cancer prevention and treatment. Cell Metab 2018;27:10–21.
- 150. Wan R, Chen Y, Feng X, et al. Exercise potentially prevents colorectal cancer liver metastases by suppressing tumor epithelial cell stemness via rps4x downregulation. Heliyon 2024;10:e26604.
- 151. Wang T, Cai G, Qiu Y, et al. Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. ISME J 2012;6:320–329.
- 152. Warbeck C, Dowd AJ, Kronlund L, et al. Feasibility and effects on the gut microbiota of a 12-week high-intensity interval training plus lifestyle education intervention on inactive adults with celiac disease. Appl Physiol Nutr Metab 2021;46:325–336.
- 153. Craven J, Cox AJ, Bellinger P, et al. The influence of exercise training volume alterations on the gut microbiome in highly-trained middle-distance runners. Eur J Sport Sci 2022;22:1222–1230.
- 154. Aoi W, Inoue R, Mizushima K, et al. Exercise-acclimated microbiota improves skeletal muscle metabolism via circulating bile acid deconjugation. Iscience 2023;26.
- 155. Lai Z-L, Tseng C-H, Ho HJ, et al. Fecal microbiota transplantation confers beneficial metabolic effects of

1681

1704

1705

1706

1707

1708

1709

1710

1711

1712

1713

diet and exercise on diet-induced obese mice. Scientif Rep 2018;8:15625.

- 1682
 1683
 1684
 1684
 1685
 1685
 1686
 1687
 1687
 1687
 1688
 1689
 1680
 1680
 1680
 1681
 1682
 1681
 1682
 1682
 1682
 1682
 1683
 1684
 1684
 1685
 1685
 1685
 1685
 1686
 1687
 1687
 1687
 1687
 1688
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687
 1687</l
- 157. Gilbert JA, Quinn RA, Debelius J, et al. Microbiome-wide association studies link dynamic microbial consortia to disease. Nature 2016;535:94–103.
- 1688
1689
1690158. Clooney AG, Eckenberger J, Laserna-Mendieta E, et al.
Ranking microbiome variance in inflammatory bowel
disease: a large longitudinal intercontinental study. Gut
2021;70:499–510.
- 1692
 1693
 159. Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. Nat Rev Gastroenterol Hepatol 2022;19:565–584.
- 1695
 1696
 160. Claesson MJ, Jeffery IB, Conde S, et al. Gut microbiota composition correlates with diet and health in the elderly. Nature 2012;488:178–184.
- 1698
 161. Sallis JF. Age-related decline in physical activity: a synthesis of human and animal studies. Med Sci Sports Exerc 2000;32:1598–1600.
- 1701
 162. Grosicki GJ, Fielding RA, Lustgarten MS. Gut microbiota contribute to age-related changes in skeletal muscle
 1703

size, composition, and function: biological basis for a gut-muscle axis. Calcif Tissue Int 2018;102:433–442.

- 163. Ticinesi A, Nouvenne A, Cerundolo N, et al. Gut microbiota, muscle mass and function in aging: a focus on physical frailty and sarcopenia. Nutrients 2019;11:1633.
- 164. Allen J, Mailing L, Cohrs J, et al. Exercise traininginduced modification of the gut microbiota persists after microbiota colonization and attenuates the response to chemically-induced colitis in gnotobiotic mice. Gut Microbes 2018;9:115–130.

Received September 12, 2024. Accepted January 25, 2025.

Correspondence

Address correspondence to: John A. Hawley, PhD, Australian Catholic University, Mary MacKillop Institute for Health Research, L 5 215 Spring St, Melbourne, VIC 3000, Australia. e-mail: John.hawley@acu.edu.au.

Conflicts of interest

The authors disclose the following: Samuel C. Forster is a consultant to BiomeBank Australia and has acted as an advisor to Microbiotica. Edward M. Giles is an advisor to AbbVie and Janssen; and has received speaker fees from Takeda, Janssen, Nutricia, Aspen Nutritionals, AbbVie, Evolution Health, and H&H Group. The remaining author discloses no conflicts

Funding

No direct funding was received for this work.

1736

1737

1738

1739

1740

1741

1742

1743

1744

1745 1746

1714

1715

1716

1717

1718