

#### Please cite the Published Version

Journé, Valentin ២, Bogdziewicz, Michał ២, Courbaud, Benoit, Kunstler, Georges 몓, Qiu, Tong 몓, Acuña, Marie-Claire Aravena, Ascoli, Davide 몓, Bergeron, Yves, Berveiller, Daniel, Boivin, Thomas, Bonal, Raul, Caignard, Thomas, Cailleret, Maxime, Calama, Rafael, Camarero, J Julio 🔟, Chang-Yang, Chia-Hao 🔟, Chave, Jerome, Chianucci, Francesco, Curt, Thomas, Cutini, Andrea, Das, Adrian, Daskalakou, Evangelia, Davi, Hendrik, Delpierre, Nicolas, Delzon, Sylvain, Dietze, Michael 💿, Calderon, Sergio Donoso, Dormont, Laurent, Espelta, Josep Maria, Farfan-Rios, William, Fenner, Michael, Franklin, Jerry, Gehring, Catherine, Gilbert, Gregory, Gratzer, Georg, Greenberg, Cathryn H, Guignabert, Arthur 🕑, Guo, Qinfeng, Hacket-Pain, Andrew, Hampe, Arndt, Han, Qingmin, Hanley, Mick E, Lambers, Janneke Hille Ris, Holík, Jan, Hoshizaki, Kazuhiko, Ibanez, Ines, Johnstone, Jill F, Knops, Johannes MH 🕑, Kobe, Richard K, Kurokawa, Hiroko 🕑, Lageard, Jonathan <sup>(D)</sup>, LaMontagne, Jalene <sup>(D)</sup>, Ledwon, Mateusz, Lefèvre, François <sup>(D)</sup>, Leininger, Theodor, Limousin, Jean-Marc, Lutz, James 💿, Macias, Diana, Mårell, Anders 💿, McIntire, Eliot 🔟, Moran, Emily V, Motta, Renzo, Myers, Jonathan, Nagel, Thomas A, Naoe, Shoji ២, Noguchi, Mahoko, Norghauer, Julian, Oguro, Michio, Ourcival, Jean-Marc, Parmenter, Robert, Pearse, Ian, Pérez-Ramos, Ignacio M, Piechnik, Łukasz, Podgórski, Tomasz, Poulsen, John, Redmond, Miranda D, Reid, Chantal D, Samonil, Pavel, Scher, C Lane D, Schlesinger, William H, Seget, Barbara, Sharma, Shubhi, Shibata, Mitsue, Silman, Miles, Steele, Michael, Stephenson, Nathan, Straub, Jacob, Sutton, Samantha, Swenson, Jennifer J, Swift, Margaret, Thomas, Peter A, Uriarte, Maria 💿, Vacchiano, Giorgio, Whipple, Amy, Whitham, Thomas, Wright, S Joseph 💿, Zhu, Kai 💿, Zimmerman, Jess, Żywiec, Magdalena 厄 and Clark, James S (2024) The Relationship Between Maturation Size and Maximum Tree Size From Tropical to Boreal Climates. Ecology Letters, 27 (9). e14500 ISSN 1461-023X

#### DOI: https://doi.org/10.1111/ele.14500

(cc) BY

Publisher: Wiley

Version: Accepted Version

Downloaded from: https://e-space.mmu.ac.uk/636094/

Usage rights:

Creative Commons: Attribution 4.0

**Additional Information:** This is an accepted manuscript of an article which appeared in final form in Ecology Letters, published by Wiley

**Data Access Statement:** Data and code supporting our results are archived on the Open Science Framework (OSF) Repository: <a href="https://doi.org/10.17605/OSF.IO/U23VY">https://doi.org/10.17605/OSF.IO/U23VY</a>. All analyses used R Core Team (2023) (v4.3.0) and published R packages.

#### Enquiries:

If you have questions about this document, contact openresearch@mmu.ac.uk. Please include the URL of the record in e-space. If you believe that your, or a third party's rights have been compromised through this document please see our Take Down policy (available from https://www.mmu.ac.uk/library/using-the-library/policies-and-guidelines)



## The Relationship between Maturation Size and Maximum Tree Size from Tropical to Boreal Climates

Valentin Journé, Bogdziewicz Michal, Courbaud Benoit, Kunstler Georges, Qiu Tong, Ascoli Davide, Bergeron Yves, Berveiller Daniel, Boivin Thomas, Bonal Raul, et al.

## ▶ To cite this version:

Valentin Journé, Bogdziewicz Michal, Courbaud Benoit, Kunstler Georges, Qiu Tong, et al.. The Relationship between Maturation Size and Maximum Tree Size from Tropical to Boreal Climates. Ecology Letters, In press. hal-04718869

## HAL Id: hal-04718869 https://hal.science/hal-04718869v1

Submitted on 2 Oct 2024

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.



Distributed under a Creative Commons Attribution 4.0 International License

# The Relationship between Maturation Size and <sup>2</sup> Maximum Tree Size from Tropical to Boreal Climates

3

4

Valentin Journé,<sup>1,2</sup> Michał Bogdziewicz,<sup>2</sup> Benoit Courbaud,<sup>1</sup> Georges Kunstler,<sup>1</sup> Tong Qiu,<sup>3</sup> 5 Marie-Claire Aravena Acuña,<sup>4</sup> Davide Ascoli,<sup>5</sup> Yves Bergeron,<sup>6</sup> Daniel Berveiller,<sup>7</sup> Thomas Boivin,<sup>8</sup> 6 Raul Bonal,<sup>9</sup> Thomas Caignard,<sup>10</sup> Maxime Cailleret,<sup>11</sup> Rafael Calama,<sup>12</sup> J. Julio Camarero,<sup>13</sup> Chia-Hao Chang-Yang,<sup>14</sup> Jerome Chave,<sup>15</sup> Francesco Chianucci,<sup>16</sup> Thomas Curt,<sup>17</sup> Andrea Cutini,<sup>18</sup> Adrian Das,<sup>19</sup> Evangelia Daskalakou,<sup>20</sup> Hendrik Davi,<sup>8</sup> Nicolas Delpierre,<sup>7</sup> Sylvain Delzon,<sup>10</sup> Michael Dietze,<sup>21</sup> Sergio Donoso Calderon,<sup>22</sup> Laurent Dormont,<sup>23</sup> Josep Maria Espelta,<sup>24</sup> William Farfan-10 Rios,<sup>25</sup> Michael Fenner,<sup>26</sup> Jerry Franklin,<sup>27</sup> Catherine Gehring,<sup>28</sup> Gregory Gilbert,<sup>29</sup> Georg Gratzer,<sup>30</sup> 11 Cathryn H. Greenberg,<sup>31</sup> Arthur Guignabert,<sup>32</sup> Qinfeng Guo,<sup>33</sup> Andrew Hacket-Pain,<sup>34</sup> Arndt Hampe,<sup>35</sup> 12 Qingmin Han,<sup>36</sup> Mick E. Hanley,<sup>37</sup> Janneke Hille Ris Lambers,<sup>38</sup> Jan Holík,<sup>39</sup> Kazuhiko Hoshizaki,<sup>40</sup> 13 Ines Ibanez,<sup>41</sup> Jill F. Johnstone,<sup>42</sup> Johannes M. H. Knops,<sup>43</sup> Richard K. Kobe,<sup>44</sup> Hiroko Kurokawa,<sup>45</sup> 14 Jonathan Lageard,<sup>46</sup> Jalene LaMontagne,<sup>47</sup> Mateusz Ledwon,<sup>48</sup> François Lefèvre,<sup>8</sup> Theodor Leininger,<sup>49</sup> 15 Jean-Marc Limousin,<sup>50</sup> James Lutz,<sup>51</sup> Diana Macias,<sup>52</sup> Anders Mårell,<sup>53</sup> Eliot McIntire,<sup>54</sup> Emily V. 16 Moran,<sup>55</sup> Renzo Motta,<sup>5</sup> Jonathan Myers,<sup>56</sup> Thomas A. Nagel,<sup>57</sup> Shoji Naoe,<sup>58</sup> Mahoko Noguchi,<sup>58</sup> 17 Julian Norghauer,<sup>59</sup> Michio Oguro,<sup>45</sup> Jean-Marc Ourcival,<sup>50</sup> Robert Parmenter,<sup>60</sup> Ian Pearse,<sup>61</sup> Ig-18 nacio M. Pérez-Ramos,<sup>62</sup> Łukasz Piechnik,<sup>63</sup> Tomasz Podgórski,<sup>64</sup> John Poulsen,<sup>65</sup> Miranda D. 19 Redmond,<sup>66</sup> Chantal D. Reid,<sup>67</sup> Pavel Samonil,<sup>39</sup> C. Lane Scher,<sup>67</sup> William H Schlesinger,<sup>67</sup> Barbara 20 Seget, 63 Shubhi Sharma, 68 Mitsue Shibata, 45 Miles Silman, 69 Michael Steele, 70 Nathan Stephenson, 19 21 Jacob Straub,<sup>71</sup> Samantha Sutton,<sup>67</sup> Jennifer J. Swenson,<sup>72</sup> Margaret Swift,<sup>67</sup> Peter A. Thomas,<sup>73</sup> 22 Maria Uriarte,<sup>74</sup> Giorgio Vacchiano,<sup>75</sup> Amy Whipple,<sup>76</sup> Thomas Whitham,<sup>76</sup> S. Joseph Wright,<sup>77</sup> Kai 23 Zhu,<sup>78</sup> Jess Zimmerman,<sup>79</sup> Magdalena Żywiec,<sup>63</sup> James S. Clark,<sup>1,67</sup> 24 25

<sup>26</sup> <sup>1</sup>Universite Grenoble Alpes, Institut National de Recherche pour Agriculture, Alimentation et Environnement

27 (INRAE), Laboratoire EcoSystemes et Societes En Montagne (LESSEM), 38402 St. Martin-d'Heres, France.

<sup>28</sup> <sup>2</sup>Forest Biology Center, institute of environmental biology, Adam Mickiewicz University in Poznan, Poland.

<sup>3</sup>Department of Ecosystem Science and Management, Pennsylvania State University, University Park, PA
 16802 USA.

<sup>31</sup> <sup>4</sup>Centro Austral de Investigaciones Científicas (CADIC), Consejo Nacional de Investigaciones Científicas y

<sup>32</sup> Técnicas (CONICET), B. Houssay 200 (9410) Ushuaia, Tierra del Fuego, Argentina.

<sup>33</sup> <sup>5</sup>Department of Agriculture, Forest and Food Sciences, University of Torino, 10095 Grugliasco, TO, Italy.

- <sup>6</sup>Forest Research Institute, University of Quebec in Abitibi-Temiscamingue, Rouyn-Noranda, QC J9X 5E4,
   <sup>35</sup> Canada.
- <sup>36</sup> <sup>7</sup>Université Paris-Saclay, CNRS, AgroParisTech, Ecologie Systématique et Evolution, 91190, Gif-sur-Yvette, <sup>37</sup> France.
- <sup>38</sup> <sup>8</sup>Institut National de Recherche pour Agriculture, Alimentation et Environnement (INRAE), Ecologie des
- <sup>39</sup> Forets Mediterranennes, 84000 Avignon, France.
- <sup>40</sup> <sup>9</sup>Department of Biodiversity, Ecology and Evolution, Complutense University of Madrid, 28040 Madrid, Spain.
- <sup>41</sup> <sup>10</sup>Universite Bordeaux, Institut National de Recherche pour Agriculture, Alimentation et Environnement (IN-
- RAE), Biodiversity, Genes, and Communities (BIOGECO), 33615 Pessac, France.
- <sup>43</sup> <sup>11</sup>INRAE, Aix-Marseille University, UMR RECOVER, Aix-en-Provence, France.
- <sup>44</sup> <sup>12</sup>ICIFOR (Forest Research Institute). INIA-CSIC. 28040 Madrid. Spain.
- <sup>45</sup> <sup>13</sup>Instituto Pirenaico de Ecologia, Consejo Superior de Investigaciones Científicas (IPE-CSIC), 50059 Zaragoza,
   <sup>46</sup> Spain.
- <sup>47</sup> <sup>14</sup>Department of Biological Sciences, National Sun Yat-sen University, Kaohsiung 80424, Taiwan.
- <sup>48</sup> <sup>15</sup>Unité Evolution et Diversité Biologique (EDB), CNRS, IRD, UPS, 118 route de Narbonne, 30162 Toulouse,
- 49 France.
- <sup>50</sup> <sup>16</sup>CREA Research Ventre for Forestry and Wood, Viale S. Margherita 80, 52100 Arezzo, Italy.
- <sup>51</sup> <sup>17</sup>Aix Marseille universite, Institut National de Recherche pour Agriculture, Alimentation et Environnement
- 52 (INRAE), 13182 Aix-en-Provence, France.
- <sup>53</sup> <sup>18</sup>Research Centre for Forestry and Wood, Arezzo, Italy.
- <sup>54</sup> <sup>19</sup>USGS Western Ecological Research Center, Three Rivers, CA, 93271 USA.
- <sup>55</sup> <sup>20</sup>Institute of Mediterranean and Forest Ecosystems, Hellenic Agricultural Organization, 11528 Athens, Greece.
- <sup>56</sup> <sup>21</sup>Earth and Environment, Boston University, Boston, MA, 02215 USA.
- <sup>57</sup> <sup>22</sup>Universidad de Chile, Facultad de Ciencias Forestales y de la Conservacion de la Naturaleza (FCFCN), La
- <sup>58</sup> Pintana, 8820808 Santiago, Chile.
- <sup>59</sup> <sup>23</sup>Centre d'Ecologie Fonctionnelle et Evolutive (CEFE), Centre National de la Recherche Scientifique (CNRS),
- 60 34293 Montpellier, France.
- <sup>61</sup> <sup>24</sup>Centre de Recerca Ecologica i Aplicacions Forestals (CREAF), Bellaterra, Catalunya 08193, Spain.
- <sup>62</sup> <sup>25</sup>Biology Department, Center for Energy, Environment, and Sustainability, Wake Forest University, Winston
- 63 Salem, NC, United States.
- <sup>64</sup> <sup>26</sup>Biology Department, University of Southampton, United Kingdom.
- <sup>65</sup> <sup>27</sup>Forest Resources, University of Washington, Seattle, WA 98195 USA.
- <sup>66</sup> <sup>28</sup>Department of Biological Sciences and Center for Adaptive Western Landscapes.
- <sup>67</sup> <sup>29</sup>Department of Environmental Studies, University of California, Santa Cruz, CA 95064 USA.
- <sup>30</sup>Institute of Forest Ecology; Department of Forest- and Soil Sciences, University of Natural Resources and
- <sup>69</sup> Life Sciences, Vienna.
- <sup>70</sup> <sup>31</sup>Bent Creek Experimental Forest, USDA Forest Service, Asheville, NC 28801 USA.
- <sup>71</sup> <sup>32</sup>INRAE, Bordeaux Sciences Agro, UMR 1391 ISPA, Villenave d'Ornon, France.
- <sup>72</sup> <sup>33</sup>Eastern Forest Environmental Threat Assessment Center, USDA Forest Service, Southern Research Sta-
- tion, Research Triangle Park, NC 27709 USA.
- <sup>74</sup> <sup>34</sup>Department of Geography and Planning, School of Environmental Sciences, University of Liverpool, Liver-

- 75 pool, United Kingdom.
- <sup>76</sup> <sup>35</sup>BIOGECO, INRA, University of Bordeaux, Cestas, France.
- <sup>77</sup> <sup>36</sup>Department of Plant Ecology Forestry and Forest Products Research Institute (FFPRI), Tsukuba, Ibaraki,
- 78 305-8687 Japan.
- <sup>79</sup> <sup>37</sup>School of Biological and Marine Sciences, University of Plymouth, Plymouth, United Kingdom.
- <sup>38</sup>Plant Ecology, Institute of Integrative Biology, D-USYS, ETH Zürich, Zürich, Switzerland.
- <sup>39</sup>Department of Forest Ecology, Silva Tarouca Research Institute, 60200 Brno, Czech Republic.
- <sup>40</sup>Department of Biological Environment, Akita Prefectural University, Akita 010-0195, Japan.
- <sup>83</sup><sup>41</sup>School for Environment and Sustainability, University of Michigan, Ann Arbor, MI 48109.
- <sup>42</sup>Institute of Arctic Biology, University of Alaska, Fairbanks, AK 99700, USA.
- $^{43}$ Health and Environmental Sciences Department, Xian Jiaotong-Liverpool University, Suzhou, China, 215123.
- <sup>44</sup>Department of Plant Biology, Program in Ecology, Evolutionary Biology, and Behavior, Michigan State Uni-
- versity, East Lansing, MI 48824.
- <sup>45</sup>Department of Forest vegetation, Forestry and Forest Products Research Institute, Tsukuba, Ibaraki, 305-
- 89 8687, Japan.
- <sup>90</sup> <sup>46</sup>Department of Natural Sciences, Manchester Metropolitan University, Manchester M1 5GD, UK.
- <sup>91</sup> <sup>47</sup>Department of Biological Sciences, DePaul University, Chicago, IL 60614 USA.
- <sup>92</sup> <sup>48</sup>Institute of Systematics and Evolution of Animals, Polish Academy of Sciences, Slawkowska 17, 31-016
- 93 Krakow, Poland.
- <sup>49</sup>USDA, Forest Service, Southern Research Station, PO Box 227, Stoneville, MS 38776.
- <sup>95</sup> <sup>50</sup>CEFE, Univ Montpellier, CNRS, EPHE, IRD, 1919 route de Mende, 34293 Montpellier Cedex 5, France.
- <sup>96</sup> <sup>51</sup>Department of Wildland Resources, and the Ecology Center, Utah State University, Logan, UT 84322 USA.
- <sup>97</sup> <sup>52</sup>Department of Biology, University of New Mexico, Albuquerque, NM 87131 USA.
- 98 <sup>53</sup>INRAE, UR EFNO, FR-45290 Nogent-sur-Vernisson, France.
- <sup>99</sup> <sup>54</sup>Pacific Forestry Centre, Victoria, British Columbia, V8Z 1M5 Canada.
- <sup>55</sup>School of Natural Sciences, UC Merced, Merced, CA 95343 USA.
- <sup>56</sup>Department of Biology, Washington University in St. Louis, St. Louis, MO.
- <sup>57</sup>Department of forestry and renewable forest resources, Biotechnical Faculty, University of Ljubljana, Ljubljana, Slovenia.
- <sup>104</sup> <sup>58</sup>Tohoku Research Center, Forestry and Forest Products Research Institute, Morioka, Iwate, 020-0123, <sup>105</sup> Japan.
- <sup>106</sup> <sup>59</sup>Institute of Plant Sciences, University of Bern, Altenbergrain 21, Bern 3013, Switzerland.
- <sup>60</sup>Valles Caldera National Preserve, National Park Service, Jemez Springs, NM 87025 USA.
- <sup>108</sup> <sup>61</sup>Fort Collins Science Center, 2150 Centre Avenue, Bldg C, Fort Collins, CO 80526 USA.
- <sup>62</sup>Inst. de Recursos Naturales y Agrobiologia de Sevilla, Consejo Superior de Investigaciones Científicas
   (IRNAS-CSIC), Seville, Andalucia, Spain.
- <sup>63</sup>W. Szafer Institute of Botany, Polish Academy of Sciences, Lubicz 46, 31-512 Krakow, Poland.
- <sup>112</sup> <sup>64</sup>Department of GameManagement and Wildlife Biology, Faculty of Forestry and Wood Sciences, Czech Uni-
- versity of Life Sciences Prague; Kamýcká 129, 165 00 Prague, Czech Republic.
- <sup>114</sup> <sup>65</sup>The Nature Conservancy, Boulder, CO 80302.
- <sup>115</sup> <sup>66</sup>Department of Environmental Science Policy and Management, University of California Berkeley, Berkeley,

- 116 California, USA.
- <sup>67</sup>Nicholas School of the Environment, Duke University, Durham, NC 27708 USA.
- <sup>118</sup> <sup>68</sup>Ecology and Evolutionary Biology Department, Yale University, 165 Prospect Street, New Haven, USA.
- <sup>69</sup>Department of Biology, Wake Forest University, 1834 Wake Forest Rd, Winston-Salem, NC 27106 USA.
- <sup>120</sup> <sup>70</sup>Department of Biology, Wilkes University, 84 West South Street, Wilkes-Barre, PA 18766 USA.
- <sup>121</sup> <sup>71</sup>Department of Environmental Science and Ecology, State University of New York-Brockport, Brockport, NY

122 14420 USA.

- <sup>72</sup>Data Science, College of William and Mary, Williamsburg, VA, USA.
- <sup>124</sup> <sup>73</sup>School of Life Sciences, Keele University, Staffordshire ST5 5BG, UK.
- <sup>125</sup> <sup>74</sup>Department of Ecology, Evolution and Environmental Biology, Columbia University, 1113 Schermerhorn
- 126 Ext., 1200 Amsterdam Ave., New York, NY 10027.
- <sup>127</sup> <sup>75</sup>Department of Agricultural and Environmental Sciences Production, Territory, Agroenergy (DISAA), Uni-
- versity of Milan, 20133 Milano, Italy.
- <sup>76</sup>Department of Biological Sciences, Northern Arizona University, Flagstaff, AZ 86011 USA.
- <sup>130</sup> <sup>77</sup>Smithsonian Tropical Research Institute, Apartado 0843n03092, Balboa, Republic of Panama.
- <sup>78</sup>School for Environment and Sustainability, University of Michigan, Ann Arbor, Michigan, USA.
- <sup>132</sup> <sup>79</sup>Department of Environmental Sciences, University of Puerto Rico, Rio Piedras, PR 00936 USA

133

The fundamental trade-off between current and future reproduction has 134 long been considered to result in a tendency for species that can grow 135 large to begin reproduction at a larger size. Due to the prolonged time 136 required to reach maturity, estimates of tree maturation size remain very 137 rare and we lack a global view on the generality and the shape of this 138 trade-off. Using seed production from five continents, we estimate tree 139 maturation sizes for 486 tree species spanning tropical to boreal climates. 140 Results show that a species' maturation size increases with maximum 141 size, but in a non-proportional way: the largest species begin reproduc-142 tion at smaller sizes than would be expected if maturation were simply 143 proportional to maximum size. Furthermore, the decrease in relative mat-144 uration size is steepest in cold climates. These findings on maturation 145 size drivers are key to accurately represent forests' responses to distur-146 bance and climate change. 147

- *keywords:* tree fecundity | size | seed production | tree maturation | life-history | allometry
- 149

#### **Running title:**

<sup>151</sup> Relation between Maturation and Maximum Tree Size

- 152
- <sup>153</sup> Type of article: Synthesis

#Total word count abstract: ~149 #Total word count main text: ~ 5300 # 78 references # 1
 Table and 6 Figures (color) # 1 supplementary material

156

#### 157 Data and code availability statement

Data and code supporting our results are archived on the Open Science Framework (OSF)
 Repository: https://doi.org/10.17605/0SF.IO/U23VY. All analyses used R Core Team
 (2023) (v4.3.0) and published R packages.

161

#### **162** Author contributions

<sup>163</sup> V.J., G.K. and J.S.C performed analyses, led the paper, and designed the study. V.J., M.B.,
 <sup>164</sup> B.C., G.K., T.Q, and J.S.C co-wrote the paper. J.S.C compiled the MASTIF network, and
 <sup>165</sup> wrote the MASTIF model and software. All authors contributed data and revised the paper.

#### 167 correspondence:

- Journé Valentin\*: journe.valentin@gmail.com & James S. Clark†: jimclark@duke.edu
- <sup>169</sup> \*Forest Biology Center, institute of environmental biology, Adam Mickiewicz University in Poznan, Poland.
- 170 † Nicholas School of the Environment, Duke University, Durham, NC 27708 USA.

## 171 Introduction

The size or age at maturity is critical for tree population fitness and forest regeneration because recruitment opportunities can occur when trees are any size or age (Dietze & Clark, 2008; McDowell *et al.*, 2020; Qiu *et al.*, 2021). In trees, reproduction follows an extended maturation phase (Thomas, 1996; Clark *et al.*, 2004). Juvenile allocation to leaves and the roots and architecture that supports them build the large light- and waterharvesting capacity characteristic of the tree life form. Allocation can then shift to include reproduction. This delayed maturation of trees is linked to the fundamental trade-off between current and future reproduction (Stearns, 1989).

On one hand, delayed reproduction sacrifices early seed production to reap future benefits. In environments marked by intense competition and a reliable future, delayed reproduction benefits from large size and the resources that accumulate if juveniles allocate to growth and survival (Falster & Westoby, 2003; Wenk & Falster, 2015). On the other hand, the advantages of large size can come with costs, including water transport high into the crown and biomechanical risk of bole fracture or windthrow (Niklas, 1994; Koch *et al.*, 2004; Dietze & Clark, 2008; Lines *et al.*, 2012). Current reproduction avoids the risks of an uncertain future, and it contributes most to fitness in non-competitive or frequently disturbed environments (Charlesworth, 2000).

Among tree species, the potential trade-off between current and future reproduction might require a balance of benefits and risks and result in a positive association across species in maturation and maximum size or age (Loehle, 1988; Thomas, 1996; Davies & Ashton, 1999; Westoby *et al.*, 2002; Falster & Westoby, 2003; Wenk & Falster, 2015; Visser *et al.*, 2016). Our understanding of the variation of maturation size among tree species is, however, extremely limited and there are no large-scale studies on this topic. We thus have a poor understanding of how maturation size varies with species maximum size and the relative importance of other factors such as species climate niche and functional traits.

In the absence of maturation estimates, earth system models (ESMs) incorporate assumptions that are 193 expected to bias lifetime reproduction. Many ESMs omit impacts of life history on disturbance response 194 entirely (see McDowell et al. 2020 for a review). In models that do accommodate life history, maturation size 195  $-d_{mat}$  (for diameter) – is independent of maximum size  $-d_{max}$  (Kohler & Huth, 2004; Wallentin *et al.*, 2008; 196 Yang et al., 2022) (Fig. 1, black dotted line). An alternative hypothesis is that maturation size is proportional 197 to maximum size. It is consistent with a study at Barro Colorado Island in Panama, where Visser et al. 198 (2016) reported that  $d_{mat} = d_{max}/2$  (red dashed line in Fig. 1). This is related to the classical prediction 199 of a proportionate increase in maturation age with increased maximum age from simple optimization models 200 in animals (Charnov & Berrigan, 1990, 1991; Jensen, 1996; Thorson et al., 2017) and trees (Clark, 1991). 201 Indeed, if the predictions for age also apply to size, then this proportionate risk model means that the 202 maturation delay incurred for increased maximum size is the same for species large and small, represented 203 by the red line in Fig. 1. 204

In fact, it would be remarkable if this relationship was the same for species of all sizes because the constraints on the large size and the relative contribution to fitness of early seed production could vary widely depending on the species' maximum size. If mortality risks and allocation demands change with age and size (Charnov & Berrigan, 1990), then the strictly proportional relationship between maturation size ( $d_{mat}$ ) and maximum size ( $d_{max}$ ) can be generalized to a power relationship,

$$d_{mat} = \alpha \times d_{max}^{\beta_d} \tag{1}$$

The proportionate model means that  $\beta_d = 1$  in Eq. (1) (Fig. 1a). An **accelerating risk model** refers to the case where the maturation delay required for large species to increase maximum size is greater than for small species. If  $\beta_d > 1$  (purple in Fig. 1), the relative size at maturation ( $d_{rel} = d_{mat}/d_{max}$ ) continues to increase for species in the largest size classes (Fig. 1b). Consider, for example, an expected fitness gain from extending end-of-life reproduction that comes with increased size and age. The early life investments in structural support or defenses needed for an incremental increase in maximum size might be especially high for the largest species.

Alternatively, a **diminishing risk model** refers to the case where the need to further delay maturation size declines for species already at large size. If  $0 < \beta_d < 1$ , then species that reach large size do not incur the same proportionate delay risk as small species. As  $\beta_d$  approaches zero, the largest species mature at nearly the same size as the small species. Then, the relative size at maturation  $d_{rel}$  declines with maximum size (Fig. 1b).

Improved understanding of maturation size confronts challenges posed by slow dynamics and limited 222 observation. Where there is a dense canopy, the reproductive status of individual trees is often not detected, 223 and crop failures (little or no seed production in trees that have reached maturity) are common. Likewise, seed 224 traps often fail to recover seeds from nearby reproductive trees, especially the low seed production of newly 225 mature individuals (LaDeau & Clark, 2001). Detection error can be minimized where observations come from 226 above the canopy or open settings like savannas, seed orchards, or common gardens (LaDeau & Clark, 2001; 227 Caignard et al., 2021). Still, a time series of observations is needed because an individual reproducing this 228 year will have matured at some time in the past. Estimates of maturation status from time series data allow 229 for detection error similar to the way it is used in capture-recapture models. Maturation can be treated as 230 a hidden Markov process (see detailed Supplements to Clark et al. 2004, 2019). As in capture-recapture 231 models, the probability that an individual is mature in a given year depends not only on failure to detect in the 232 current year but also on the history and future of observations on the same individual. The more times that 233 reproduction is not detected in the past (or future), the lower the probability that a tree is mature now. In tree-234 fecundity studies, the complexity is compounded by the "masting" phenomenon, where quasi-synchronous, 235 quasi-periodic crops require observations over several years, making a large number of observations in a 236 single year insufficient. Most studies where maturation size has been estimated focus on open-grown trees 237 and/or have limited taxonomic breadth, habitat variation, or both (Wenk et al., 2018; Thomas, 1996; Davies & 238 Ashton, 1999; Kohyama et al., 2003; Wright et al., 2005; Visser et al., 2016; Minor & Kobe, 2019). 239

Inferring the relationship between maturation and maximum size has also to control for the environment (Wenk & Falster, 2015) and species characteristics (Visser *et al.*, 2016). While the effects of climate on maturation size are unknown, tree fecundity responds to seasonal temperature and moisture, soils, and light availability, which depends on the local competitive environment (Clark *et al.*, 2014; Caignard *et al.*, 2017; Minor & Kobe, 2019; Le Roncé *et al.*, 2021; Qiu *et al.*, 2022; Journé *et al.*, 2022). Also, fast growth and accelerated competition that comes from long growing seasons in the wet tropics do not necessarily imply small or large maturation sizes.

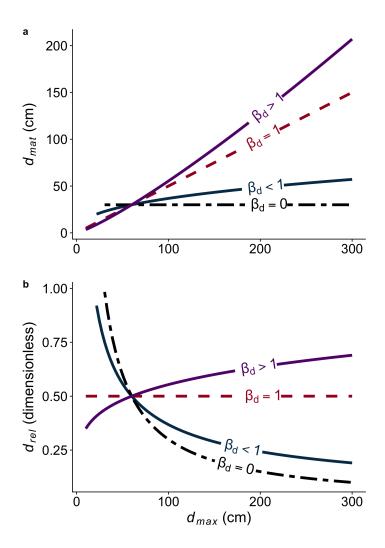


Figure 1: Hypothesized association between maturation size  $(d_{mat})$  and maximum size  $(d_{max})$  (**a**) and the relative size at maturation  $(d_{rel} = d_{mat}/d_{max})$  (**b**) (Eq. (1)). To highlight the effects of size (parameter  $\beta_d$ ), values of parameter  $\alpha$  are selected to yield an equivalent diameter at  $d_{max} = 60$  cm. Two "baseline" hypotheses (dashed lines) are independence between  $d_{mat}$  and  $d_{max}$  (black dotted) and proportionate delay (red dotted), the latter is expected if increased size incurs the same maturation delay at all size classes. Two alternative hypotheses are increasing (purple - accelerating risk model) or decreasing (blue - diminishing risk model) maturation delays in the largest size classes.

The relationship between maturation and maximum size could be associated with other plant functional 247 traits, that would reflect diverse plant strategies. Fast growth in open environments is often associated with 248 low wood density and high specific leaf area (SLA) (Moles et al., 2004, 2006; Thomas et al., 2015; Visser 249 et al., 2016; Wenk et al., 2018). However, it is unclear whether the same traits that are involved in fast growth 250 are also associated with maturation at a small size. Furthermore, if large-seeded species need to accumulate 251 resource reserves, then there could be a positive association between seed size and maturation size (Moles 252 et al., 2004). Relatedly, high reproductive expenditures, measured as seed size × seed number (Qiu et al., 253 2022), might be associated with delayed maturation size. Due to their co-dependence, it is necessary to 254 model all of these traits jointly, while accounting for the effects of habitat and phylogenetic groups (Clark, 255 2016; Seyednasrollah & Clark, 2020; Bogdziewicz et al., 2023; Qiu et al., 2023). 256

In this study, we provide the first comprehensive estimates of tree maturation size, obtained for 486 tree 257 species on five continents, incorporating effects of the environment over a large range of tree diameters 258 and habitats. We use the Masting Inference and Forecasting (MASTIF) network and modeling framework 259 to accommodate the dependence between observations between trees and within trees over time (Clark 260 et al., 2021; Sharma et al., 2022; Qiu et al., 2022; Journé et al., 2022). Based on MASTIF estimates we 261 derive maturation size as tree diameter at the onset of female reproductive function allowing us to compare 262 maturation sizes across species that vary in reproductive biology (e.g., Pinaceae commonly produce male 263 cones earlier than female cones; many species have no such separation) and where pollen production can 264 be hard to quantify. We first evaluate how maturation varies with species' maximum size and test the three 265 alternative models of Fig. 1 with our estimates of maturation size and estimates of species' maximum size. 266 Then, we evaluate how the relationship between maturation size and maximum size is influenced by climate 267 and its association with other plant functional traits. 268

## <sup>269</sup> Materials and Methods

Our analysis includes three elements (Fig. 2). We first parameterize a model for individual maturation status 270 and fecundity based on diameter, shade conditions, and environmental variables (Fig. 2a). The year in which 271 an individual achieves maturity is almost never observed. Instead, seeds counted in traps or in crowns vary 272 from year to year. Successive observations represent a time series for every tree. This first step estimates 273 maturation status and conditional fecundity (seeds per tree per year given that it is mature) for all trees in 274 the network. From this fitted model, we generate predictive distributions of maturation status across diameter 275 with other variables held at intermediate values to estimate  $d_{mat}$  from the model. Again, this prediction 276 from the model is necessitated by the fact that true maturation status is an estimate, not a state that is directly 277 observed. Second, we estimated the model of Fig. 1 to obtain estimates of  $\alpha$  and  $\beta_d$  (Fig. 2b), while controlling 278 for other variables that could affect their relationship. Finally, we evaluate the species-level trait relationship 279 that includes maturation size (Fig. 2c). The following section describes these elements of the analysis. 280

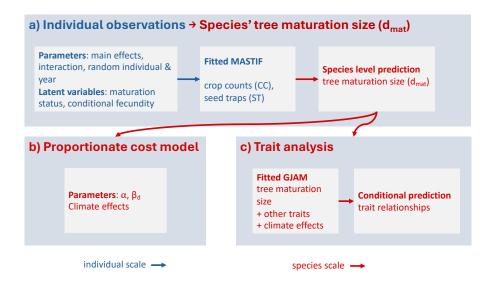


Figure 2: Three elements of the analysis include a) an individual-scale analysis (blue) to estimate maturation status each year and to parameterize relationships that control maturation. This fitted model is the basis for species-level prediction of maturation size (red). b) Species-level expected maturation size based on the proportionate risk model, controlling for species' differences in their climate domains. c) Analysis of species-level trait relationships with maturation size.

#### 281 MASTIF data and model

The MASTIF model and data summarized here are detailed in Clark et al. (2019) and its extended Supplement 282 (see also Qiu et al. 2021, 2022; Journé et al. 2022). Data are of two types, crop counts on trees and seed traps 283 in mapped inventory plots (MASTIF) (Clark et al., 2019) (Fig. S1). The initial sample size is approximately 12 284 million tree-years from five continents on 898 species and 112 families. The majority of observations (99%) 285 are derived from longitudinal studies, involving repeated observations of all trees on a plot or individual trees. 286 The remaining crop count observations (1%) are collected opportunistically through the iNaturalist project 287 MASTIF (Clark et al., 2019). The number of species observed per plot ranges from 1 to 221 species. The 288 number of species observations is larger for seed trap monitoring (476 species in total, 22,929 tree-year 289 observations on average) than for crop count monitoring (130 species in total, 1,058 tree-year observations 290 on average). Most plots are localized in North America and central Europe (97%), whereas most species 291 observations are coming from South America (54%). On average, 75% of individual tree year observation are 292 coming from the tropics. Additional information is provided in Table S1 and Supplementary Files 1 and 2. For 293 both data types, observations include species, diameter, shade class (ranging from "full sun", class 1, to "full 294 shade", class 5), number of fruiting structures, and an estimate of the fraction of the total crop represented 295 by the count. For crop counts, the data model is beta-binomial, with binomial uncertainty for the counts given 296 crop fraction, and beta uncertainty for crop fraction. The seed traps data additionally include mapped locations 297 of trees and seed traps, which is used to jointly estimate fecundity, dispersal, and, for seeds identified only to 298

genus, species identity. For seed traps, the data model is Poisson for counts given dispersal and species, a
 bivariate Student's t (i.e. 2Dt) redistribution kernel for dispersal (Clark *et al.*, 1999), and a multinomial species
 probability (many seeds are identified only to genus level).

The MASTIF model is a dynamic model for year-to-year and tree-to-tree seed production. The model 302 allows for conditional independence in crop counts and seed traps data through latent states. It estimates 303 maturation state and conditional fecundity (seed production given the individual is mature), which depend 304 on tree size, shading, local climate, and soil conditions. Random effects on individuals and years allow for 305 wide variation between trees and over time. The posterior distribution includes the parameters and latent 306 states presented in Clark et al. (2019), and summarized in Qiu et al. (2022) and Journé et al. (2022). Model 307 fitting was accomplished with Gibbs sampling, a Markov chain Monte Carlo technique based on sampling 308 from conditional distributions. Model structure and methodology are implemented with the R package Mast 309 Inference and Forecasting (mastif, v1.0.1) (Clark et al., 2019). 310

#### **Derivation of tree maturation size from fitted MASTIF model**

Tree maturation size (d<sub>mat</sub>) is derived from an individual-scale model fitted to each species with MASTIF. We 312 define tree maturation size (dmat) as the diameter when a tree is mature and has the capacity to produce 313 enough seed to construct one fruiting structure, f<sub>min</sub>. For species that produce one-seeded fruits (e.g., 314 Quercus, Juglandaceae),  $f_{min} = 1$ . For species that produce cones (e.g., Pinaceae, Cupressaceae), pods 315 (e.g., Fabaceae, Bignoniaceae), or other capsules that house multiple seeds (e.g., Fagus capsules), fmin 316 is the number of seeds contained in that structure. The data and definitions we use to determine  $d_{mat}$ 317 differ from those employed in previous studies (e.g. Visser et al. 2016), as we use both crop count and 318 seed trap observation and not only maturation status. The estimation of individual fecundities, obtained 319 through MASTIF model, is also included, taking into account tree characteristics and environment (Clark 320 et al., 2021; Qiu et al., 2021; Journé et al., 2022). MASTIF models the effects of environmental predictors on 321 conditional fecundity (given mature status), because immature trees do not respond to predictors (it is always 322 zero). [Modeling environmental effects on (unconditional) fecundity would make no more sense than including 323 immature individuals in studies of masting intervals or synchronicity.] Conditional fecundity  $\psi$  is represented 324 by a log-normal distribution, which allows for the effects of the environment. The log-normal is undefined 325 for zero seeds. Zeros are accommodated by the fact that trees can be in the immature state ( $\rho = 0$ ), or 326 conditional fecundity can be below the threshold  $f_{min}$ , as in a failed seed crop (Clark *et al.*, 2004, 2019), 327

$$f = \begin{cases} \psi & \rho = 1\\ 0 & (\rho = 0) + (\rho = 1)(\psi < f_{min}) \end{cases}$$
(2)

An individual is immature until the first time fecundity rises above the threshold for producing fruit, i.e., ( $\rho_{i,t} = 1$ )( $\psi_{i,t} > f_{min}$ ). Specifically for tree *i* in year *t*,

$$f_{i,t} = \psi_{i,t} \times \rho_{i,t}$$

$$\rho_{i,t} | \rho_{i,t-1}, \rho_{i,t+1} \sim Bernoulli\left(\rho_{i,t-1} + (1 - \rho_{i,t-1})\rho_{i,t+1}\Phi(\beta_0^\rho + \beta_1^\rho d_{i,t})\right)$$

$$\log \psi_{i,t} \sim N(\mathbf{x}'_{i,t}\boldsymbol{\beta}^x + \dots, \sigma^2)$$
(3)

where  $\Phi(\cdot)$  is the standard normal cumulative distribution function for the probit probability of transitioning to 330 the mature state, depending on tree diameter  $d_{i,t}$ . Importantly,  $\Phi(\beta_0^{\rho} + \beta_1^{\rho} d_{i,t})$  it is the probability of making 331 the transition for an individual that is now in the immature state. For this reason, the coefficients  $\beta_0^{\rho}, \beta_1^{\rho}$  for 332 maturation in the second line of Eq. (3) engage only for the transition tree-years,  $[\rho_{i,t}|\rho_{i,t-1}=0,\rho_{i,t+1}=1]$ . 333 Predictors in the design vector for conditional fecundity  $\mathbf{x}'_{i,t}$  include the mean climate variables (defined at 334 the species level) tested here and competition by neighbors, and  $\beta^x$  is the estimated parameter vector. The 335 ellipses (...) in Eq. (3) includes individual effects (subscript i) and year effects (subscript t) (Clark et al., 336 2019). The variance not assigned to predictors is  $s^2 = \sigma^2 + Var(individuals) + Var(years)$ . 337

Setting all other fitted variables at their mean values and intermediate shade (shade class 3 on the scale from 1 to 5), we obtained (unconditional) fecundity f from the fitted model. We first factored the joint distribution of conditional fecundity and maturation,

$$[\psi > f_{min}, \rho = 1] = [\psi > f_{min}|\rho = 1][\rho = 1]$$
(4)

<sup>341</sup> Using Bayes' theorem, the cumulative distribution function for maturation diameter is

$$[d_{matr} > d|\psi > f_{min}, \rho = 1] \propto [\psi > f_{min}, \rho = 1|d][d]$$
  
=  $[\psi > f_{min}|\rho = 1, d][\rho = 1][d]$   
=  $\Phi(z_1) \Phi(z_2) [d]$  (5)

where  $z_1 = \frac{\log f_{min} - \mathbf{x}' \boldsymbol{\beta}^{\psi}}{s}$  (log normal fecundity),  $z_2 = \beta_0^{\rho} + \beta_0^{\rho} d$  (probit maturation), and again,  $s^2$  is the marginal variance for conditional fecundity. We have taken the diameter distribution [d] to be uniform. The distribution of maturation size is obtained using inverse distribution sampling from Eq. (5), and we estimated  $d_{mat}$  as the mean of this distribution. We selected species for which maturation and fecundity schedules could be estimated with confidence. The selection was based on estimates of maturation status from the MASTIF model, and we retained species with at least 10 immature and 10 mature individuals. This included 486 species observed over a range of values for diameters.

#### **Trait and climate data**

Like maximum tree height or age, maximum tree diameter is a useful concept, despite the fact that it cannot be known. To incorporate the concept of size differences, we use extreme sizes available from literature and our inventory data, recognizing that the concept of a maximum becomes most meaningful with large sample sizes, which are not available for all species. For the final analysis here, we kept the highest estimates

of d<sub>max</sub>. Sources in Table S3 include tropical species from large plots in central Panama (189 species) 354 and French Guyana (33 species), which together represent 45.7% of values. Estimates extracted from the 355 internet (e.g. encyclopedia, online flora) include 173 species (35.6%) (Table S3). For species not estimated 356 in other sources, we used forest inventory data, evaluated by two approaches, both based on order statistics. 357 We avoided using the absolute largest reported value in forest/MASTIF inventories due to the high noise 358 levels associated with extremes. Order statistics were preferred over quantiles, the latter being determined 359 by whether there are huge numbers of small trees in the data set; guantiles are based on the entire stand 360 structure, whereas here the goal is to estimate the largest sizes, regardless of whether there are few or 361 many small trees. For species present in national forest inventories we estimated  $d_{max}$  following Qiu et al. 362 (2021) by using the tenth largest order statistic (38 species, 7.8%). For the remaining species present in 363 MASTIF inventories, and with at least more than 90 unique individuals, we used the fifth largest order statistics 364 (representing in total 8.4%). For species having only maximal plant height (Liu et al., 2019), but no d<sub>max</sub>, 365 we converted them to  $d_{max}$  using allometric equations of Feldpausch et al. (2011) (12 species, <2.5%). 366 Observations of  $d_{max}$  coming from the internet are usually higher than data from National Forest Inventories, 367 allometric predictions, and MASTIF inventories (Fig. S2). Seed size estimates came from measurements in 368 our lab (Clark et al., 2021), the primary literature, and the TRY Plant Trait Database (Kattge et al., 2011). 369 Wood density and SLA are from the compilation of Carmona et al. (2021). We used genus- or family-level 370 means for seed size, SLA, and wood density values that were missing at the species level (15%, 28%, and 371 26%, respectively). We defined a species' seed productivity as (mass per seed)  $\times$  (mean seeds per tree 372 basal area) (Qiu et al., 2022). 373

For species' climate, we extracted average temperature (in  $^{\circ}$  C) and moisture deficit (evapotranspiration minus precipitation, in mm) for each species based on all occurrences in the Global Biodiversity Information Facility (GBIF) through the R package rgbif (Chamberlain & Boettiger, 2017). The GBIF request is available from reference GBIF.org (2022). For species that are absent from GBIF, we extracted temperature and deficit from the MASTIF sites where those species were reported (162 species, 33%). Climate variables were obtained from CHELSA (Karger *et al.*, 2017).

#### Maturation and maximum size

To test the alternative hypotheses that the maturation diameter decreases ( $\beta_d < 1$ ) or increases ( $\beta_d > 1$ ) with maximum species size (Fig. 2b), we estimated parameters in Eq. (1) with the model

$$\log_{10}(d_{mat_s}) = \log_{10}(\alpha) + \beta_d \times \log_{10}(d_{max_s}) + \dots + \epsilon_s$$
  
$$\epsilon \sim N(0, \sigma^2)$$
(6)

for species *s*, where the ellipsis includes climatic variables (moisture deficit and temperature) and their interactions with  $d_{max}$ . We tested alternative models including independence between maturation and maximum size (fitted  $\alpha$  with  $\beta_d$  fixed at zero), proportionate increase (fitted  $\alpha$  with  $\beta_d = 1$ ), and changing relationship with size (both  $\alpha$  and  $\beta_d$  estimated). Models were fitted with regression by using species average estimates of  $d_{mat}$  as a response, and we included the inverse of the standard error of  $d_{mat}$  as weights. Model selection and fit were evaluated with AIC and root-mean-square error (RMSE). Regression dilution could cause underestimation of the strength between here  $d_{mat}$  and  $d_{max}$  when a predictor (i.e.  $d_{max}$ ) contains errors (Frost & Thompson, 2000; Detto *et al.*, 2019). We thus ran additional analyses to test the robustness of our results to the regression dilution effect (see Supplementary material A.2). First, we corrected the parameter  $\hat{\beta}_d$  from measurement error by using the R package mecor (Nab, 2021) (v1.0). Secondly, we tested if the relationship between  $d_{mat}$  and  $d_{max}$  varies depending on the origin of  $d_{max}$ .

#### **Joint trait analysis**

We evaluated the association between maturation size and other species' traits from the ability of  $d_{mat}$  to pre-395 dict other trait values while allowing for climate and phylogeny effects (Fig. 2c). The marginal correlations that 396 are commonly used for this purpose do not account for the many ways that traits can be related to one another. 397 For instance, maturation size might be associated with maximum size because both tend to be high in warm 398 climates, or in the phylogenetic groups that tend to occur in warm climates. To accommodate co-dependence 399 between trait values we used Generalized Joint Attribute Modeling (GJAM) with traits as responses (Clark 400 2016). To account for phylogeny in the joint traits model, we diverged from traditional assumptions concerning 401 residual covariance. Instead, we adopted a direct inference of the effects of phylogenetic groups. Traditional 402 approaches of phylogenetic correction build on highly specific assumptions for the residual variance (random 403 walk, or more complex models representing stabilizing selection such as the Ornstein-Uhlenbeck model). 404 Our departure from these assumptions stems from the recognition that natural selection does not operate 405 uniformly, neither within a given species pair nor across a broad spectrum of species. Our GJAM analysis 406 explored phylogenetic contributions, with species groups treated as random effects and covariance that is 407 unconstrained by assumptions on divergence rates (Qiu et al., 2023). Explanatory variables included temper-408 ature, moisture deficit, and their interaction. Traits included wood density (g  $m^{-3}$ ), specific leaf area (SLA) 409 (mm<sup>2</sup> mg<sup>-1</sup>), species seed productivity (kg m<sup>-2</sup> basal area), seed size (g), maximum diameter ( $d_{max}$ ) (cm), 410 and maturation diameter ( $d_{mat}$ ) (cm). All traits were log-transformed. We included a random phylogenetic 411 group effect in the joint trait analysis (Qiu et al., 2022; Bogdziewicz et al., 2023; Qiu et al., 2023). For species 412 in speciose genera (more than 10 species), genus was used as the phylogenetic group. For species in less 413 speciose genera but belonging to families with more than five species, family was used as the phylogenetic 414 group. For the remaining species (<25% of the total), an 'other' category was used. To estimate the direct 415 effect of traits (i.e. SLA, wood density, species seed productivity, seed size) and climatic variables on  $d_{mat}$ , we 416 report conditional parameters from GJAM. Conditional parameters are estimated by extracting the parameters 417 of the conditional distribution of traits conditioned on  $d_{mat}$ . Conditional parameters estimate the direct associ-418 ations between traits while accounting for climate and phylogeny. Conditional parameters were obtained with 419 the gjam R package (v2.6.2) (Supplementary Material, Section A.1). 420

#### <sup>421</sup> Relation of $d_{mat}$ along the phylogeny

We visualized how  $d_{rel}$  varies across species phylogeny by making a phylogenetic tree plot. We used the 422 phylogeny from Zanne et al. (2014), and retrieved phylogenetic information for 400 out of the 486 studied 423 species. Of the species missing from the phylogeny (i.e. 86 species), the relative proportion of missing 424 phylogenetic information is about 13.2% for temperate species and about 19.4% for tropical species. We then 425 tested for a phylogenetic signal in  $d_{rel}$  and  $d_{mat}$  using Pagel's  $\lambda$  (Pagel, 1999) (which test for a Brownian 426 motion evolutionary signal), with values close to 0 indicating low phylogenetic signal and values close to 1 427 suggesting a phylogenetic correlation. We plotted the phylogenetic tree with ggtree R package (v3.8) (Yu 428 et al., 2017). We estimated the Pagel's  $\lambda$  by using the phylosig function from phytools (v1.5) (Revell, 2012). 429

Table 1: Coefficient estimates and fit to Eq. (6). The selected model with the lowest AIC (bold font at top) includes temperature ( $\beta_T$ ) and the interaction between  $d_{max}$  and temperature ( $\beta_{dT}$ ). The proportional cost model has  $\beta_d$  fixed at 1. The independence model has  $\beta_d$  fixed at 0. Additional models that include moisture deficit and temperature have higher AIC values (Table S4).

α	$\beta_d$	$\beta_T$	$\beta_{dT}$	$\sigma$	AIC	RMSE
3.71 [1.94, 7.07]	0.30 [0.15, 0.46]	-0.023 [-0.035, -0.011]	0.012 [0.0058, 0.019]	0.089	-62	10.2
1.08 [0.93, 1.25]	0.59 [0.55, 0.63]	-	-	0.090	-52	10.0
0.24 [0.23 , 0.26]	1	-	-	0.12	248	18.8
9.25 [8.69, 9.85]	0	-	-	0.15	447	15.4

## 430 **Results**

Maturation size is associated with maximum size, but not proportionately so (Fig. 3a). Large inter-specific 431 variation in  $d_{mat}$  estimates had 95% quantiles that ranged from 4.0 to 51 cm, with relative maturation size 432  $(d_{rel} = d_{mat}/d_{max})$  quantiles of (0.07, 0.65). Contrary to the baseline independence model ( $\beta_d = 0$ ), trees 433 did not start to reproduce at a constant size (dashed black line in Fig. 3a). If we force proportionality (fix 434  $\beta_d$  at 1), the estimate of  $\hat{\alpha} = 0.24(0.23, 0.26)$  (line 3 of Table 1) is consistent with Loehle's (1988) range for 435 hardwoods (1/5 to 1/4), but far outside his range for conifers (1/15 to 1/10). The 95% CI that is well below 0.5. 436 This differs from the Visser et al. (2016)'s estimate of 1/2 for Barro Colorado Island (N = 60 species), Panama, 437 and with Minor & Kobe (2019) La Selva, Costa Rica (N = 16 species). It is crucial to acknowledge that the 438 aforementioned authors employed a distinct definition of  $d_{mat}$  and estimated larger  $d_{mat}$  (Fig. S3). Moreover, 439 this proportional cost model ( $\beta_d = 1$ ) fits poorly, with twice the RMSE and a higher AIC than the best-fitting 440 model (Table 1). 441

Fitting both  $\alpha$  and  $\beta_d$  (line 2 of Table 1) shows strong support for the diminishing risk model ( $0 < \hat{\beta}_d < 1$ ). Allowing for environmental predictors further decreases the estimate to  $\hat{\beta}_d = 0.30$  (0.15, 0.46). The exponent  $0 < \beta_d < 1$  means that relative size at maturation ( $d_{rel}$ ) decreases in large species (blue in Fig. 3a).

The best-fitting model (lowest AIC and RMSE) includes a negative effect of temperature (maturation at small size for species most common in cold climates) and a positive interaction between temperature T and  $d_{max}$  (Table 1). This positive interaction means that the relationship between maturation and maximum size

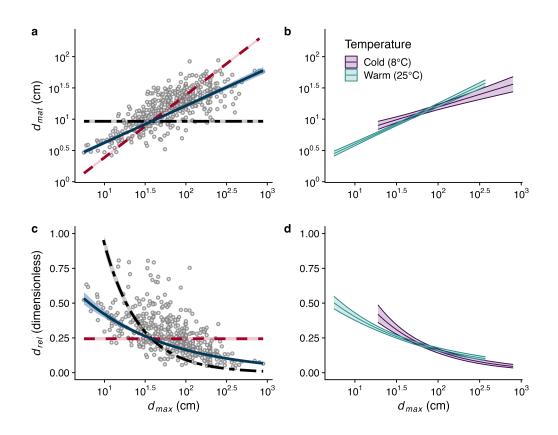


Figure 3: Tree maturation size (a, b), and relative size at maturation (c, d) for 486 species. Each dot represents one species. Alternative models are dashed lines, black for independence between maturation size and maximum size ( $\beta_d = 0$ ), and red for the proportional cost model ( $\beta_d = 1$ ). The best fitting model (blue with 95%CI) supports the diminishing risk model ( $\beta_d < 1$ , Table 1). Panels b and d are predictions from the fitted model with an interaction between continuous  $d_{max}$  and temperature (line 1 of Table 1). This model gives a continuous surface plot of maturation size as a function of maximum size and temperature (see Fig. S4). However, for clarity, we represent only the prediction at cold (8°C, purple) and warm temperatures (25°C, green) spanning observed diameter ranges.

tends to steepen for species in warm climates (Fig. 3b, d), approaching the proportionate risk model (Table 1); the rise in  $d_{mat}$  with  $d_{max}$  increases with temperature. However, the main plus interaction effect remains below 1 even in warm climates showing that the diminishing risk model is supported across this temperature range. The  $\beta_d$  remained below 1 even when we restricted the analysis to a single source of  $d_{max}$  (Table A1 and Figure A1, see Supplementary Section 2). After correcting for risks of regression dilution, the average value of  $\beta_d$  remained below 1, with corrected  $\beta_d = 0.73$ , however, the confidence interval is between 0.03 to 1.43

454 (Table A2, see Supplementary Section 2).

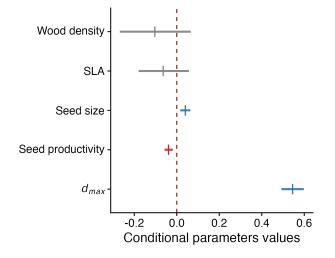


Figure 4: Conditional parameter estimates for the direct effect of traits on tree size at maturation diameter  $(d_{mat})$  while accounting for trait covariance, climate, and phylogeny. Conditional parameters are evaluated on a standardized scale (predictors are centered and standardized) making trait effects on  $d_{mat}$  respective to their variation in the data set. Shown are posterior means and 95% credible intervals. Blue and red represent positive and negative associations where 95% of the posterior does not include zero. SLA = specific leaf area

The joint trait model incorporating random phylogenetic group and climate exhibits a root mean square 455 prediction error 1.17 units smaller compared to the model that includes only climate. Conditional parameter 456 estimates from the joint trait analysis show that  $d_{max}$  has a stronger effect on  $d_{mat}$  than other traits. There is 457 a weak positive association with seed size, and a negative association with species fecundity (see Methods, 458 Trait and climate data section) (Fig. 4). There are no meaningful associations with wood density or SLA. The 459 joint trait analysis also confirms the absence of a direct climate effect on  $d_{mat}$  after accounting for  $d_{max}$  in the 460 conditional trait analysis (see Table S5 for joint trait model and Table S6 for conditional parameters). Previous 461 linear models showed that temperature was significant only in the interaction with  $d_{max}$  (Table S4). Joint trait 462 analysis indicates that the temperature effect on maturation size in Fig. 2c could be due to the abundance of 463 small species (small  $d_{max}$ ) in warm climates (Figure S5). The trait relationships do not depend on the source 464 of climatic data: i) GBIF species occurrence in Fig. 4 or ii) more narrowly, where they occur in the MASTIF 465 data network (Fig. S6). 466 Both maturation ( $d_{mat}$ ) and relative size ( $d_{rel}$ ) show evidence of phylogenetic conservation ( $\lambda_{mat} = 0.83$ , 467

468 p < 0.0001;  $\lambda_{rel} = 0.51$ , p < 0.0001, n = 400, Fig. S7a), yet with substantial variation within some groups.

The two-sample t-test for unequal variances shows differences between gymnosperms and angiosperms. 469 Gymnosperms have high mean values for both  $d_{mat}$  and  $d_{max}$  and low mean values for  $d_{rel}$  (all p < 0.0001). 470 Within gymnosperms, Pinales and Cupressaceae mature at large size, but large  $d_{max}$  gives them lower  $d_{rel}$ 471 than most angiosperms (Fig. S8, S9). Most Pinaceae (Picea, Pinus) and Cupressaceae (Thuja, Sequoia), 472 Fagaceae (Quercus and Fagus), and Juglandaceae have low  $d_{rel}$  (Fig. S10). Plant groups with both tree 473 and shrub habits, such as Rosales, Magnoliales, Rubiaceae, and Fabaceae, have mixed  $d_{rel}$ . However, we 474 did not find a significant effect of tree versus shrub habit on  $d_{rel}$ , possibly due to high variation in the data 475 (Fig. S10). Shrubs may tend to have high  $d_{rel}$ , but higher  $d_{rel}$  is also observed in trees genera like *Magnolia* 476 and Poulsenia. 477

## 478 Discussion

Our analysis suggests a diminishing risk model for the relationship between maturation and maximum size 479 (i.e.  $d_{mat}$  and  $d_{max}$ ). The novelty here comes from the low coupling we find. Indeed, the coefficient  $\hat{\beta}_d = 0.30$ 480 in Table 1 that we found is closer to zero (no relationship) than one. In contrast, the estimated exponent values 481 fitted to vertebrates are greater than 1/2 (Prothero, 1993; Herculano-Houzel, 2019), twice the value of  $\hat{eta}_d$  we 482 find for trees. Nevertheless, the comparison across groups is complex due to the uncertainty on maximum 483 size  $(d_{max})$ , which could influence the value of the exponent  $\hat{\beta}_d$ , but this issue remained unexplored in other 484 taxa. The biological difference of  $\hat{\beta}_d$  may arise because trees differ from other species groups in the gains that 485 come from allocation to growth, as the gain is due to the relative difference in height with other competitive 486 individuals. In most tree species, individuals in the understory produce no seed at all, while dominant stature 487 can yield multi-order-of-magnitude gains in fecundity over crowded neighbors (Clark et al., 2004). In contrast, 488 in vertebrates, improved parental condition and size can translate to incremental increases in clutch size or 489 survival of well-provisioned offspring. Gestation times and physical limits on clutch size (e.g., one offspring) 490 may allow only muted near-term benefits of reproductive delay. 491

In trees, large size comes with uncertainty that could weaken the potential benefit of delaying maturation. 492 Wind exposure and risk of hydraulic failure both increase with size (Bennett et al., 2015; Jackson et al., 493 2021; Gardiner, 2021; Barrere et al., 2023). The vanishing probability that a seed survives to large size, 494 combined with the fact that fecundity can plateau and even decline late in life (Qiu et al., 2021) means that 495 the competitive advantages of extremely large size can rarely make up for lost benefits of early reproduction. 496 At a stand scale, the risk of stand-replacing disturbances can increase with stand age and development (e.g., 497 accumulated fuels increase fire risk), such that species that fail to reach minimum reproductive size before 498 the next disturbance can be excluded from communities (Clark, 1991; McDowell et al., 2020). The fact that 499 maturation size increases with maximum size means that the two are not independent. But the cost is not 500 proportionate (Fig. 3). 501

The fact that some correlation exists does not conflict with a disproportionate importance of near-term gains that can follow delayed maturation. Instead, it suggests that the benefits of large size probably do not come at the end of life. The capacity to reach a large size pays benefits throughout life, contributing with many other variables to current size and fecundity, not just as a tree approaches the maximum.

Both climate and species traits contribute to the relationships between maturation and maximum size. 506 The negative main effect of temperature and its positive interaction steepens the relationship with maximum 507 size in warm climates (Fig. 3), where growth and mortality rates are generally higher than in temperate forests 508 (Stephenson & Van Mantgem, 2005; Locosselli et al., 2020). Abundant resources may offer a disproportionate 509 advantage to early maturation (van Noordwijk & de Jong, 1986; Kozłowski, 1992; Wenk & Falster, 2015). Long 510 growing seasons in warm climates might have similar effects. However, intense competition on nutrient-rich 511 sites might also favor delayed reproduction as trees compete for canopy access. Theoretical studies (Falster 512 et al., 2017; Detto et al., 2022) have shown that a trade-off between maximum size and maturation size can 513 promote niche diversification and maintain species coexistence, and can be typically observed in tropical 514 where there is a wide range of maximum sizes forests (Falster et al., 2017). 515

We did not find that high specific leaf area (SLA) is associated with maturation at small size (Visser 516 et al., 2016) or early age (Wenk et al., 2018). The relationship reported in Wenk et al. (2018) includes leaf 517 area from one year and one site, and the correlation estimated in that study does not appear to control for 518 phylogeny. Similarly, lack of association with wood density in our study does not agree with suggestions that 519 shade-tolerant species with high wood density mature at small size (Thomas et al., 2015). The inclusion of a 520 wider range of plant species may reveal a different pattern of traits. For example, a comparison over a large 521 number of perennial plant species such as herbs, graminoids, shrubs, and trees shows that traits that promote 522 longevity are associated with greater variability in seed production (Journé et al., 2023). The fact that species 523 that produce large seeds also allocate more to reproductive effort (Qiu et al., 2022) could contribute in a small 524 way to delayed maturation. The differences between our result and previous work may be due to the larger 525 species coverage, and to the control of the effect of climate and phylogeny in our joint analysis. 526

Results highlight the importance of large data sets and how they are modelled. This first compilation of 527 tree maturation size for hundreds of species on five continents shows strong support for a diminishing risk 528 model -trees that can get big can still mature at relatively small sizes. The result is a decline in the relative 529 size of maturation for large trees (Fig. 3d). The benefits of extensive data here parallel the shift from early 530 theory that argued for a constant relative maturation size  $(d_{rel})$  in fish (Charnov & Berrigan, 1990), followed 531 by studies showing an exponent that is less than one (Froese & Binohlan, 2000; Tsikliras & Stergiou, 2014; 532 Thorson et al., 2017). It is, however, important to acknowledge that our coverage of tree species diversity 533 is still patchy, with most data coming from Europe and North America and limited spatial coverage in Africa, 534 South America, Asia, and Oceania as data are concentrated in a few large plots in these areas (Daru & 535 Rodriguez, 2023). 536

New insight from this analysis comes first from extending observations beyond a small number of tropical 537 sites, few species, or limited sample size (Thomas, 1996; Wright et al., 2005; Thomas, 2011; Visser et al., 538 2016; Minor & Kobe, 2019). The expanded coverage of species and sites permitted the incorporation of 539 climatic drivers into the analysis of  $d_{mat}$  in relation to  $d_{max}$ , which influenced the estimation of  $\alpha$  and  $\beta_d$ . 540 Secondly, this study also benefited from accommodating detection and temporal dependence to infer mat-541 uration. The estimation of  $d_{mat}$  was possible by combining diverse datasets, either based from direct crop 542 measurement and seed trap monitoring and by the use of MASTIF model which could estimate jointly a prob-543 ability of maturation and individual fecundities. For instance, estimates of d<sub>mat</sub> from Visser et al. (2016) are, 544 545 on average, approximately 1.8 times larger than our estimates for the species in common in the two studies (Fig. S3). This discrepancy could be attributed to a different definition of size at maturation and methods of
 analysis, as our method also includes the number of seeds produced.

Due to the high juvenile mortality, the maturation sizes quantified here are expected to impact predictions 548 from demographic vegetation models, including earth system models (ESMs) that include effects of maturation 549 size. In one ESM study that considered the effects of maturation height, variation in a single value applied 550 to all species did not have a large impact on simulated stand productivity (Raczka et al., 2018). However, 551 when differences in species maturation size are accounted for in models, the effect can be larger. Few 552 individuals survive to large size and, thus, their ability to reproduce early can be important. The fact that 553 species capable of large size tend to retain this capacity to reproduce while still small highlights the importance 554 of understanding maturation size. Accurate estimation of maturation size (d<sub>mat</sub>) is likewise important for 555 assessing response to disturbance regimes, especially as the time to maturity begins to exceed the interval 556 between disturbances. For instance, several species that compared pairs of species found that species with 557 smaller size at maturation can have better post-disturbance dynamics than species with larger maturation size 558 (Alfaro-Sánchez et al., 2022; Andrus et al., 2020). Our results provided the data to test such hypotheses at 559 a much larger scale. This effect can even scale up at the ecosystem scale. In boreal habitats, exposure to 560 more frequent disturbances that exceed the tree maturation time can completely change a tree community to 561 a grass-dominated community without a return to a forest stand within centuries (Buma et al., 2013). 562

Current ESMs suffer from limited information on allocation to reproduction (Wenk & Falster, 2015), in-563 cluding empirical data (Hanbury-Brown et al., 2022b). Similarly, management actions intended to assure 564 regeneration from seed also need to consider if the minimum harvest diameter is smaller than maturation 565 size (Ouédraogo et al., 2018). Maturation size may play an important role in the ability of species to respond 566 to disturbance and climate change (McDowell et al., 2020) when tree maturation can be reached faster un-567 der elevated CO<sub>2</sub> exposure (LaDeau & Clark, 2001). Considering the difference in maturation size between 568 species may be crucial, although it can be more challenging to comprehend due to the impact of  $CO_2$  on 569 maturation size. Understanding how fecundity strategies differ between species and phylogenetic groups, 570 such as lower relative size at maturation  $(d_{rel})$  for gymnosperms than angiosperms, may open a new avenue 571 to better understand species diversification and responses to disturbances (Bond, 1989; Verdu, 2002; Qiu 572 et al., 2022). Developing a model that represents the size at which a species begins to produce seeds could 573 improve the representation of the regeneration of each functional type (Hanbury-Brown et al., 2022a) and 574 colonization rates (Snell, 2014) and improve our understanding of species coexistence. Our study contributes 575 to the maturation sizes needed for each of these objectives. 576

### 577 References

Alfaro-Sánchez, R., Johnstone, J.F., Cumming, S.G., Day, N.J., Mack, M.C., Walker, X.J. *et al.* (2022). What
 Drives Reproductive Maturity and Efficiency in Serotinous Boreal Conifers? *Frontiers in Ecology and Evolution*, 10, 1–12.

Andrus, R.A., Harvey, B.J., Hoffman, A. & Veblen, T.T. (2020). Reproductive maturity and cone abundance vary with tree size and stand basal area for two widely distributed conifers. *Ecosphere*, 11.

Barrere, J., Reineking, B., Cordonnier, T., Kulha, N., Honkaniemi, J., Peltoniemi, M. *et al.* (2023). Func tional traits and climate drive interspecific differences in disturbance-induced tree mortality. *Global Change Biology*, 29, 2836–2851.

Bennett, A.C., McDowell, N.G., Allen, C.D. & Anderson-Teixeira, K.J. (2015). Larger trees suffer most during
 drought in forests worldwide. *Nature Plants*, 1, 15139.

Bogdziewicz, M., Acuña, M.C.A., Andrus, R., Ascoli, D., Bergeron, Y., Brveiller, D. *et al.* (2023). Linking seed
 size and number to trait syndromes in trees. *Global Ecology and Biogeography*, 32, 683–694.

Bond, W.J. (1989). The tortoise and the hare: ecology of angiosperm dominance and gymnosperm persistence. *Biological Journal of the Linnean Society*, 36, 227–249.

<sup>592</sup> Buma, B., Brown, C.D., Donato, D.C., Fontaine, J.B. & Johnstone, J.F. (2013). The impacts of changing <sup>593</sup> disturbance regimes on serotinous plant populations and communities. *BioScience*, 63, 866–876.

Caignard, T., Kremer, A., Bouteiller, X.P., Parmentier, J., Louvet, J.M., Venner, S. *et al.* (2021). Counter gradient variation of reproductive effort in a widely distributed temperate oak. *Functional Ecology*, 35,
 1745–1755.

Caignard, T., Kremer, A., Firmat, C., Nicolas, M., Venner, S. & Delzon, S. (2017). Increasing Spring Temper atures Favor Oak Seed Production in Temperate Areas. *Scientific Reports*, 7, 1–8.

- Carmona, C.P., Bueno, C.G., Toussaint, A., Träger, S., Díaz, S., Moora, M. *et al.* (2021). Fine-root traits in the
   global spectrum of plant form and function. *Nature*, 597, 683–687.
- <sup>601</sup> Chamberlain, S. & Boettiger, C. (2017). R python, and ruby clients for gbif species occurrence data. *PeerJ* <sup>602</sup> *PrePrints*.
- <sup>603</sup> Charlesworth, B. (2000). Fisher, medawar, hamilton and the evolution of aging. *Genetics*, 156, 927–931.

<sup>604</sup> Charnov, E.L. & Berrigan, D. (1990). Age of Maturity Versus the Adult Lifespan. *Evolutionary Ecology*, 4,
 <sup>605</sup> 273–275.

<sup>606</sup> Charnov, E.L. & Berrigan, D. (1991). Evolution of life history parameters in animals with indeterminate growth,
 <sup>607</sup> particularly fish. *Evolutionary Ecology*, 5, 63–68.

- <sup>608</sup> Clark, J.S. (1991). Disturbance and tree life history on the shifting mosaic landscape. *Ecology*, 72, 1102– <sup>609</sup> 1118.
- Clark, J.S. (2016). Why species tell more about traits than traits about species: Predictive analysis. *Ecology*,
   97, 1979–1993.
- Clark, J.S., Andrus, R., Aubry-Kientz, M., Bergeron, Y., Bogdziewicz, M., Bragg, D.C. *et al.* (2021). Continent wide tree fecundity driven by indirect climate effects. *Nature Communications*, 12, 1–11.
- Clark, J.S., Bell, D.M., Kwit, M.C. & Zhu, K. (2014). Competition-interaction landscapes for the joint response
   of forests to climate change. *Global Change Biology*, 20, 1979–91.
- Clark, J.S., LaDeau, S. & Ibanez, I. (2004). Fecundity of trees and the colonization–competition hypothesis.
   *Ecological Monographs*, 74, 415–442.
- Clark, J.S., Nuñez, C.L. & Tomasek, B. (2019). Foodwebs based on unreliable foundations: spatiotemporal
   masting merged with consumer movement, storage, and diet. *Ecological Monographs*, 89, 1–24.
- Clark, J.S., Silman, M., Kern, R., Macklin, E. & HilleRisLambers, J. (1999). Seed dispersal near and far:
   Patterns across temperate and tropical forests. *Ecology*, 80, 1475–1494.
- Daru, B.H. & Rodriguez, J. (2023). Mass production of unvouchered records fails to represent global biodi versity patterns. *Nature Ecology and Evolution*, 7, 816–831.
- Davies, S.J. & Ashton, P.S. (1999). Phenology and fecundity in pioneer species of Euphorbiaceae. *American Jornal of Botany*, 86, 1786–1795.
- Detto, M., Levine, J.M. & Pacala, S.W. (2022). Maintenance of high diversity in mechanistic forest dynamics
   models of competition for light. *Ecological Monographs*, 92.
- <sup>628</sup> Detto, M., Visser, M.D., Wright, S.J. & Pacala, S.W. (2019). Bias in the detection of negative density depen-<sup>629</sup> dence in plant communities. *Ecology Letters*, 22, 1923–1939.
- Dietze, M.C. & Clark, J.S. (2008). Changing the gap dynamics paradigm: Vegetative regeneration control on
   forest response to disturbance. *Ecological Monographs*, 78, 331–347.
- Falster, D.S., Brännström, Å., Westoby, M. & Dieckmann, U. (2017). Multitrait successional forest dynamics
   enable diverse competitive coexistence. *Proceedings of the National Academy of Sciences of the United States of America*, 114, E2719–E2728.
- Falster, D.S. & Westoby, M. (2003). Plant height and evolutionary games. *Trends in Ecology and Evolution*,
   18, 337–343.
- Feldpausch, T.R., Banin, L., Phillips, O.L., Baker, T.R., Lewis, S.L., Quesada, C.A. *et al.* (2011). Height diameter allometry of tropical forest trees. *Biogeosciences*, 8, 1081–1106.

Froese, R. & Binohlan, C. (2000). Empirical relationships to estimate asymptotic length, length at first maturity
 and length at maximum yield per recruit in fishes, with a simple method to evaluate length frequency data.
 *Journal of Fish Biology*, 56, 758–773.

Frost, C. & Thompson, S.G. (2000). Correcting for regression dilution bias: Comparison of methods for
a single predictor variable. *Journal of the Royal Statistical Society. Series A: Statistics in Society*, 163,
173–189.

Gardiner, B. (2021). Wind damage to forests and trees: a review with an emphasis on planted and managed
 forests. *Journal of Forest Research*, 26, 248–266.

GBIF.org (2022). Occurrence download https://doi.org/10.15468/dl.wevh3v.

Hanbury-Brown, A.R., Powell, T.L., Muller-Landau, H.C., Wright, S.J. & Kueppers, L.M. (2022a). Simulating
 environmentally-sensitive tree recruitment in vegetation demographic models. *New Phytologist*, 235, 78–
 93.

Hanbury-Brown, A.R., Ward, R.E. & Kueppers, L.M. (2022b). Forest regeneration within Earth system models:
 current process representations and ways forward. *New Phytologist*, 235, 20–40.

Herculano-Houzel, S. (2019). Longevity and sexual maturity vary across species with number of cortical
 neurons, and humans are no exception. *Journal of Comparative Neurology*, 527, 1689–1705.

Jackson, T.D., Shenkin, A.F., Majalap, N., Bin Jami, J., Bin Sailim, A., Reynolds, G. *et al.* (2021). The mechanical stability of the world's tallest broadleaf trees. *Biotropica*, 53, 110–120.

Jensen, A.L. (1996). Beverton and Holt life history invariants result from optimal trade-off of reproduction and survival. 822, 820–822.

Journé, V., Andrus, R., Aravena, M.C., Ascoli, D., Berretti, R., Berveiller, D. *et al.* (2022). Globally, tree fecundity exceeds productivity gradients. *Ecology Letters*, 25, 1471–1482.

Journé, V., Hacket-Pain, A. & Bogdziewicz, M. (2023). Evolution of masting in plants is linked to investment in low tissue mortality. *Nature communications*, 14, 7998.

Karger, D.N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R.W. *et al.* (2017). Climatologies at
 high resolution for the earth's land surface areas. *Scientific Data*, 4, 1–20.

Kattge, J., Díaz, S., Lavorel, S., Prentice, I.C., Leadley, P., Bönisch, G. *et al.* (2011). TRY - a global database
 of plant traits. *Global Change Biology*, 17, 2905–2935.

Koch, G.W., Stillet, S.C., Jennings, G.M. & Davis, S.D. (2004). The limits to tree height. Nature, 428, 851–854.

Kohler, P.K. & Huth, A. (2004). Simulating Growth Dynamics in a South-East Asian Rainforest Threatened By
 Recruitment Shortage and. *Climatic Change*, 67, 95–117.

- Kohyama, T., Suzuki, E., Partomihardjo, T., Yamada, T. & Kubo, T. (2003). Tree species differentiation in
   growth, recruitment and allometry in relation to maximum height in a Bornean mixed dipterocarp forest.
   *Journal of Ecology*, 91, 797–806.
- Kozłowski, J. (1992). Optimal allocation of resources to growth and reproduction: Implications for age and
   size at maturity. *Trends in Ecology and Evolution*, 7, 15–19.
- LaDeau, S.L. & Clark, J.S. (2001). Rising co2 levels and the fecundity of forest trees. Science, 292, 95–8.
- Le Roncé, I., Gavinet, J., Ourcival, J.M., Mouillot, F., Chuine, I. & Limousin, J.M. (2021). Holm oak fecundity does not acclimate to a drier world. *New Phytologist*, 231, 631–645.
- Lines, E.R., Zavala, M.A., Purves, D.W. & Coomes, D.A. (2012). Predictable changes in aboveground allome try of trees along gradients of temperature, aridity and competition. *Global Ecology and Biogeography*, 21,
   1017–1028.
- Liu, H., Gleason, S.M., Hao, G., Hua, L., He, P., Goldstein, G. *et al.* (2019). Hydraulic traits are coordinated with maximum plant height at the global scale. *Science Advances*, 5.
- Locosselli, G.M., Brienen, R.J.W., Leite, M.d.S., Gloor, M., Krottenthaler, S., Oliveira, A.A.d. *et al.* (2020).
   Global tree-ring analysis reveals rapid decrease in tropical tree longevity with temperature. *Proceedings of the National Academy of Sciences*, 117, 33358–33364.
- Loehle, C. (1988). Tree life history strategies: the role of defenses. *Canadian Journal of Forest Research*, 18,
   209–222.
- McDowell, N., Allen, C., Anderson-Teixeira, K., Aukema, B., Bond-Lamberty, B., Chini, L. *et al.* (2020). Pervasive shifts in forest dynamics in a changing world. *Science*, 368, eaaz9463.
- Minor, D.M. & Kobe, R.K. (2019). Fruit production is influenced by tree size and size-asymmetric crowding in
   a wet tropical forest. *Ecology and Evolution*, 9, 1458–1472.
- Moles, A.T., Ackerly, D.D., Tweddle, J.C., Dickie, J.B., Smith, R., Leishman, M.R. *et al.* (2006). Global patterns
   in seed size. *Global Ecology and Biogeography*, 16, 109–116.
- <sup>694</sup> Moles, A.T., Falster, D.S., Leishman, M.R. & Westoby, M. (2004). Small-seeded species produce more seeds <sup>695</sup> per square metre of canopy per year, but not per individual per lifetime. *Journal of Ecology*, 92, 384–396.
- Nab, L. (2021). *mecor: Measurement Error Correction in Linear Models with a Continuous Outcome*. R
   package version 1.0.0.
- <sup>698</sup> Niklas, K.J. (1994). *Plant Allometry. The Scaling of Form and Process.* Chicago.
- van Noordwijk, A. & de Jong, G. (1986). Acquisition and Allocation of Resources: Their Influence on Variation
   in Life History Tactics. *The American Naturalist*, 128, 137–142.

Ouédraogo, D.Y., Doucet, J.L., Daïnou, K., Baya, F., Biwolé, A.B., Bourland, N. *et al.* (2018). The size at reproduction of canopy tree species in central Africa. *Biotropica*, 50, 465–476.

<sup>703</sup> Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature*, 401, 877–884.

Prothero, J. (1993). Adult life span as a function of age at maturity. *Experimental Gerontology*, 28, 529–536.

Qiu, T., Andrus, R., Aravena, M.C., Ascoli, D., Bergeron, Y., Berretti, R. *et al.* (2022). Limits to reproduction and
 seed size-number trade-offs that shape forest dominance and future recovery. *Nature Communications*,
 13.

Qiu, T., Aravena, M.C., Andrus, R., Ascoli, D., Bergeron, Y., Berretti, R. *et al.* (2021). Is there tree senescence?
 The fecundity evidence. *Proceedings of the National Academy of Sciences of the United States of America*, 118, 1–10.

Qiu, T., Aravena, M.C., Ascoli, D., Bergeron, Y., Bogdziewicz, M., Boivin, T. et al. (2023). Masting is uncommon

in trees that depend on mutualist dispersers in the context of global climate and fertility gradients. *Nature* 

- 713 *Plants*, 9, 1044–1056.
- R Core Team (2023). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical
   Computing, Vienna, Austria.

Raczka, B., Dietze, M.C., Serbin, S.P. & Davis, K.J. (2018). What limits predictive certainty of long-term carbon
 uptake? *Journal of Geophysical Research: Biogeosciences*, 123, 3570–3588.

- Revell, L.J. (2012). phytools: An R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3, 217–223.
- Seyednasrollah, B. & Clark, J.S. (2020). Where Resource-Acquisitive Species Are Located: The Role of
   Habitat Heterogeneity. *Geophysical Research Letters*, 47, 1–12.

Sharma, S., Andrus, R., Bergeron, Y., Bogdziewicz, M., Bragg, D.C., Brockway, D. *et al.* (2022). North
 American tree migration paced by climate in the West, lagging in the East. *Proceedings of the National Academy of Sciences of the United States of America*, 119.

- Snell, R.S. (2014). Simulating long-distance seed dispersal in a dynamic vegetation model. *Global Ecology and Biogeography*, 23, 89–98.
- 727 Stearns, S.C. (1989). Trade-Offs in Life-History Evolution. *Functional Ecology*, 3, 259–268.
- Stephenson, N.L. & Van Mantgem, P.J. (2005). Forest turnover rates follow global and regional patterns of
   productivity. *Ecology Letters*, 8, 524–531.
- Thomas, S.C. (1996). Relative size at onset of maturity in rain forest trees: a comparative analysis of 37
   Malaysian species. *Oikos*, 76, 1450154.

- Thomas, S.C. (2011). Age-Related Changes in Tree Growth and Functional Biology: The Role of Reproduc-
- tion. In: Size- and Age-Related Changes in Tree Structure and Function (eds. Meinzer, F.C., Lachenbruch,
- B. & Dawson, T.E.). Springer Netherlands, Dordrecht, vol. 4, pp. 33–64.
- Thomas, S.C., Martin, A.R. & Mycroft, E.E. (2015). Tropical trees in a wind-exposed island ecosystem:
   Height-diameter allometry and size at onset of maturity. *Journal of Ecology*, 103, 594–605.
- Thorson, J.T., Munch, S.B., Cope, J.M. & Gao, J. (2017). Predicting life history parameters for all fishes
   worldwide. *Ecological Applications*, 27, 2262–2276.
- Tsikliras, A.C. & Stergiou, K.I. (2014). Size at maturity of Mediterranean marine fishes. *Reviews in Fish Biology and Fisheries*, 24, 219–268.
- 741 Verdu, M. (2002). Age at maturity and diversification in woody angiosperms '. Evolution, 56, 1352–1361.

Visser, M.D., Bruijning, M., Wright, S.J., Muller-Landau, H.C., Jongejans, E., Comita, L.S. *et al.* (2016).
 Functional traits as predictors of vital rates across the life cycle of tropical trees. *Functional Ecology*, 30, 168–180.

- Wallentin, G., Tappeiner, U., Strobl, J. & Tasser, E. (2008). Understanding alpine tree line dynamics: An
   individual-based model. *Ecological Modelling*, 218, 235–246.
- Wenk, E.H., Abramowicz, K., Westoby, M. & Falster, D.S. (2018). Investment in reproduction for 14 iteroparous
   perennials is large and associated with other life-history and functional traits. *Journal of Ecology*, 106, 1338–1348.
- Wenk, E.H. & Falster, D.S. (2015). Quantifying and understanding reproductive allocation schedules in plants.
   *Ecology and Evolution*, 5, 5521–5538.
- Westoby, M., Falster, D.S., Moles, A.T., Vesk, P.A. & Wright, I.J. (2002). Plant ecological strategies: Some
   leading dimensions of variation between species. *Annual Review of Ecology and Systematics*, 33, 125–
   159.
- Wright, S.J., Jaramillo, M.A., Pavon, J., Condit, R., Hubbell, S.P. & Foster, R.B. (2005). Reproductive size
   thresholds in tropical trees: Variation among individuals, species and forests. *Journal of Tropical Ecology*,
   21, 307–315.
- Yang, X., Angert, A.L., Zuidema, P.A., He, F., Huang, S., Li, S. *et al.* (2022). The role of demographic compensation in stabilising marginal tree populations in North America. *Ecology Letters*, 25, 1676–1689.
- Yu, G., Smith, D.K., Zhu, H., Guan, Y. & Lam, T.T.Y. (2017). Ggtree: an R Package for Visualization and
   Annotation of Phylogenetic Trees With Their Covariates and Other Associated Data. *Methods in Ecology and Evolution*, 8, 28–36.
- Zanne, A.E., Tank, D.C., Cornwell, W.K., Eastman, J.M., Smith, S.A., FitzJohn, R.G. *et al.* (2014). Three keys
   to the radiation of angiosperms into freezing environments. *Nature*, 506, 89–92.

## 765 Acknowledgements

We thank the National Ecological Observatory Network (NEON) for access to sites and vegetation struc-766 ture data. The project has been funded by grants to JSC from the National Science Foundation, most 767 recently DEB-1754443, and by the Belmont Forum (1854976), NASA (AIST16-0052, AIST18-0063), and 768 the Programme d'Investissement d'Avenir under project FORBIC (18-MPGA-0004) (Make Our Planet Great 769 Again). Jerry Franklin's data remain accessible through NSF LTER DEB-1440409. Puerto Rico data were 770 funded by NSF grants, most recently, DEB 0963447 and LTREB 11222325. Data from the Andes Biodiversity 771 and Ecosystem Research Group were funded by the Gordon and Betty Moore Foundation and NSF LTREB 772 1754647. VJ was supported by project FORBIC (18-MPGA-0004) and project No. 2021/43/P/NZ8/01209 co-773 funded by the Polish National Science Centre and the EU H2020 research and innovation programme under 774 the MSCA GA No. 945339. For the purpose of Open Access, the author has applied a CC-BY public copyright 775 licence to any Author Accepted Manuscript (AAM) version arising from this submission. MB was supported 776 by grant no. 2019/35/D/NZ8/00050 from the Polish National Science Centre, and Polish National Agency for 777 Academic Exchange Bekker programme PPN/BEK/2020/1/00009/U/00001. Research by the USDA Forest 778 Service and the USGS was funded by these agencies. Any use of trade, firm, or product names is for de-779 scriptive purposes only and does not imply endorsement by the U.S. Government. 780

781

#### 782 Competing interests

783 The authors declare no competing interests

784

#### 785 Supplementary Materials

- 786 Section A1 A2
- 787 Table S1 S6
- 788 Fig S1 S9

789

## Supplementary material

#### 791 A.1 Conditional parameters

One way to evaluate relationships between traits is to ask how well a trait like  $d_{mat}$  predicts other traits, while controlling for effects of climate and phylogeny. We start with the joint distribution of M traits for each species s = 1, ..., S fitted with GJAM (Methods). All traits were log-transformed. The joint distribution is

$$\begin{aligned} [\mathbf{T}_{s}|P,X] &= MVN_{M}(\mathbf{T}_{s}|\mathbf{B}'\mathbf{x}_{s} + \mathbf{g}[s],\Sigma) \\ \mathbf{g} &\sim MVN(\mathbf{0},\Omega) \end{aligned} \tag{A1}$$

where MVN is the multivariate normal distribution,  $\mathbf{g}[s]$  is a random vector for the phylogenetic group to which s belongs, and  $\Omega$  is the  $M \times M$  covariance between traits taken over phylogenetic groups (Clark *et al.*, 2016). With this fitted model, we consider the effects of  $d_{mat}$  on all other traits, organized in the vector  $\mathbf{T} = [\mathbf{u}, \mathbf{d}]$ , where  $\mathbf{d}$  is the length-S vector of maturation sizes, and  $\mathbf{u}$  is a  $S \times M - 1$  matrix holding all traits in  $\mathbf{T}$  other than  $d_{mat}$ .

<sup>800</sup> We partition the coefficients in **B** and trait covariance  $\Sigma$  as

$$\mathbf{B} = \begin{bmatrix} \mathbf{B}_u \\ \mathbf{B}_d \end{bmatrix}, \Sigma = \begin{bmatrix} \Sigma_{u,u} & \Sigma_{u,d} \\ \Sigma_{d,u} & \Sigma_{d,d} \end{bmatrix}$$
(A2)

For *M* traits and *Q* climate predictors in  $\mathbf{x}_s$ ,  $\mathbf{B}_u$  is the  $Q \times M - 1$  matrix of climate effects on traits other than  $d_{mat}$ ,  $\mathbf{B}_d$  is the  $Q \times 1$  vector of climate effects on  $d_{mat}$ , with similar partition of  $\Sigma$ . We then write the conditional distribution of responses in **u** as

$$\begin{aligned} \mathbf{u}_{s}|d_{mat,s} &\sim MVN(\mathbf{A}d_{mat,s} + \mathbf{C}\mathbf{x}_{s} + \mathbf{g}[s], \mathbf{P}) \\ \mathbf{A} &= \Sigma_{u,d} \Sigma_{d,d}^{-1} \\ \mathbf{C} &= \mathbf{B}'_{u} - \mathbf{A}\mathbf{B}'_{d} \\ \mathbf{P} &= \Sigma_{u,u} - \mathbf{A}\Sigma_{d,u} \end{aligned}$$
(A3)

**A** is the vector of effects of  $d_{mat}$  on each response in **u**, **C** holds the effects of **x**, and **P** is the conditional residual covariance. Other applications can be found in Qiu *et al.* (2021); Bogdziewicz *et al.* (2023). We report in Fig. 4 the estimates from **A**.

#### <sup>807</sup> A.2 Impact of error on estimates of maximum size ( $d_{max}$ )

## A.2.1 Robustness of the model relating maturation size to maximum size to data sources

In this additional analysis, we aimed to test the relationship between maturation size  $(d_{mat})$  and maximum 810 size  $(d_{max})$  using different data sources for  $d_{max}$ . The estimates  $d_{max}$  used in the main analysis are based 811 on online open sources and are on average higher than other sources of  $d_{max}$  (Figure S2) which could 812 underestimate the parameter  $\beta_d$ . The analysis was restricted to the most abundant data source of  $d_{max}$ . We 813 conducted separate regression analyses using  $d_{max}$  estimates from unpublished data (J. Wright, N = 233 814 species), National Forest Inventories (N = 158 species), and MASTIF inventories (N = 346 species) based on 815 the model from Equation (6). In the three different models (i.e. one model per data source of  $d_{max}$ ), we used 816 the inverse of the standard error of  $d_{mat}$  as weights in the models. We then extracted coefficient estimates 817 for all three models. Results are reported in Table A1 and visualized in Figure A1. 818

Table A1: Coefficient estimates from the model Eq. (6) according to the origin of  $d_{max}$ . Data sources of  $d_{max}$  are National Forest Inventories (NFI), MASTIF inventories, and unpublished field tropical measurements.

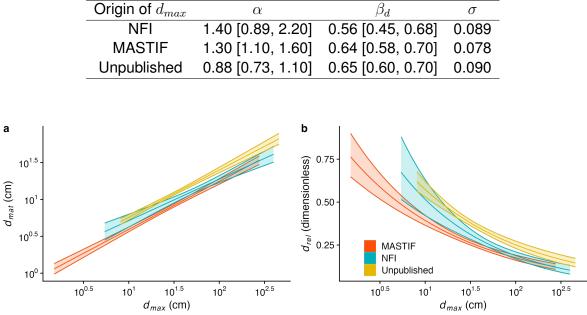


Figure A1: Tree maturation size and relative size at maturation relationship to maximum size according to the origin of maximum size observations. The orange line represents predictions from the regression between  $d_{mat}$  and  $d_{max}$  based on unpublished data (obtained from J. Wright field observation, N = 233 species). In blue, the prediction from the regression between  $d_{mat}$  and  $d_{max}$  comes only from National Forest Inventories estimates (N = 158 species). In yellow, the prediction from the regression between  $d_{max}$  comes only from MASTIF inventories (N = 346 species).

#### **A.2.2** Correction of maximum size error measurement

Measurement error in a covariate is common and often ignored which could have implications in the estimation 820 of the relationship between a response and a covariate (Nab et al., 2021). Indeed, the true value of a covariate, 821 here of  $d_{max}$ , is not available, and using a log-log model exposed to regression dilution could flatten the slope 822  $(\beta_d)$  (Detto *et al.*, 2019). To correct for measurement error we benefit here from the R package mecor (Nab, 823 2021) by using additional observation of our variable  $d_{max}$ . We specified here the error-prone measurement of 824  $d_{max}$ , coming from the highest estimates of  $d_{max}$ . We then used here four different additional observations of 825 dmax, where this covariate can be obtained from Online Open-access resources, National Forest Inventories, 826 Unpublished data, and MASTIF inventories. We ran the analysis on a simple model from Equation (6), that 827 would include only  $d_{max}$ , or both  $d_{max}$  and temperature as covariates. The function does not allow to take into 828 account weights in the analysis. To make a fair comparison, we reported here both coefficients, uncorrected 829 and corrected (Table A2). 830

 Table A2: Coefficient estimates uncorrected and corrected from measurement errors fitted to Eq. (6). Values are reported with a mean estimate and 95% CI.

Model	<b>Coefficient Parameter</b>	Uncorrected value	Corrected value
$d_{max}; T$	α	1.404 [1.092, 1.804]	0.826 [0.022, 31.637]
	$eta_d$	0.58 [0.53, 0.62]	0.732 [-0.049, 1.51]
	$\beta_T$	-0.0005 [-0.0029, 0.0019]	-0.0003 [-0.012, 0.011]
$d_{max}$	α	1.357 [1.132, 1.627]	0.810 [0.048, 13.555]
	$eta_d$	0.582 [0.539, 0.625]	0.733 [0.031, 1.434]

## **Supplementary Tables and Figures**

Table S1: Numbers of species, plots, tree-year observations coming from crop count and seeds traps, and individual species tree year observations according to region. Additional details about MASTIF network are provided in Supplementary Files 1 and 2, and in Table S2.

Region	Nb. of plots	Nb. of species	Crop count x year	Seed-trap x year	Nb. Ind/Year
Africa	2	19	416	9,705	12
Asia	4	59	0	1,589,946	1,475
Eastern North America	155	92	20,983	1,389,998	142
Western North America	280	29	83,042	715,132	576
Europe	156	21	30,763	158,991	28
South America	7	267	2,280	7,050,620	863

832

Table S2: MASTIF plots listed by WWF eco-regions, with principal investigator list and references. The column ST/CC indicates seed traps (ST) and crop counts (CC) sites.

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
A/W Turkey sclerophyllous/mixed						
	PNPG	E. Daskalakou	24	38	CC	Daskalakou <i>et al.</i> (2019)
Alps conifer/mixed forests						
	ROTH	G. Gratzer	15	48	ST	
	BELLE	G. Kunstler; B. Courbaud	6	45	ST CC	
	RNNPT	L. Dormon	7	45	CC	Dormont <i>et al.</i> (2006)
	PNPP	D. Ascoli; R. Motta; R. Berretta; G. Vacchiano	12	46	ST	Hacket-Pain <i>et al.</i> (2019)
Appalachian-Blue Ridge forests						
	BCEF	C. Greenberg	-83	35	CC	Rose <i>et al.</i> (2012)
	CWT	J. S. Clark	-83	35	ST CC	Clark <i>et al.</i> (2004)
	EPENN	M. Steele	-76	41	CC	
	GRAN	C. Greenberg	-82	36	CC	Rose <i>et al.</i> (2012)
	GRSM	J. S. Clark	-83	36	ST	
	MARS	J. S. Clark; S. Pearson	-83	36	ST CC	Clark <i>et al.</i> (2014)
	MLBS	J. S. Clark	-81	37	ST CC	
	PISG	C. Greenberg	-83	35	CC	Rose <i>et al.</i> (2012)
	SCBI	B. McShea	-78	39	ST	Bourg et al. (2013)
Appalachian mixed mesophytic forests						
Arizona Mountains forests						
	MOPA	M. Redmond	-106	33	CC	Redmond et al. (2012)
	REMO	A. Whipple; C. Gering; T. Whitham	-112	36	CC	Whipple <i>et al.</i> (2019)
	SICI	A. Wion; M. Redmond	-108	33	CC	
	WHIT	A. Wion; M. Redmond	-109	33	CC	
	WINO	A. Whipple; C. Gering; T. Whitham	-111	35	CC	Whipple et al. (2019)

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
Atlantic coastal pine barrens						
	BARBEAU	N. Delpierre; D. Berveiller	3	48	ST	
Atlantic mixed forests						
Balkan mixed forests						
Baltic mixed forests						
British Columbia mainland coastal forests						
	GLCR1	J. Franklin	-122	49	CC	
	GLCR2	J. Franklin	-122	49	CC	
	HEME	J. Franklin	-122	49	CC	
	STPA	J. Franklin	-121	47	CC	
California Central Valley grasslands						
California interior chaparral/woodlands						
California montane chaparral/woodlands						
	HNHR	J. Knops; W. Koenig	-122	36	CC	Knops & Koenig (2012)
Carpathian montane forests						
	BGNP	M. Zywiec; L. Piechnik; B. Seget; M. Ledwon	20	50	CC	
Cascade Mountains leeward forests						
	TUCR	J. Franklin	-121	48	CC	
Celtic broadleaf forests						
	BENWE	A. Hacket-Pain	-2	55	CC	Bogdziewicz et al. (2020)
	CONGL	A. Hacket-Pain	-2	53	CC	Bogdziewicz et al. (2020)
	GILLF	A. Hacket-Pain	-2	54	CC	Bogdziewicz et al. (2020)
	HIMLE	A. Hacket-Pain	-2	53	CC	Bogdziewicz et al. (2020)
	KEELE	A. Hacket-Pain	-2	53	CC	Bogdziewicz et al. (2020)
	KILLE	A. Hacket-Pain	-3	51	CC	Bogdziewicz et al. (2020)
	RIPON	A. Hacket-Pain	-1	54	CC	Bogdziewicz et al. (2020)
	SPENN	A. Hacket-Pain	-2	55	CC	Bogdziewicz et al. (2020)

#### Table S2 – continued from previous page

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
	WOODB	A. Hacket-Pain	-3	51	CC	Bogdziewicz et al. (2020)
Central Canadian Shield forests						
	COCH	Y. Bergeron; Y. Messaoud	-81	49	CC	Messaoud et al. (2007)
	LDUPT	Y. Bergeron; Y. Messaoud	-79	48	CC	Messaoud et al. (2007)
	MASK	Y. Bergeron; Y. Messaoud	-79	50	CC	Messaoud et al. (2007)
Central European mixed forests						
Central forest-grasslands transition						
	UKFS	J. S. Clark	-95	39	ST CC	
	WUSL	J. Myers	-91	39	ST	
Central Pacific coastal forests						
	MAPK	J. Franklin	-124	45	CC	
Central Ranges xeric scrub						
	LS	B. Wright	132	-24	CC	Wright & Zuur (2014)
Central tall grasslands						
Central/S Cascades forests						
	BAMT	J. Franklin	-122	46	ST	
	BERK	J. Franklin	-122	43	CC	
	BLLK	J. Franklin	-122	46	CC	
	DECU	J. Franklin	-122	45	CC	
	IRMT	J. Franklin	-122	44	CC	
	MOLK	J. Franklin	-122	46	CC	
	MORA	J. HilleRisLambers	-122	47	ST	
	PEPR	J. Franklin	-122	46	CC	
	SAMT	J. Franklin	-122	44	CC	Redmond et al. (2012)
	SAPA	J. Franklin	-122	45	CC	
	SIRK	J. Franklin	-122	46	CC	
	SLBE	J. Franklin	-122	46	CC	

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
	STMT	J. Franklin	-122	46	CC	
	TIRD	J. Franklin	-122	46	CC	
	WIMT1	J. Franklin	-122	44	CC	
	WIMT2	J. Franklin	-122	44	CC	
	WISP	J. Franklin	-122	43	CC	
	WREF	J. S. Clark; J. HilleRisLambers	-122	46	ST CC	
Chihuahuan desert						
	FOBA	M. Redmond	-108	33	CC	Redmond et al. (2012)
Colorado Plateau shrublands						
	ALBU	A. Wion; M. Redmond	-106	35	CC	Wion <i>et al.</i> (2020)
	CEBO	A. Wion; M. Redmond	-106	36	CC	Rodman <i>et al.</i> (2020)
	DOLO	A. Wion; M. Redmond	-109	38	CC	Rodman <i>et al.</i> (2020)
	GLPA	A. Wion; M. Redmond	-109	39	CC	Rodman <i>et al.</i> (2020)
	HOND	A. Wion; M. Redmond; K. Rodman	-106	37	CC	
	HOTC	A. Wion; M. Redmond; K. Rodman	-108	39	CC	Rodman <i>et al.</i> (2020)
	LASA	A. Wion; M. Redmond	-109	39	CC	Rodman <i>et al.</i> (2020)
	MAGD	A. Wion; M. Redmond	-107	34	CC	Rodman <i>et al.</i> (2020)
	MONT	A. Wion; M. Redmond	-108	38	CC	Rodman <i>et al.</i> (2020)
	NATU	A. Wion; M. Redmond	-109	38	CC	Rodman <i>et al.</i> (2020)
	SEV	R. Zlotin; D. Macias	-107	34	CC	Parmenter et al. (2018)
	SUCR	A. Whipple; C. Gering; T. Whitham	-111	36	CC	Whipple et al. (2019)
Colorado Rockies forests						
	BOCA	I. Pearse	-105	40	CC	
	CANJ	A. Wion; M. Redmond	-106	36	СС	Rodman <i>et al.</i> (2020)
	HAYM	A. Wion; M. Redmond	-105	39	СС	
	LAK	K. Rodman	-106	36	СС	Rodman <i>et al.</i> (2020)
	LV	M. Redmond	-105	36	CC	Redmond et al. (2012)

Table S2 – continued from previous page

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
-	MG	K. Rodman	-106	36	CC	Rodman <i>et al.</i> (2020)
	MON	K. Rodman	-106	36	CC	Rodman <i>et al.</i> (2020)
	MR	K. Rodman	-106	36	CC	Rodman <i>et al.</i> (2020)
	MVG	K. Rodman	-106	36	CC	Rodman <i>et al.</i> (2020)
	NIWO	J. S. Clark	-106	40	ST CC	
	PC	K. Rodman	-106	36	CC	Rodman <i>et al.</i> (2020)
	PECO	M. Redmond	-106	36	CC	Redmond <i>et al.</i> (2012)
	POND	A. Wion; M. Redmond	-107	36	CC	Wion <i>et al.</i> (2020)
	RATN	M. Redmond	-104	37	CC	Redmond et al. (2012)
	SAFE	A. Wion; M. Redmond	-106	36	CC	Rodman <i>et al.</i> (2020)
	WACA	A. Wion; M. Redmond	-105	39	CC	
	WEMO	A. Wion; M. Redmond	-105	38	CC	
Cross-Sanaga-Bioko coastal forests						
	KNP	J. Norghauer	9	5	CC	Norghauer & Newbery (2015)
Dinaric Mountains mixed forests						
E Cascades forests						
E forest-boreal transition						
	LMONT	Y. Bergeron; Y. Messaoud	-79	48	CC	Messaoud et al. (2007)
E Great Lakes lowland forests						
English Lowlands beech forests						
	BEECH	A. Hacket-Pain	0	52	CC	Bogdziewicz et al. (2020)
	BUCKH	A. Hacket-Pain	-2	52	CC	Bogdziewicz et al. (2020)
	FISHH	A. Hacket-Pain	-2	52	CC	Bogdziewicz et al. (2020)
	NETTL	A. Hacket-Pain	-1	52	CC	Bogdziewicz et al. (2020)
	PAINS	A. Hacket-Pain	-2	52	CC	Bogdziewicz et al. (2020)
	PATCH	A. Hacket-Pain	-0	51	CC	Bogdziewicz et al. (2020)
	STP	M. Fenner M. Hanley	-1	51	СС	Hanley <i>et al.</i> (2018)

Table S2 – continued from previous page

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
Great Basin shrub steppe						
	DSP	M. Redmond	-119	39	CC	
Iberian sclerophyllous/semi-deciduous						
	CARB	C. Perez-Izquierdo	-6	40	CC	
	HUEC	R. Bonal	-4	40	CC	
	SIOE	R. Calama	-4	40	CC	
	VALT	J. Espelta	-4	41	CC	
Illyrian deciduous forests						
Interior Alaska-Yukon lowland taiga						
	EAPL	J. Johnstone	-137	66	CC	Viglas <i>et al.</i> (2013)
	BONA	J. Johnstone	-148	65	ST	
	FAIR	J. Johnstone	-148	65	CC	Viglas <i>et al.</i> (2013)
	LAGE	J. Johnstone	-145	64	CC	Viglas <i>et al.</i> (2013)
	LELA	J. Johnstone	-138	64	CC	Viglas <i>et al.</i> (2013)
	SMR	J. Johnstone	-141	64	CC	Viglas <i>et al.</i> (2013)
Interior Yukon-Alaska alpine tundra						
	CHIC	J. Johnstone	-143	63	CC	Viglas <i>et al.</i> (2013)
Isthmian-Atlantic moist forests						
	BCI	S.J. Wright	-80	9	ST	
Italian sclerophyllous/semi-deciduous						
Klamath-Siskiyou forests						
	ASRN	J. Franklin	-123	42	CC	
	MEOV	J. Franklin	-123	42	CC	
Mid Atlantic coastal forests						
	BLSF	D. Brockway	-79	35	CC	Chen <i>et al.</i> (2018)
	CALL	J. S. Clark	-79	35	ST CC	
	CROA	S. Cohen	-77	35	СС	

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
	GRSW	J. S. Clark	-78	34	ST	
	SASF	D. Brockway	-81	34	CC	Chen <i>et al.</i> (2018)
Mississippi lowland forests						
	CHICK	J. Straub; T. Leininger	-90	36	CC	Straub <i>et al.</i> (2016)
	DELTA	J. Straub; T. Leininger	-91	33	CC	Straub et al. (2016)
	MINGO	J. Straub; T. Leininger	-90	37	CC	Straub <i>et al.</i> (2016)
	TENAS	J. Straub; T. Leininger	-91	32	CC	Straub et al. (2016)
	WHITE	J. Straub; T. Leininger	-91	34	CC	Straub <i>et al.</i> (2016)
Montana Valley/Foothill grasslands						
N California coastal forests						
	UCSC	G. Gilbert; Kai Zhu	-122	37	ST	
N Central Rockies forests						
	WBP	E. McIntire	-114	48	CC	
N short grasslands						
NE coastal forests						
NE Spain/S France Mediterranean						
	ISS	H. Davi	6	44	CC	Davi <i>et al.</i> (2016)
	PCMEJEAN	T. Curt	3	44	CC	Debain <i>et al.</i> (2003)
	PUECHEXP1	J. Limousin; J. Ourcival	4	44	ST	
	RBI	T. Boivin	5	44	CC	Doublet <i>et al.</i> (2019)
	RBLL	H. Davi	6	44	CC	Davi <i>et al.</i> (2016)
	VALLI	F. Lefevre; F. Courbet	5	44	CC	
	VEN	H. Davi	5	44	CC	Davi <i>et al.</i> (2016)
	VENT	H. Davi	5	44	CC	Davi <i>et al.</i> (2016)
	VES	H. Davi	7	44	СС	Davi <i>et al.</i> (2016)
New England-Acadian forests						
	ASWP	C. Moore; J. S. Clark	-69	45	ST CC	

39

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
•	BART	I. Fer; M. Dietze	-71	44	ST	
	COMPT	W. Schlesinger	-67	45	CC	
	HARV	J. S. Clark	-72	42	ST CC	
	HBEF	T. Fahey; N. Cleavitt	-72	44	ST	Cleavitt & Fahey (2017)
Nihonkai montane deciduous forests			. –		0.	
	KANU	K. Hoshizaki	141	39	ST	
	JNP	Q. Han; D. Kabeya; K. Noguchi	139	37	ST	Han <i>et al.</i> (2014)
NW Congolian lowland forests	••••			0.	0.	( <u>_</u> )
	CONGO	J. Poulson; C. Nunez	16	2	ST	
Pannonian mixed forests	0011010			-	5.	
Peruvian Yungas						
	ABERG	M. Silman; W. Farfan	-72	-13	ST	
Piney Woods forests					01	
	KINF	D. Brockway	-92	31	СС	Chen <i>et al.</i> (2018)
Pontic steppe		2. 2.00.004	02	01	00	
Puerto Rican dry forests						
	GUA	M. Uriarte	-67	18	ST	Uriarte <i>et al.</i> (2012)
Puerto Rican moist forests		Wi. Charlo	07	10	01	
	LUQ	M. Uriarte	-66	18	ST	Uriarte <i>et al.</i> (2012)
Puget lowland forests	200			.0	0.	
Pyrenees conifer/mixed forests						
	PNVO	J. Camarero	-1	43	ST	de Andrés <i>et al.</i> (2014)
	PNP	S. Delzon; T. Caignard	-0	43	CC	Caignard <i>et al.</i> (2017)
Rodope montane mixed forests			Ŭ	10		
S Central Rockies forests						
	YELL	J. S. Clark	-110	45	ST CC	
S Great Lakes forests		0. 0. Olark	110	70	0100	

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
	ANNA	I. Ibanez	-84	42	ST	Redmond et al. (2012)
SE conifer forests						
	APNF	D. Brockway	-85	30	CC	Chen <i>et al.</i> (2018)
	BRSF	D. Brockway	-87	31	CC	Chen <i>et al.</i> (2018)
	DSNY	J. S. Clark	-81	28	ST CC	
	EAFB	D. Brockway	-87	30	CC	Chen <i>et al.</i> (2018)
	EEF	D. Brockway	-87	31	CC	Chen <i>et al.</i> (2018)
	JERC	D. Brockway	-84	31	CC	Chen <i>et al.</i> (2018)
	OSBS	J. S. Clark	-82	30	ST CC	
	STCB	D. Brockway	-85	31	CC	Chen <i>et al.</i> (2018)
	TTRS	D. Brockway	-86	31	CC	Chen <i>et al.</i> (2018)
SE mixed forests						
	DUKE	J. S. Clark	-79	36	ST CC	Berdanier & Clark (2016)
	FBMB	D. Brockway	-85	32	CC	Chen <i>et al.</i> (2018)
	SERC	J. S. Clark	-77	39	ST CC	van Mantgem et al. (200
	TALL	J. S. Clark	-87	33	ST CC	
Sierra Nevada forests						
	SEQU	A. Das; N. Stephenson	-119	37	ST	van Mantgem <i>et al.</i> (200
	SOAP	J. S. Clark	-119	37	ST CC	
	YOSE	A. Das; N. Stephenson	-120	38	ST	van Mantgem <i>et al.</i> (200
SW Iberian Mediterranean S/M						
	ALCO	I. Perez-Ramos	-6	36	CC	Pérez-Ramos <i>et al.</i> (201
	PNLA	A. Hampe	-6	37	CC	Hampe & Bairlein (2000)
	SJDV	F. Rodriguez-Sanchez	-6	37	CC	
Taiwan subtropical evergreen forests						
	FFDF	C. Chang-Yang; I-Fang Sun	122	25	ST	
Upper Midwest forest-savanna transition						

Eco-region	Plot	PI(s)	lon	lat	ST/CC	Citation
Valdivian temperate forests						
	LNP	J. Sanguinetti; T. Kitzberger	-71	-39	СС	Sanguinetti & Kitzberger (2008
	ARAU	M. Aravena; S. Donoso Calderon	-71	-38	CC	
W European broadleaf forests						
W Great Lakes forests						
	CADI	R. Kobe	-86	44	ST CC	
	MANI	R. Kobe	-86	44	ST CC	
	MICH	J. LaMontagne	-88	47	CC	
	PAFA	J. LaMontagne	-90	46	CC	
	TREE	J. S. Clark	-90	45	ST CC	
	UMBS	I. Ibanez	-85	46	ST	
	UNDE	J. S. Clark	-90	46	ST CC	
	WILW	J. LaMontagne	-90	46	CC	
	WORU	J. LaMontagne	-90	46	CC	
W Gulf coastal grasslands						
W short grasslands						
	CMNM	M. Redmond	-104	37	CC	Redmond et al. (2012)
	KENT	M. Redmond	-103	37	CC	Redmond et al. (2012)
Willamette Valley forests						
	CAMT	J. Franklin	-123	45	CC	
Wyoming Basin shrub steppe						
	NORT	A. Wion; M. Redmond	-109	41	CC	

Data source of <i>d<sub>max</sub></i>	Nb of species	Comments
National Forest Inventories	38	Used near maximum diameter forest inventory and analysis in the United States (Granet al., 2012); forest inventory in Europe (Kunstler <i>et al.</i> , 2021); tree census data in Japan
		(Ishihara <i>et al.</i> , 2011).
Unpublished field tropical measurements	222	Maximum diameter value observed from French Guyana (33 species) and from Panama (189 species).
Allometric equation	12	Maximum trait height (Liu <i>et al.</i> , 2019; Car mona <i>et al.</i> , 2021) and allometric equation from pan-tropical for Africa region, South America equation for South America region and Asia equation for Asia region.
MASTIF inventories	41	Used tree size observations and near max mum diameter for species with > 90 individua unique diameter.
Online open sources	173	<pre>Wikipedia, https://www.wikipedia.org/ American conifers society, https //conifersociety.org/; Monumental tree https://www.monumentaltrees.com/en/; iPlantz, https://www.iplantz.com/, Use ful tropical plants, https://tropical theferns.info/)</pre>

Table S3: Origin of maximum diameter ( $d_{max}$ ), with the number of species for each data source retained for the analysis used in this study.

Table S4: Summary of all fitted linear models between maturation size  $(d_{mat})$  and maximum size  $(d_{max})$ in  $\log_{10}$ - $\log_{10}$  testing alternative effects of climatic variables. The mean value of estimates with a confidence interval of 95%, and their p-values (p) are reported. All models have the initial structure  $\log_{10}(d_{mat_s}) = \alpha + \beta_d \times \log_{10}(d_{max_s})$  represented here by the ellipse (...). Other parameters included are temperature (T)and deficit (D).

Parameters	Estimate	2.5%	97.5%	p	AIC
$\frac{1}{\dots + \beta_T \times T}$				ľ	
$\alpha$	3.7100	1.94000	7.0700	< 0.001	-62.84
$eta_d$	0.3050	0.15100	0.4590	< 0.001	
$\beta_T$	-0.0233	-0.03520	-0.0114	< 0.001	
$\beta_{dT}$	0.0126	0.00586	0.0193	< 0.001	
					$\log_{10}(d_{max_s})$
$\alpha$		1.99e+00	8.65e+00		-59.29
$\beta_d$	2.81e-01	1.09e-01	4.53e-01		
	-2.43e-02	-3.66e-02	-1.20e-02	< 0.001	
$\beta_D$	2.73e-05	-7.98e-05	1.34e-04	0.617000	
$\beta_{dT}$		6.18e-03		< 0.001	
$\beta_{dD}$		-7.62e-05	5.03e-05	0.688000	
$\alpha$	1.080	0.929	1.250	0.325	-52
$\beta_d$	0.592	0.552	0.631	< 0.001	
$\frac{\beta^{\mu}u}{\dots+\beta_D\times D}$					
$\alpha$		8.17e-01		0.657	-48.12
$\beta_d$	5.93e-01	5.27e-01	6.59e-01	< 0.001	
$\beta_D$		-1.13e-04		0.910	
$\beta_{dD}$	1.40e-06	-6.18e-05	6.47e-05	0.964	
$\frac{1}{\dots + \beta_D \times D}$	$+\beta_T \times T$				
$\alpha$		9.32e-01	1.51e+00	0.165	-49.48
$\beta_d$	5.83e-01	5.42e-01	6.25e-01	< 0.001	
$\beta_D$	-3.90e-06	-2.53e-05	1.75e-05	0.721	
$\beta_T$	-1.59e-03	-4.29e-03	1.10e-03	0.246	
$\frac{\beta + 1}{\dots + \beta_D \times D}$					
$\alpha$	1.07e+00	9.09e-01	1.25e+00	0.433	-50.12
$eta_d$	5.92e-01	5.53e-01	6.31e-01	< 0.001	
$\beta_D$	-3.70e-06	-2.52e-05	1.77e-05	0.732	
$\frac{1}{\dots + \beta_T \times T}$					
$\alpha$	1.20000	0.94800	1.52000	0.129	-51.35
$\beta_d$	0.58300	0.54100	0.62500	< 0.001	
$\beta_T$	-0.00159	-0.00429	0.00111	0.248	
$\frac{\alpha}{\alpha+1\times\beta_d}$					
$\alpha$	0.244	0.232	0.257	0	248.1
$\beta_d$	1	2		-	
$\frac{\beta^{\alpha} u}{\alpha}$		2	+4		
$\alpha$	9.25	8.69	9.85	0	447.83
$\beta_d$	0	-	-	-	
1~ u	5				

Table S5: Summary of the best joint trait model. The best model has been selected based on the lowest DIC value (Table S7). Traits have been included as responses ( $d_{max}$ ,  $d_{mat}$ , SLA, Wood density, seed size, and species seed productivity), with temperature (T), deficit (D), and their interaction (T : D) as predictors, with genus as a random effect. For each response, estimate, standard error (SE), and credible interval (95%) with significance are reported (CI does not overlap 0).

Climate variable	Estimate	SE	2.5%	97.5%	significance
$d_{max}$					
Т	-0.230	0.068	-0.360	-0.097	*
D	0.150	0.050	0.054	0.250	*
T:D	-0.096	0.065	-0.220	0.031	
$d_{mat}$					
Т	-0.190	0.065	-0.320	-0.064	*
D	0.077	0.049	-0.018	0.170	
T:D	0.010	0.064	-0.110	0.140	
SLA					
Т	-0.006	0.073	-0.150	0.140	
D	0.190	0.051	0.083	0.280	*
T:D	-0.047	0.065	-0.180	0.079	
Wood density					
Т	0.160	0.067	0.030	0.300	*
D	0.064	0.050	-0.033	0.160	
T:D	0.019	0.064	-0.110	0.150	
Seed size					
Т	0.046	0.071	-0.093	0.180	
D	-0.180	0.050	-0.270	-0.078	*
T:D	-0.075	0.064	-0.200	0.054	
Seed productivity					
Т	-0.130	0.061	-0.250	-0.012	*
D	-0.061	0.048	-0.150	0.032	
T:D	-0.037	0.064	-0.160	0.085	

Table S6: Summary of conditional parameters for the effect on  $d_{mat}$  from the GJAM joint trait model. Conditioning was done on SLA, wood density, species seed productivity, seed size, and  $d_{max}$ . Standardized coefficient values are coming from matrix **A**, for direct trait effect, and **C**, for direct climate effect.

Conditional variable	Estimate	SE	2.5%	97.5%	significance
Traits					
$d_{max}$	0.5460	0.0266	0.4940	0.5960	*
SLA	-0.0569	0.0576	-0.1720	0.0552	
Wood density	-0.1300	0.0856	-0.3020	0.0384	
Seed size	0.0403	0.0120	0.0169	0.0639	*
Seed productivity	-0.0385	0.0100	-0.0574	-0.0180	*
Climate					
Intercept	8.36e-01	2.69e-01	3.22e-01	1.36e+00	*
Т	4.11e-03	4.66e-03	-4.93e-03	1.33e-02	
D	-1.59e-04	9.53e-05	-3.49e-04	3.28e-05	
T:D	8.30e-06	4.60e-06	-8.00e-07	1.73e-05	

Climatic predictors in GJAM	DIC
$T \times D$	20,575
T + D	20,603
Т	20,661
D	20,898

Table S7: Joint traits model selection (based on the lowest DIC values). GJAM models ran with different combinations of climate covariates (T, temperature, and D, deficit).

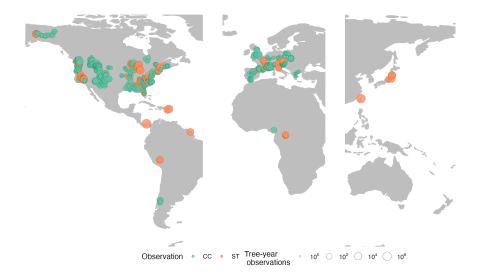


Figure S1: MASTIF data network, including seed traps data and crop count data limited here to speciesgenus used. The dot size represents the number of initial tree-year observations at the log10 scale. Crop count data (green dots, CC) includes 137,484 tree years observations and seed traps (orange dots, ST) 10,914,392 observations in total.

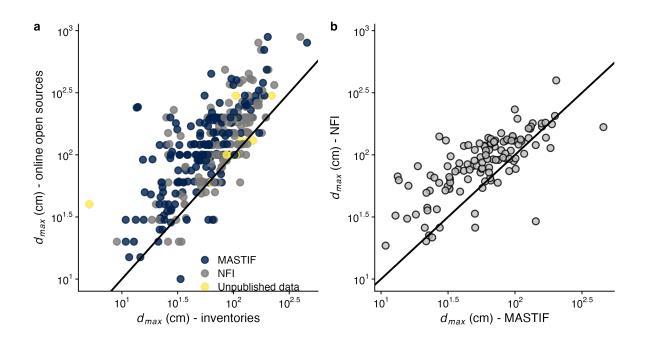


Figure S2: a) Comparison of  $d_{max}$  obtained from online open sources to  $d_{max}$  obtained from National Forest Inventories (NFI), MASTIF inventories, and from the unpublished dataset (n=191 species). b) Comparison of  $d_{max}$  obtained from National Forest Inventories (NFI) to  $d_{max}$  obtained from MASTIF inventories (N = 118 species). For both panels, the black line is the 1:1 relationship.

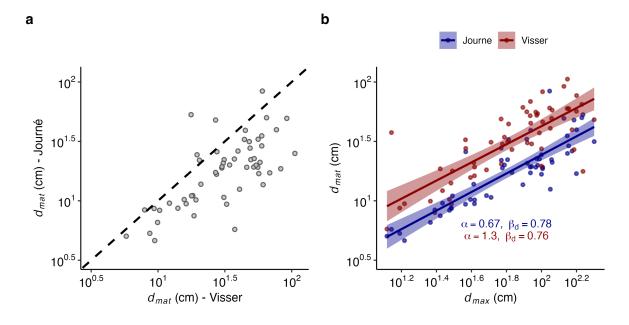


Figure S3: a) Comparison of maturation size  $(d_{mat})$  from the main analysis (probability to produce the first fruiting structure) to maturation size (probability to reproduce at 50%) based on the reference in Visser *et al.* (2016), restricted to Barro Colorado Island, Panama. Each dot represents a single species, with the black dotted line indicating a 1:1 relationship (N = 56 species). b) Relationship between  $d_{mat}$  and  $d_{max}$  restricted to species used by Visser *et al.* (2016). The model fitted between  $d_{mat}$  and  $d_{max}$  is shown in blue for the model based on our estimates of  $d_{mat}$  and in red for estimates of  $d_{mat}$  from Visser *et al.* (2016). The regressions are reported with a confidence interval of 0.95. The average parameters  $\alpha$  and  $\beta_d$  are reported for both models.

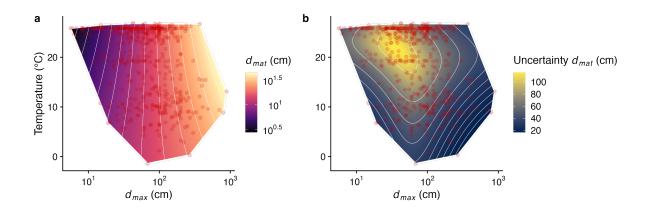


Figure S4: a) Maturation size response to  $d_{max}$  and temperature and b) Uncertainty of maturation size. Convex hulls are defined by observations (red), including the 486 tree species. In b) the surface scale color decreases as the inverse of the predictive standard error-blue edges reflect increased uncertainty at data extremes.

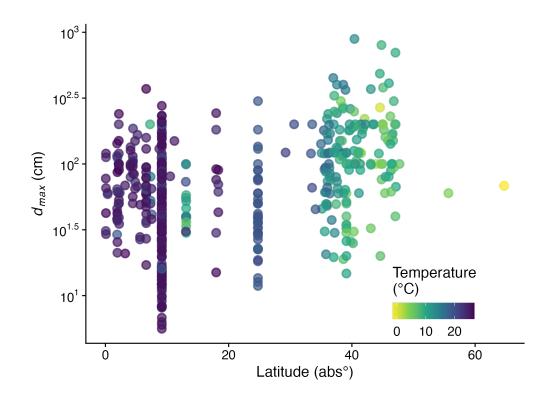


Figure S5: Relationship between  $d_{max}$  with latitude (in absolute degree) accross species. The color gradient represents the average temperature (in °C). The average latitude for each species was determined by using MASTIF inventories. Each dot represents one species.

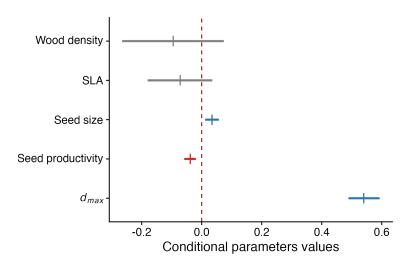


Figure S6: Conditional parameter estimates for the direct effect of traits on maturation size  $(d_{mat})$ , while accounting for trait covariance, climate and phylogeny. The climate used here has been extracted from the MASTIF inventories. Conditional parameters are evaluated on a standardized scale, making trait effects on  $d_{mat}$  respective to their variation in the data set. Points represent the posterior mean with their 95% credible intervals. Blue and red represent positive and negative associations where 95% of the posterior does not include zero. SLA, specific leaf area.

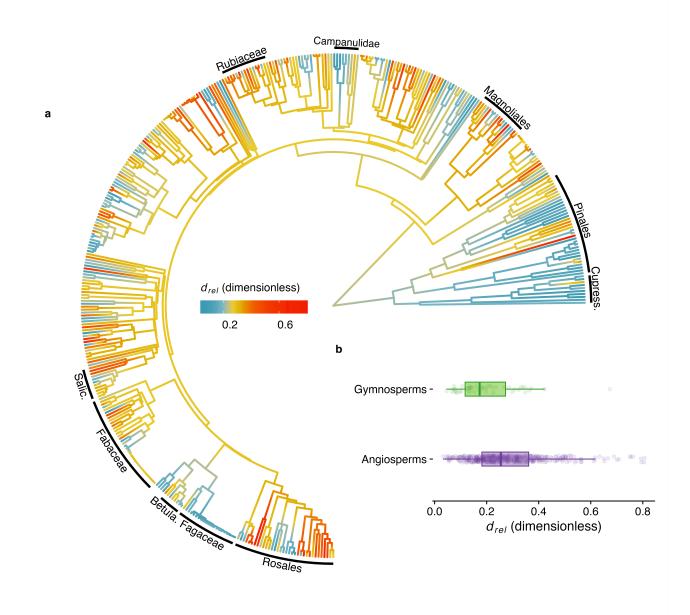


Figure S7: (a) Relative size at maturation  $(d_{rel})$  includes a phylogenetic signal (400 species in our data have phylogenies in Zanne *et al.* 2014, Pagel's  $\lambda = 0.51$ , p < 0.0001). (b) Boxplot of relative size at maturation  $(d_{rel})$  for gymnosperms and angiosperms (number of species is 48 for gymnosperm and 438 for angiosperm).

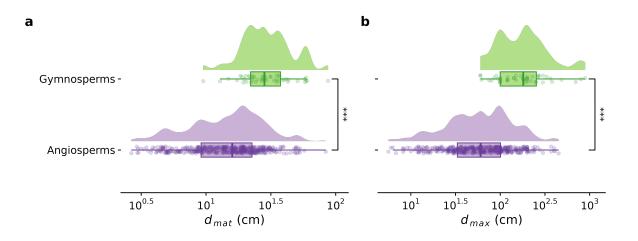


Figure S8: Boxplot of a)  $d_{mat}$  and b)  $d_{max}$  for gymnosperms (n = 48) and angiosperms (n = 438). \*\*\* indicates p < 0.0001 based on the sample t-test for unequal variances.

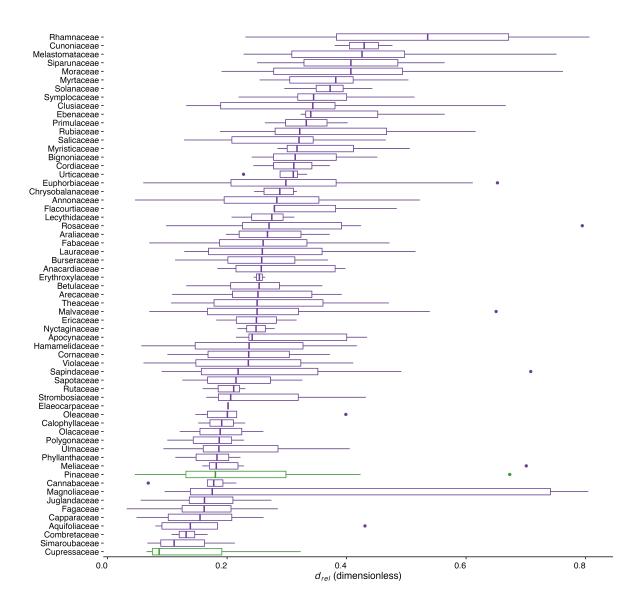


Figure S9: Boxplot of  $d_{rel}$  for families with more than one species. Green is for gymnosperms and purple for angiosperms.

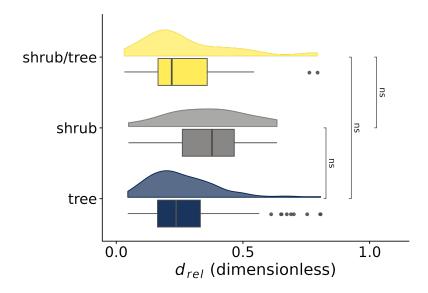


Figure S10: Boxplot with violin of variation of  $d_{rel}$  across growth forms (n = 419 species). The violin here is a mirrored density plot and showed the distribution of the data. Growth form follows a compilation from Díaz *et al.* (2022), with samples: trees, n = 361 species; shrubs, n = 17 species; shrub/tree n = 41 species. Groups were compared with a t-test for unequal variance and detected no differences according to plant growth forms (non-significant adjusted p-values with p > 0.05).

## **References supplementary material**

- de Andrés, E.G., Camarero, J.J., Martínez, I. & Coll, L. (2014). Uncoupled spatiotemporal patterns of seed
   dispersal and regeneration in pyrenean silver fir populations. *Forest Ecology and Management*, 319, 18–
   28.
- Berdanier, A.B. & Clark, J.S. (2016). Divergent reproductive allocation trade-offs with canopy exposure across
   tree species in temperate forests. *Ecosphere*, 7, e01313–n/a.
- Bogdziewicz, M., Acuña, M.C.A., Andrus, R., Ascoli, D., Bergeron, Y., Brveiller, D. *et al.* (2023). Linking seed
   size and number to trait syndromes in trees. *Global Ecology and Biogeography*, 32, 683–694.
- Bogdziewicz, M., Kelly, D., Thomas, P.A., Lageard, J.G.A. & Hacket-Pain, A. (2020). Climate warming disrupts
  mast seeding and its fitness benefits in european beech. *Nature Plants*, 6, 88–94.

Bourg, N.A., McShea, W.J., Thompson, J.R., McGarvey, J.C. & Shen, X. (2013). Initial census, woody
 seedling, seed rain, and stand structure data for the scbi sigeo large forest dynamics plot. *Ecology*, 94,
 2111–2112.

- Caignard, T., Kremer, A., Firmat, C., Nicolas, M., Venner, S. & Delzon, S. (2017). Increasing spring tempera tures favor oak seed production in temperate areas. *Scientific Reports*, 7, 8555.
- Carmona, C.P., Bueno, C.G., Toussaint, A., Träger, S., Díaz, S., Moora, M. *et al.* (2021). Fine-root traits in the
   global spectrum of plant form and function. *Nature*, 597, 683–687.
- Chen, X., Brockway, D.G. & Guo, Q. (2018). Characterizing the dynamics of cone production for longleaf pine
   forests in the southeastern united states. *Forest Ecology and Management*, 429, 1–6.
- <sup>852</sup> Clark, J.S., Bell, D.M., Kwit, M.C. & Zhu, K. (2014). Competition-interaction landscapes for the joint response
   <sup>853</sup> of forests to climate change. *Global Change Biology*, 20, 1979–1991.
- <sup>854</sup> Clark, J.S., Iverson, L., Woodall, C.W., Allen, C.D., Bell, D.M., Bragg, D.C. *et al.* (2016). The impacts of
   <sup>855</sup> increasing drought on forest dynamics, structure, and biodiversity in the united states. *Global Change* <sup>856</sup> *Biology*, 22, 2329–52.
- Clark, J.S., LaDeau, S. & Ibanez, I. (2004). Fecundity of trees and the colonization–competition hypothesis.
   *Ecological Monographs*, 74, 415–442.
- Cleavitt, N.L. & Fahey, T.J. (2017). Seed production of sugar maple and american beech in northern hardwood
   forests, new hampshire, usa. *Canadian Journal of Forest Research*, 47, 985–990.
- Daskalakou, E.N., Koutsovoulou, K., Ioannidis, K., Koulelis, P.P., Ganatsas, P. & Thanos, C.A. (2019). Masting
   and regeneration dynamics of abies cephalonica, the greek endemic silver fir. *Seed Science Research*,
   29, 227–237.

Davi, H., Cailleret, M., Restoux, G., Amm, A., Pichot, C. & Fady, B. (2016). Disentangling the factors driving
 tree reproduction. *Ecosphere*, 7, e01389.

Debain, S., Curt, T., Lepart, J. & Prevosto, B. (2003). Reproductive variability in pinus sylvestris in southern
 france: Implications for invasion. *Journal of Vegetation Science*, 14, 509–516.

Detto, M., Visser, M.D., Wright, S.J. & Pacala, S.W. (2019). Bias in the detection of negative density dependence in plant communities. *Ecology Letters*, 22, 1923–1939.

Díaz, S., Kattge, J., Cornelissen, J.H., Wright, I.J., Lavorel, S., Dray, S. *et al.* (2022). The global spectrum of
 plant form and function: enhanced species-level trait dataset. *Scientific Data*, 9, 1–18.

Dormont, L., Baltensweiler, W., Choquet, R. & Roques, A. (2006). Larch- and pine-feeding host races of
 the larch bud moth (zeiraphera diniana) have cyclic and synchronous population fluctuations. *Oikos*, 115,
 299–307.

<sup>875</sup> Doublet, V., Gidoin, C., Lefèvre, F. & Boivin, T. (2019). Spatial and temporal patterns of a pulsed resource <sup>876</sup> dynamically drive the distribution of specialist herbivores. *Scientific Reports*, 9, 17787.

Gray, A.N., Brandeis, T.J., Shaw, J.D., McWilliams, W.H. & Miles, P. (2012). Forest inventory and analysis
database of the united states of america (fia). *In: Dengler, J.; Oldeland, J.; Jansen, F.; Chytry, M.; Ewald, J., Finckh, M.; Glockler, F.; Lopez-Gonzalez, G.; Peet, RK; Schaminee, J. HJ, eds. Vegetation databases*for the 21st century. Biodiversity and Ecology. *4: 225-231.*, pp. 225–231.

Hacket-Pain, A., Ascoli, D., Berretti, R., Mencuccini, M., Motta, R., Nola, P. *et al.* (2019). Temperature and
 masting control norway spruce growth, but with high individual tree variability. *Forest Ecology and Management*, 438, 142–150.

Hampe, A. & Bairlein, F. (2000). Modified dispersal-related traits in disjunct populations of bird-dispersed
 frangula alnus (rhamnaceae): a result of its quaternary distribution shifts? *Ecography*, 23, 603–613.

Han, Q., Kabeya, D., Iio, A., Inagaki, Y. & Kakubari, Y. (2014). Nitrogen storage dynamics are affected by
 masting events in faguscrenata. *Oecologia*, 174, 679–687.

Hanley, M.E., Cook, B.I. & Fenner, M. (2018). Climate variation, reproductive frequency and acorn yield in
 english oaks. *Journal of Plant Ecology*, 12, 542–549.

Ishihara, M.I., Suzuki, S.N., Nakamura, M., Enoki, T., Fujiwara, A., Hiura, T. *et al.* (2011). Forest stand
 structure, composition, and dynamics in 34 sites over japan. *Ecological Research*, 26, 1007–1008.

Knops, J.M.H. & Koenig, W.D. (2012). Sex allocation in california oaks: Trade-offs or resource tracking? *PLOS ONE*, 7, e43492.

- Kunstler, G., Guyennon, A., Ratcliffe, S., Rüger, N., Ruiz-Benito, P., Childs, D.Z. *et al.* (2021). Demographic
   performance of European tree species at their hot and cold climatic edges. *Journal of Ecology*, 109, 1041–1054.
- Liu, H., Gleason, S.M., Hao, G., Hua, L., He, P., Goldstein, G. *et al.* (2019). Hydraulic traits are coordinated with maximum plant height at the global scale. *Science Advances*, 5.
- van Mantgem, P.J., Stephenson, N.L. & Keeley, J.E. (2006). Forest reproduction along a climatic gradient in
   the sierra nevada, california. *Forest Ecology and Management*, 225, 391–399.

Messaoud, Y., Bergeron, Y. & Asselin, H. (2007). Reproductive potential of balsam fir (abies balsamea), white
 spruce (picea glauca), and black spruce (p. mariana) at the ecotone between mixedwood and coniferous
 forests in the boreal zone of western quebec. *American Journal of Botany*, 94, 746–754.

Nab, L. (2021). *mecor: Measurement Error Correction in Linear Models with a Continuous Outcome*. R
 package version 1.0.0.

Nab, L., van Smeden, M., Keogh, R.H. & Groenwold, R.H. (2021). Mecor: An R package for measurement
 error correction in linear regression models with a continuous outcome. *Computer Methods and Programs in Biomedicine*, 208, 106238.

- Norghauer, J.M. & Newbery, D.M. (2015). Tree size and fecundity influence ballistic seed dispersal of two
   dominant mast-fruiting species in a tropical rain forest. *Forest Ecology and Management*, 338, 100–113.
- Parmenter, R.R., Zlotin, R.I., Moore, D.I. & Myers, O.B. (2018). Environmental and endogenous drivers of tree
   mast production and synchrony in piñon–juniper–oak woodlands of new mexico. *Ecosphere*, 9, e02360.
- Pérez-Ramos, I.M., Aponte, C., García, L.V., Padilla-Díaz, C.M. & Marañón, T. (2014). Why is seed production
   so variable among individuals? a ten-year study with oaks reveals the importance of soil environment.
   *PLOS ONE*, 9, e115371.
- Qiu, T., Sharma, S., Woodall, C.W. & Clark, J.S. (2021). Niche Shifts From Trees to Fecundity to Recruitment
   That Determine Species Response to Climate Change. *Frontiers in Ecology and Evolution*, 9, 1–12.
- Redmond, M.D., Forcella, F. & Barger, N.N. (2012). Declines in pinyon pine cone production associated with
   regional warming. *Ecosphere*, 3, art120.
- Rodman, K.C., Veblen, T.T., Chapman, T.B., Rother, M.T., Wion, A.P. & Redmond, M.D. (2020). Limitations
   to recovery following wildfire in dry forests of southern colorado and northern new mexico, usa. *Ecological Applications*, 30, e02001.
- Rose, A.K., Greenberg, C.H. & Fearer, T.M. (2012). Acorn production prediction models for five common oak
   species of the eastern united states. *The Journal of Wildlife Management*, 76, 750–758.

- Sanguinetti, J. & Kitzberger, T. (2008). Patterns and mechanisms of masting in the large-seeded southern
   hemisphere conifer araucaria araucana. *Austral Ecology*, 33, 78–87.
- Straub, J.N., Kaminski, R.M., Leach, A.G., Ezell, A.W. & Leininger, T. (2016). Acorn yield and masting traits
   of red oaks in the lower mississippi river alluvial valley. *Forest Science*, 62, 18–27.
- <sup>929</sup> Uriarte, M., Clark, J.S., Zimmerman, J.K., Comita, L.S., Forero-Montana, J. & Thompson, J. (2012). Multidi <sup>930</sup> mensional trade-offs in species responses to disturbance: implications for diversity in a subtropical forest.
   <sup>931</sup> *Ecology*, 93, 191–205.
- <sup>932</sup> Viglas, J., Brown, C. & Johnstone, J. (2013). Age and size effects on seed productivity of northern black
   <sup>933</sup> spruce. *Canadian Journal of Forest Research-Revue Canadienne De Recherche Forestiere*, 43, 534–543.
- Whipple, A.V., Cobb, N.S., Gehring, C.A., Mopper, S., Flores-Rentería, L. & Whitham, T.G. (2019). Long-term
   studies reveal differential responses to climate change for trees under soil- or herbivore-related stress.
   *Frontiers in Plant Science*, 10.
- Wion, A.P., Weisberg, P.J., Pearse, I.S. & Redmond, M.D. (2020). Aridity drives spatiotemporal patterns of
   masting across the latitudinal range of a dryland conifer. *Ecography*, 43, 569–580.
- Wright, B.R. & Zuur, A.F. (2014). Seedbank dynamics after masting in mulga (acacia aptaneura): Implications
   for post-fire regeneration. *Journal of Arid Environments*, 107, 10–17.