

1 Thermographic imaging in sports and exercise medicine: a Delphi study and consensus statement on
2 the measurement of human skin temperature

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25
26 **Word count:** 4091

27

1 **Abstract**

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The importance of using infrared thermography (IRT) to assess skin temperature (t_{sk}) is increasing in clinical settings. Recently, its use has been increasing in sports and exercise medicine; however, no consensus guideline exists to address the methods for collecting data in such situations. The aim of this study was to develop a checklist for the collection of t_{sk} using IRT in sports and exercise medicine. We carried out a Delphi study to set a checklist based on consensus agreement from leading experts in the field. Panelists (n = 24) representing the areas of sport science (n=8; 33%), physiology (n=7; 29%), physiotherapy (n=3; 13%) and medicine (n=6; 25%), from 13 different countries completed the Delphi process. An initial list of 16 points was proposed which was rated and commented on by panelists in three rounds of anonymous surveys following a standard Delphi procedure. The panel reached consensus on 15 items which encompassed the participants' demographic information, camera/room or environment setup and recording/analysis of t_{sk} using IRT. The results of the Delphi produced the checklist entitled "Thermographic Imaging in Sports and Exercise Medicine (TISEM)" which is a proposal to standardize the collection and analysis of t_{sk} data using IRT. It is intended that the TISEM can also be applied to evaluate bias in thermographic studies and to guide practitioners in the use of this technique.

Keywords: Infrared thermography, guideline, protocol, checklist, thermoregulation.

1 Introduction

2

3 The growing importance of infrared thermography (IRT) measures of human skin temperature (t_{sk}) in
4 health and disease has been evidenced by the increase in the number of publications with this technique. IRT
5 is characterized by the use of a camera which can detect radiation and produce thermal images, called
6 thermograms (Ring and Ammer, 2000). The thermograms contain temperature data that can be analyzed by
7 specific software which provides temperature of a region of interest (ROI) (Costello et al., 2012b; Selfe et al.,
8 2006). Common applications of IRT include: prevention and treatment of sports injuries (Hadžić et al., 2015;
9 Hildebrandt et al., 2010), detection of delayed onset muscle soreness (Hani et al., 2012), evaluation of
10 cryotherapy protocols (Adamczyk et al., 2016; Costello et al., 2012b; Selfe et al., 2014; Silva et al., 2017),
11 assessment of brown adipose tissue activation (Robinson et al., 2016), evaluation of t_{sk} during cold (Fournet
12 et al., 2013) and hot environment exposure (Gerrett et al., 2015; Vainer, 2005), following aerobic (Priego-
13 Quesada et al., 2015b), anaerobic (Adamczyk et al., 2014) and resistance exercises (Ferreira et al., 2008).
14 These applications have considerably growth in the use of IRT in recent years, due in part to improvements in
15 the accuracy, functionality and affordability of camera technology, thus making IRT an emerging method of
16 t_{sk} measurement in sports and exercise medicine (Bach et al., 2015b; Costello et al., 2012b; Fernandes et al.,
17 2014).

18 IRT is a rapid emerging technique for the assessment of t_{sk} as it is versatile, non-invasive, wireless,
19 and requires no contact with the individual (Bach et al., 2015b; Fernandes et al., 2014; Formenti et al., 2016).
20 Due to its image capture capability, the selection of ROIs permits an evaluation of the t_{sk} distributions in
21 different areas, consequently allowing its application in studies that require the analysis of several areas
22 simultaneously (Fournet et al., 2013; Gerrett et al., 2015). Moreover, the thermograms allow the visualization
23 of hot and cold areas. This has important implications in studies aimed at determining what location hot or
24 cold t_{sk} is generated (Costello et al., 2012a; Eglin et al., 2013; Maley et al., 2014; Robinson et al., 2016; Selfe
25 et al., 2010). Another advantage of IRT is the portability of cameras which can be used in a wide array of
26 conditions and locations in both the laboratory and the field (Fernandes et al., 2016; Hildebrandt et al., 2012).

1 Previous reports have demonstrated poor agreement between t_{sk} measurements by IRT and contact
2 devices (e.g. thermistors, thermocouples, iButtons) (Bach et al., 2015a; Bach et al., 2015b; Fernandes et al.,
3 2014). However, there are some inherent limitations related to the methodology of IRT data acquisition that
4 could act as confounding factors, thereby influencing temperatures outcomes (Fernández-Cuevas et al., 2015).
5 For example, the distance the camera is from the subject and the room temperature of the laboratory where
6 IRT recording is conducted can affect the data (Fernández-Cuevas et al., 2015). If thermograms from different
7 distances (fields of view) of the same subject are compared, the variable number of pixels within the ROI can
8 lead to inaccurate data (Ring and Ammer, 2000). Ammer (2015) compared the results of local thermograms
9 with a total body thermogram and showed differences in the t_{sk} of the anterior thigh of up to 1.09 ± 0.93 °C
10 (CI: 2.91; -0.74 °C). Likewise, a room without adequate temperature regulation can result in variable air
11 temperatures thereby impacting results (Bach et al., 2015b). Most published studies using IRT have employed
12 a temperature range of 18 °C to 25 °C (Fernández-Cuevas et al., 2015). However, it is well established that
13 resting metabolic rate varies as a function of ambient temperature which can markedly influence
14 thermoregulatory response (e.g. heat conservation or heat loss responses) and ultimate t_{sk} (Taylor et al., 2014).
15 Consequently, it is important to ensure that ambient temperature conditions are adequately regulated to
16 minimize any potential influence on the measurement of t_{sk} using IRT.

17 In order to prevent bias and improve the quality of data, several organizations have published their
18 own protocols and quality control guidelines (Ammer, 2008; IACT, 2002; ISO, 2004; Mercer and Ring, 2009;
19 Ring and Ammer, 2012; Ring and Ammer, 2000; Schwartz et al., 2006). Similarly, other investigators have
20 discussed the need for standardization to ensure the quality of thermal image acquisition (Ammer, 2003;
21 Ammer, 2015; Costello et al., 2012b; Ring et al., 2007a; Ring and Ammer, 2012). The acquisition of accurate
22 t_{sk} data requires knowledge of the primary factors influencing the t_{sk} measurement. Fernández-Cuevas et al.
23 (2015) defined the factors influencing the use of IRT in studies conducted on humans dividing them into
24 environmental, technical and individual factors. However, no specific guidelines or checklist was provided.
25 Given the wide array of factors that can affect the measurement of t_{sk} (Fernández-Cuevas et al., 2015), the use
26 of IRT for scientific analysis can be challenging. However, by defining appropriate measurement standards
27 and protocols, the accurate assessment of t_{sk} with IRT is possible. As reported by Costello et al. (2012b), many

1 researchers fail to report detailed information regarding the procedures and conditions under which IRT is
2 employed. As a consequence, there can be a lack of standardization between studies which can affect the
3 interpretability of the data. However, this limitation can be circumvented with the development of operational
4 standards for the use of IRT.

5 In this context, we propose a checklist based on consensus agreement from leading experts in the field.
6 Checklists tend to be more reliable than guidelines as they focus on the key points for simplicity and ease of
7 application (Kelley et al., 2003). Similar checklists such as the PEDro (Verhagen et al., 1998) and CONSORT
8 (Schulz et al., 2010) have been developed using expert consensus to improve methodological aspects of
9 research and the contribution of these instruments is well established (Moher et al., 2001; Moseley et al.,
10 2011). It is expected that a consensus instrument can contribute to the most appropriate application of IRT
11 technique, including data collection and analysis. Therefore, this study aimed to develop a detailed checklist
12 for the assessment of t_{sk} using IRT in sports and exercise medicine settings. It is intended to standardize the
13 collection and analysis of t_{sk} data by end users which include clinicians, researchers and practitioners. This
14 checklist could also be applied to evaluate bias in thermographic studies, and to guide practitioners in the use
15 of this technique.

17 **Methods**

19 **Participants**

21 Of the 30 invited experts, 25 agreed to participate in the study. Two experts declined the invitation
22 because they were too busy to participate, and a further three did not respond. One expert only completed the
23 first round leaving a total of 24 experts who participated in the full evaluation. Panelists were selected based
24 on their expertise in studies with IRT and thermal physiology, with at least three publications related to the
25 discipline or area. Once a person was identified as an expert, an e-mail was sent to him/her with an invitation
26 to participate; those who agreed received an electronic consent document. The panelists included experts who
27 self-defined themselves as working predominantly in the sport sciences (n=8; 33%), physiology (n=7; 29%),

1 physiotherapy (n=3; 13%) and medicine (n=6; 25%). They were currently working in academic and/or
2 research institutions (n=17; 71%), practicing in hospitals as medical doctors (n=3; 13%), working for a
3 company/industry (n=2; 8%), or working in the military (n=2; 8%). The panelists resided in the United
4 Kingdom (n=5; 21%), Brazil (n=3; 13%), Australia (n=2; 8%), Italy (n=2; 8%), Poland (n=2; 8%), Spain(n=2;
5 8%), United States (n=2; 8%), Austria (n=1; 4%), Canada (n=1; 4%), France (n=1; 4%), Mexico (n=1; 4%),
6 Portugal (n=1; 4%) and Russia (n=1; 4%). Panelists had a wealth of experience working with IRT,
7 thermoregulation, and the assessment of t_{sk} (Median=8 years; range 4-32), and had published a median of
8 eight (range 3-80) full peer reviewed articles related to the subject examined in the current manuscript. **In**
9 **addition, a search on the Scopus database on 06/26/2017 showed an average H index of 9 (range: 3 to 35).**
10 Participants received an information document describing the study with consent indicated by completion of
11 the Delphi survey. The first, second, third and last authors organized the work of the panel as part of the core
12 research team but did not participate as panel members. The core panel was responsible for the development
13 of the initial items, the analysis, organization and reporting of the decisions, as well as communicating with
14 the panelists.

16 Research Design

18 A Delphi procedure was applied in the present study, as previously described (Boulkedid et al., 2011;
19 Dalkey and Helmer, 1963; Hsu and Sandford, 2007; Steurer, 2011). The Delphi procedure is based on
20 developing a consensus among a group of experts through a series of questionnaires interspersed with
21 controlled feedback (Whiting et al., 2003). **In this procedure the expert evaluation**, judgment, phrasing and
22 scoring of each round is completed independently. As previously demonstrated, controlled and anonymous
23 feedback also helps the experts to gain a consensus (Boulkedid et al., 2011; Dalkey and Helmer, 1963; Hsu
24 and Sandford, 2007; Steurer, 2011). To initiate the process, a literature review was conducted in March 2016
25 to identify the available guidelines. Subsequently, instead of asking open questions to the panelists, an initial
26 list of items for inclusion in the checklist was developed. This approach is considered appropriate if basic
27 information is already available within the literature (Hsu and Sandford, 2007). In addition, as infrared

1 technology has been improved in recent years, only documents published since 2000 were included (for further
2 details and a full rationale see Costello et al. (2013) and Bach et al. (2015b)). Therefore, nine documents
3 (Ammer, 2008; Fernández-Cuevas et al., 2015; IACT, 2002; ISO, 2004; Mercer and Ring, 2009; Ring and
4 Ammer, 2000; Ring et al., 2007b; Schwartz et al., 2006; Schwartz et al., 2015) which contained
5 recommendations on the use of IRT for measurements in humans were used to develop an itemized list.

6 The core panel (authors DGM, JTC, CJB and MSQ) reviewed the nine documents to summarize the
7 empirical and theoretical evidence regarding the procedures involving IRT. A preliminary list (with 16 items
8 for inclusion in the checklist) was settled based on scientific evidence provided in the selected documents.
9 The core panel previously established that only aspects relating to the measurement of t_{sk} using IRT would be
10 included in the checklist, and other aspects in the applications of IRT were not included.

11 Although no criterion is universally accepted to address consensus, the value of 80% agreement is
12 mostly used (Bahl et al., 2016; Boulkedid et al., 2011; Hsu and Sandford, 2007; Snyder et al., 2014; Whiting
13 et al., 2003). A five point Likert Scale (strongly agree, moderately agree, neutral, moderately disagree, strongly
14 disagree) (Chipchase et al., 2012; Whiting et al., 2003) was used in all rounds to rate each item for inclusion
15 in the checklist. Therefore, we applied the criteria of 80% of the sum of responses of ‘strongly agree’ and
16 ‘moderately agree’ to approve an item. To help panel members in their decision-making, the core panel
17 summarized the evidence and provided the references which supported each item. In addition, the panelists
18 were encouraged to identify any study or practical experience that could help in the discussion. In every round,
19 all members were given an opportunity to comment on the items and suggest possible rephrasing. The panelists
20 had 15 days to respond to each round and all communication was conducted by electronic mail.

21 The responses of each Delphi round were organized to include the results of the previous round and a
22 summary of all panel members’ evaluation. All modifications were shared in the subsequent rounds as
23 previously reported (Chipchase et al., 2012; Snyder et al., 2014; Whiting et al., 2003). Furthermore, the
24 decision made in each round comprised of six distinct actions: (1) *Modify*: when an item was substantially
25 modified, either by suggestion of a panel member or to meet new evidence; (2) *Rephrase*: when an item was
26 rephrased to improve understanding without changing the meaning; (3) *Divide*: when an item was divided into
27 two different items; (4) *Exclude*: when an item did not meet the criteria and was excluded from the checklist;

1 (5) *Include*: when a panel member suggested a new item; and (6) *Approve*: when an item met the criteria and
2 was approved to be part of the checklist. Consensus was considered to be achieved for an item if: a) the criteria
3 of 80% of the sum of responses of ‘strongly agree’ and ‘moderately agree’ was reached; and b) no panelist
4 recommended a relevant change in wording or provided new or additional evidence. All comments regarding
5 rephrasing were incorporated when revising the checklist.

6 7 Round 1

8
9 The initial list of possible items for inclusion was sent to all panel members which included information
10 on the the aim of the consensus protocol and the process. The scientific reasoning for the inclusion of each
11 item was presented. All panelists were asked to analyze each item while considering two main points: (1)
12 validity, that was defined as the extent to which the characteristics of the item are appropriate to the objective
13 of the checklist (Boulkedid et al., 2011); and (2) feasibility, which was defined as the practical viability of the
14 item (Boulkedid et al., 2011). The panelists were encouraged to support their answers with information based
15 on peer-reviewed literature whenever possible. When this was not possible, it was suggested that they justify
16 their positions using their practical experience in the use of thermography.

17 18 Round 2

19
20 The results of round one were analyzed by the core panel and a report was prepared containing the
21 response rating, as well as a summary of all the comments received for each item. In addition, in this round,
22 panelists were asked to indicate which tense they preferred the checklist to be in, and whether they wanted to
23 propose a new item not included in the initial list. In this context, the following questions were asked: 1) In
24 order to improve the understanding and interpretation of the proposed items, which tense do you deem to be
25 the most appropriate for the presentation of the checklist? 2) Do you want to propose an item that is not
26 included in this checklist? If yes, please, propose the item and identify where you think it should be located in
27 the checklist.

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Round 3

Based on the results of round two, all of the items were modified according to the chosen tense (have/should/must). As some items were approved in round two (5 items), only those items that did not receive consensus were assessed further. The response ratings and a summary of all the comments received were presented again. The study was concluded in this round since we achieved the previously established threshold for consensus. The phases of the study are presented in Figure 1.

*** Insert Figure 1 here***

Results

Round 1

Decisions in this round included modification (6; 38%), rephrasing (8; 50%), and exclusion (2; 12%) of items. In general, the items obtained a high level of agreement between the experts, however some items were judged incomplete or wrong (the number of the approved item in the final checklist is expressed in table 1 – items: individual data, previous instructions, environmental condition, image background, acclimation and camera preparation) while others required alternate phrasing (items: extrinsic factors, environmental setup, equipment, image recording, camera position, emissivity, body position and image evaluation). For the incomplete items, new evidence was provided by panel members and subsequently incorporated. Likewise, suggested grammatical edits were incorporated to improve clarity. Although some items met the approval criteria, none was approved in this round, since it was deemed by the panelists that the proposed edits should be re-evaluated.

The core panel indentified two items (assessment time and method of drying the skin) to be removed since they were judged not relevant to the checklist. Five panelists argued that both items were not related to

1 the objective of the checklist and therefore they should be excluded. All edits were highlighted and explained
2 in subsequent round for evaluation.

3 4 Round 2

5
6 The decisions in this round comprised modification (4; 25%), rephrasing (6; 38%), division (1; 6%)
7 and approval (5; 31%). Five items were approved since they met the approval criteria and no further relevant
8 information was provided by the panelists (items: extrinsic factor, environmental condition, camera position,
9 emissivity and image evaluation). Ten panelists suggested a new approach to address the items “assessment
10 time” and “method of drying the skin”, which were suggested to be removed in the previous round because
11 they were not related to the goal of the checklist. Based on the feedback received on these items, a new version
12 was proposed to make them applicable. In addition, the item “assessment time” was divided into “assessment
13 time” and “assessment operators”, since most of the comments indicated that two distinct aspects were
14 addressed. Regarding the question about which tense would be the most appropriate, the majority of panel
15 members selected should/has/must sentences (15;63%) followed by past tense (4;17%), question sentences
16 (4;17%), and present tense (1;4%). On the basis of these data, the items on the checklist were modified.

17 Only one panel member proposed a new item regarding aspects that should be considered when
18 presenting IRT images in scientific articles. The core panel considered this item related to the category “body
19 position”, and it was therefore added to item 13. All decisions were communicated and submitted to the next
20 round.

21 22 Round 3

23
24 The items approved in the previous round were not included in this round. Thus, the decisions involved
25 approval (10; 92%) and exclusion (2; 8%). Two items (image background and assessment operators) did not
26 meet the criteria for inclusion and were subsequently excluded from the checklist. The responses from each
27 of the rounds, and the decisions made by the core panel are shown in table 1.

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Insert table 1 here

Final document

The final Checklist was structured as a list of 15 items (table 2) which should be marked “yes”, “no”, or “unclear”.

Insert Table 2 here

Discussion

In the absence of a consensus guideline to measure t_{sk} with IRT in sports and exercise medicine settings, a Delphi procedure was applied to develop a checklist for addressing the methodological aspects of data collection. The final checklist entitled “Thermographic Imaging in Sports and Exercise Medicine” (TISEM) contains 15 items which were approved by 24 world leading experts covering different fields of expertise (e.g. sport sciences, physiology, physiotherapy and medicine). The items encompassed the participants’ demographic information (items 1, 2 and 3), camera/room or environment setup (items 4, 5, 6, 7, 8, 9, 10 and 11) and recording/analysis (items 12, 13, 14 and 15) of t_{sk} using IRT. Considering the rapidly emerging use of IRT in sports and exercise medicine (Costello et al., 2013; Hildebrandt et al., 2010; Hildebrandt et al., 2012), TISEM addresses the relevant issues regarding the methodological aspects of data collection. The items from TISEM were organized to identify key methodological aspects regarding the use of infrared thermography in humans, especially when conducting research investigations. Similarly, it is proposed that this checklist could be used to collect t_{sk} data in a medical or clinical setting, since all items are equally relevant in non-sports and exercise medicine setting.

It is well established that age (Kenny and Journeay, 2010), sex (Gagnon and Kenny, 2012), body composition (Chudecka et al., 2015; Savastano et al., 2009), ethnicity (Maley et al., 2014), prevalence of smoking (Bornmyr and Svensson, 1991; Ijzerman et al., 2003), and others impact thermoregulation and t_{sk} .

1 As such, we advise physical characteristics (e.g. age, sex, body mass, height and body mass index), as well as
2 ethnicity and smoking history be reported. A number of studies have previously demonstrated the effect of
3 physical fitness on t_{sk} (Abate et al., 2013; Akimov and Son'kin, 2011; Chudecka and Lubkowska, 2010;
4 Formenti et al., 2013; Quesada et al., 2015a), therefore it is recommended that participants' physical activity
5 profile (e.g. frequency, duration, intensity, and activity description) and/or physical fitness (e.g. aerobic
6 capacity) be reported. Moreover, although the exact time frames of the impact of alcoholic beverages,
7 smoking, caffeine, large meals, ointments, cosmetics, showering and sunbathing may have on t_{sk} are not well
8 known (Fernández-Cuevas et al., 2015; Ring and Ammer, 2000), TISEM advises that users control these
9 variables in order to standardize the data collection procedures within and, where possible, between studies.
10 These variables should be confirmed verbally before the assessment and the use of any medicinal treatments
11 or drugs should be recorded. Additionally, t_{sk} is heavily influenced by extrinsic factors such as prior physical
12 activity (Bach et al., 2015a; Formenti et al., 2016; Tanda, 2016), as well as physical or medical treatments
13 such as massage (Adamczyk et al., 2016; IACT, 2002; Ring and Ammer, 2000), electrotherapy (Ring and
14 Ammer, 2000), ultrasound (Fernández-Cuevas et al., 2015), and by heat (Bach et al., 2015a; Gerrett et al.,
15 2014) or cold (Chesterton et al., 2002; Vellard and Arfaoui, 2016) exposure. Therefore, any intervention prior
16 to or during the assessment of t_{sk} should be recorded and detailed.

17 In relation to the experimental setup and reporting of environmental conditions it is important to
18 describe the ambient temperature and relative humidity where the assessment took place. Skin temperature is
19 influenced by the environment, especially if the skin is exposed (Bach et al., 2015b; Fernandes et al., 2014),
20 therefore TISEM recommends that mean ambient temperature ($^{\circ}\text{C}$; \pm standard deviation) and relative humidity
21 ($\%$; \pm standard deviation) be reported. Similarly, external factors, such as infrared radiation (e.g. electronic
22 devices, lightning) or airflow (e.g. under an air conditioning unit, ceiling fans, open windows/doors), are likely
23 to impact on and interact with t_{sk} . Therefore, TISEM suggests completing the assessment away from these
24 factors and reporting whether exposure to any of these conditions was unavoidable. Since a large number of
25 infrared camera models are currently available including cooled and uncooled cameras (Bach et al., 2015b;
26 Ring and Ammer, 2000), TISEM advises that manufacturer, model and accuracy of the camera is detailed.
27 Where available, it is also important to report when and where the camera was last calibrated. Regarding the

1 sensor stabilization of IRT cameras, depending on the technology, some models need to be turned on for some
2 time prior to the assessment in order to ensure consistent readings. To determine the time frame to address
3 this issue, TISEM recommends following manufacturer's guidelines or performing a quality assurance test, as
4 described by Ring et al. (2007b). In addition, when baseline measurements of t_{sk} are required, TISEM
5 recommends that an acclimation period be conducted in the examination room wherein ambient temperature
6 and humidity are regulated. Although previous research (Marins et al., 2014) showed that a 10-min acclimation
7 period is sufficient wherein differences between external and internal (testing room) temperature is less than
8 5 °C, the panelists agreed (63%) that 15 minutes should be the minimum recommended acclimation period
9 (Fernández-Cuevas et al., 2015; IACT, 2002; Ring and Ammer, 2000). However, it is important to note that
10 extreme temperature (e.g. more than 20 °C of difference between external temperature and room temperature),
11 can require more time to acclimatize because of the marked effects on t_{sk} (Taylor et al., 2014). Moreover, the
12 after-effect of the removal of clothing should be considered given that it can influence t_{sk} up to 20 minutes
13 after undressing (Vainer, 2001). Therefore, the time used must be determined in accordance with the objective
14 of the study/assessment, the ROI (open skin or that under clothes) and the environmental conditions. The
15 checklist also advises that the distance between the skin and camera and percentage of the region of interest
16 within the image should be detailed in order to guarantee reproducibility across studies. As demonstrated by
17 Ammer (2015), when the camera is placed close to the region of interest, the field of view provides a more
18 detailed temperature information. In addition, the camera should be positioned perpendicular to the region of
19 interest, otherwise the assessment can result in a critical loss of information (Tkacova et al., 2010).

20 While controversy in the literature regarding emissivity exists (Sanchez-Marin et al., 2009; Steketee,
21 1973), 96% of experts strongly agreed that an emissivity of 0.98 (ϵ) should be used for clean dry skin. Due to
22 circadian rhythm (Costa et al., 2015; Marins et al., 2015), t_{sk} is likely to change during the course of the day,
23 therefore, when individuals are assessed over multiple days or when comparisons between participants are
24 made at different time of the day, the time of day at which the images were recorded should be reported. The
25 panel of experts agreed that the use of a standardized body position as well as the selection of regions of
26 interest should be sufficiently described to ensure the reliability and reproducibility of the data. Moreover, the
27 presentation of a visual example is recommended as it may add important information or representation about

1 how regions of interests were defined. Since water impacts on emissivity (Fernández-Cuevas et al., 2015) in
2 some situations, particularly during cold water immersion or exercising in water, the experts agreed (92%)
3 that the skin could be dried. However, the method of drying (e.g. towel patting) should be clearly described
4 and reported. Recent recommendations by Seixas et al. (2014) suggested that the skin should be carefully
5 dried with a microfiber towel to limit irritation of the skin (that may occur with more abrasive fabrics). At the
6 same time, it is quantitatively demonstrated that moisturizing the skin affects t_{sk} contrast and may be used to
7 enhance the surface vessels thermal pattern (Vainer, 2001). In addition, a suitable practice within extremity
8 cooling studies is to use a thin plastic bag to prevent the extremity from becoming wet (Eglin et al., 2013;
9 Maley et al., 2014). Finally, the method of analysis including the software used and whether or not the analysis
10 was completed manually or automatically should be described. Similarly, the method employed to calculate
11 the final temperature value (e.g. average, median, maximum or minimum) should be clear described. As much
12 information about the process itself should be provided so that others can replicate the findings if needed.

13 The number of experts who have completed the process (24) is greater than other studies using a similar
14 methodological design (Boulkedid et al., 2011). In addition the experience of the panel experts (median = 8
15 years; range 3 to 32 years and publications median = 8; range 3 to 80), the multiple nationalities (13) and
16 different professional backgrounds (4) of the panelists allow a thorough and broad analysis of the use of IRT,
17 illustrating the positive characteristics of the current Delphi procedure (Boulkedid et al., 2011; Hsu and
18 Sandford, 2007). However, the study is not without limitations. While the Delphi panel consisted of experts
19 representing a range of disciplines, the process would have benefitted from a greater inclusion of practitioners
20 that use IRT daily. In addition, the invitation of panelists could have caused selection bias. Because the
21 recommendations are primarily based on experts' opinion, users should take into account the possibility of the
22 checklist does not address all issues. In this sense, additional items may be required since scientific evidence
23 has become available.

24

25 **Conclusion**

26

1 We have provided a checklist with 15 items directed at standardizing the assessment of t_{sk} using IRT
2 for a wide array of end-users including practitioners, sports scientists, exercise physicians, medical
3 professionals and others. This checklist is not limited to this setting, and may also be used in others fields such
4 occupational medicine and public health. It is intended that the TISEM can also be applied to evaluate bias in
5 thermographic studies, and to guide practitioners in the use of this technique.

6 **Acknowledgements**

8 National Council of Scientific and Technologic Development (CNPq) for the PhD Scholarship number
9 205815/2014-6 for DGM and 234243/2014-7 for CJB. Participating societies included the European European
10 Association of Thermology (EAT), American Academy of Thermology (AAT), and Polish Society of Medical
11 Thermography.

13 **Competing interests**

14 None declared.

16 **References**

- 17
- 18 Abate, M., Di Carlo, L., Di Donato, L., Romani, G., Merla, A., 2013. Comparison of cutaneous termic
19 response to a standardised warm up in trained and untrained individuals. *J Sports Med Phys Fitness* 53, 209-
20 215.
- 21 Adamczyk, J.G., Boguszewski, D., Siewierski, M., 2014. Thermographic evaluation of lactate level in
22 capillary blood during post-exercise recovery. *Kinesiology* 46, 186-193.
- 23 Adamczyk, J.G., Krasowska, I., Boguszewski, D., Reaburn, P., 2016. The use of thermal imaging to assess
24 the effectiveness of ice massage and cold-water immersion as methods for supporting post-exercise recovery.
25 *J Therm Biol* 60, 20-25.
- 26 Akimov, E., Son'kin, V., 2011. Skin temperature and lactate threshold during muscle work in athletes. *Hum*
27 *Physiol* 37, 621-628.

1 Ammer, K., 2003. Need for standardisation of measurements in thermal imaging. *Thermography and Lasers*
2 *in Medicine*. Akademickie Centrum Graficzno-Marketigowe Lodart SA, Lodz, 13-18.

3 Ammer, K., 2008. The Glamorgan Protocol for recording and evaluation of thermal images of the human
4 body. *Thermol Int* 18, 125-129.

5 Ammer, K., 2015. Do we need reference data of local skin temperatures? *Thermol Int* 25, 45-47.

6 Bach, A.J., Stewart, I.B., Disher, A.E., Costello, J.T., 2015a. A comparison between conductive and infrared
7 devices for measuring mean skin temperature at rest, during exercise in the heat, and recovery. *PLoS One* 10,
8 e0117907.

9 Bach, A.J.E., Stewart, I.B., Minett, G.M., Costello, J.T., 2015b. Does the technique employed for skin
10 temperature assessment alter outcomes? A Systematic Review. *Physiol Meas* 36 (9), 27-51.

11 Bahl, J.S., Dollman, J., Davison, K., 2016. The development of a subjective assessment framework for
12 individuals presenting for clinical exercise services: A Delphi study. *J Sci Med Sport* 19 (11), 872-876.

13 Bornmyr, S., Svensson, H., 1991. Thermography and laser- Doppler flowmetry for monitoring changes in
14 finger skin blood flow upon cigarette smoking. *Clin Physiol* 11 (2), 135-141.

15 Boulkedid, R., Abdoul, H., Loustau, M., Sibony, O., Alberti, C., 2011. Using and reporting the Delphi method
16 for selecting healthcare quality indicators: a systematic review. *PLoS One* 6, e20476.

17 Chesterton, L.S., Foster, N.E., Ross, L., 2002. Skin temperature response to cryotherapy. *Phys Med Rehabil*
18 83 (4), 543-549.

19 Chipchase, L., Schabrun, S., Cohen, L., Hodges, P., Ridding, M., Rothwell, J., Taylor, J., Ziemann, U., 2012.
20 A checklist for assessing the methodological quality of studies using transcranial magnetic stimulation to study
21 the motor system: an international consensus study. *Clin Neurophysiol* 123 (9), 1698-1704.

22 Chudecka, M., Lubkowska, A., 2010. Temperature changes of selected body's surfaces of handball players in
23 the course of training estimated by thermovision, and the study of the impact of physiological and
24 morphological factors on the skin temperature. *J Therm Biol* 35, 379-385.

25 Chudecka, M., Lubkowska, A., Leźnicka, K., Krupecki, K., 2015. The Use of Thermal Imaging in the
26 Evaluation of the Symmetry of Muscle Activity in Various Types of Exercises (Symmetrical and
27 Asymmetrical). *J Hum Kinet* 49 (11), 141-147.

1 Costa, C.M., Sillero-Quintana, M., Pinonosa Cano, S., Moreira, D.G., Brito, C.J., Fernandes, A.A., Pussieldi,
2 G.A., Marins, J.C., 2015. Daily oscillations of skin temperature in military personnel using thermography. *J*
3 *R Army Med Corps* 162 (5), 335-342.

4 Costello, J., Stewart, I.B., Selfe, J., Karki, A.I., Donnelly, A.E., 2013. Use of thermal imaging in sports
5 medicine research: a short report: short article. *Int SportMed Journal* 14 (2), 94-98.

6 Costello, J.T., Culligan, K., Selfe, J., Donnelly, A.E., 2012a. Muscle, skin and core temperature after $-110\text{ }^{\circ}\text{C}$
7 cold air and $8\text{ }^{\circ}\text{C}$ water treatment. *PloS One* 7, e48190.

8 Costello, J.T., McInerney, C.D., Bleakley, C.M., Selfe, J., Donnelly, A.E., 2012b. The use of thermal imaging
9 in assessing skin temperature following cryotherapy: a review. *J Therm Biol* 37, 103-110.

10 Dalkey, N., Helmer, O., 1963. An experimental application of the Delphi method to the use of experts. *Manag*
11 *Sci* 9 (3), 458-467.

12 Eglin, C.M., Golden, F.S., Tipton, M.J., 2013. Cold sensitivity test for individuals with non-freezing cold
13 injury: the effect of prior exercise. *Extrem Physiol Med* 2 (1), 1-8.

14 Fernandes, A.A., Amorim, P.R.S., Brito, C.J., Moura, A.G., Moreira, D.G., Costa, C.M.A., Sillero-Quintana,
15 M., Marins, J.C.B., 2014. Measuring skin temperature before, during and after exercise: a comparison of
16 thermocouples and infrared thermography. *Physiol Meas* 35 (2), 189-203.

17 Fernandes, A.A., Moreira, D.G., Brito, C.J., da Silva, C.D., Sillero-Quintana, M., Pimenta, E.M., Bach, A.J.,
18 Garcia, E.S., Marins, J.C.B., 2016. Validity of inner canthus temperature recorded by infrared thermography
19 as a non-invasive surrogate measure for core temperature at rest, during exercise and recovery. *J Therm Biol*
20 62, 50-55.

21 Fernández-Cuevas, I., Marins, J.C.B., Lastras, J.A., Carmona, P.M.G., Cano, S.P., García-Concepción, M.Á.,
22 Sillero-Quintana, M., 2015. Classification of factors influencing the use of infrared thermography in humans:
23 a review. *Infrared Phys Techn* 71, 28-55.

24 Ferreira, J.J., Mendonca, L.C., Nunes, L.A., Andrade Filho, A.C., Rebelatto, J.R., Salvini, T.F., 2008.
25 Exercise-associated thermographic changes in young and elderly subjects. *Ann Biomed Eng* 36, 1420-1427.

1 Formenti, D., Ludwig, N., Gargano, M., Gondola, M., Dellerma, N., Caumo, A., Alberti, G., 2013. Thermal
2 imaging of exercise-associated skin temperature changes in trained and untrained female subjects. *Ann*
3 *Biomed Eng* 41, 863-871.

4 Formenti, D., Ludwig, N., Trecroci, A., Gargano, M., Michielon, G., Caumo, A., Alberti, G., 2016. Dynamics
5 of thermographic skin temperature response during squat exercise at two different speeds. *J Therm Biol* 59,
6 58-63.

7 Fournet, D., Ross, L., Voelcker, T., Redortier, B., Havenith, G., 2013. Body mapping of thermoregulatory and
8 perceptual responses of males and females running in the cold. *J Therm Biol* 38, 339-344.

9 Gagnon, D., Kenny, G.P., 2012. Does sex have an independent effect on thermoeffector responses during
10 exercise in the heat? *J Physiol* 590, 5963-5973.

11 Gerrett, N., Ouzzahra, Y., Coleby, S., Hobbs, S., Redortier, B., Voelcker, T., Havenith, G., 2014. Thermal
12 sensitivity to warmth during rest and exercise: a sex comparison. *Eur J Appl Physiol* 114, 1451-1462.

13 Gerrett, N., Ouzzahra, Y., Redortier, B., Voelcker, T., Havenith, G., 2015. Female thermal sensitivity to hot
14 and cold during rest and exercise. *Physiol. Behav* 152, 11-19.

15 Hadžić, V., Širok, B., Malneršič, A., Čoh, M., 2015. Can infrared thermography be used to monitor fatigue
16 during exercise? A case study. *J Sport Health* 1-4.

17 Hani, H.A.-N., Jerrold, S.P., Michael, S.L., Lee, S.B., 2012. The use of thermal infra-red imaging to detect
18 delayed onset muscle soreness. *J Vis Exp* 59, e3551.

19 Hildebrandt, C., Raschner, C., Ammer, K., 2010. An overview of recent application of medical infrared
20 thermography in sports medicine in Austria. *Sensors* 10, 4700-4715.

21 Hildebrandt, C., Zeilberger, K., Ring, E.F.J., Raschner, C., 2012. The application of medical Infrared
22 Thermography in sports medicine, in: Zaslav, K.R. (Ed.), *An International Perspective on Topics in Sports*
23 *Medicine and Sports Injury*. InTech, p. 534.

24 Hsu, C.-C., Sandford, B.A., 2007. The Delphi technique: making sense of consensus. *Practical Assessment,*
25 *Research & Evaluation* 12, 1-8.

26 IACT, 2002. *Thermology Guidelines. Standards and protocols in Clinical Thermography Imaging.*
27 <http://www.iact-org.org/professionals/thermog-guidelines.html>. Accessed 01 Feb 2015.

1 Ijzerman, R.G., Serne, E.H., van Weissenbruch, M.M., de Jongh, R.T., Stehouwer, C.D., 2003. Cigarette
2 smoking is associated with an acute impairment of microvascular function in humans. *Clin Sci* 104, 247-252.

3 ISO, 2004. Ergonomics-Evaluation of thermal strain by physiological measurements. International
4 Standardization Organization Geneva.

5 Kelley, K., Clark, B., Brown, V., Sitzia, J., 2003. Good practice in the conduct and reporting of survey
6 research. *Int J Qual Health Care* 15, 261-266.

7 Kenny, G.P., Journeay, W.S., 2010. Human thermoregulation: separating thermal and nonthermal effects on
8 heat loss. *Front Biosci* 15, 259-290.

9 Maley, M.J., Eglin, C.M., House, J.R., Tipton, M.J., 2014. The effect of ethnicity on the vascular responses
10 to cold exposure of the extremities. *Eur J Appl Physiol* 114, 2369-2379.

11 Marins, J.C.B., Formenti, D., Costa, C.M.A., Fernandes, A.d.A., Sillero-Quintana, M., 2015. Circadian and
12 gender differences in skin temperature in militaries by thermography. *Infrared Phys Techn* 71, 322-328.

13 Marins, J.C.B., Moreira, D.G., Cano, S.P., Sillero-Quintana, M., Soares, D.D., Fernandes, A.A., Silva, F.S.,
14 Costa, C.M.A., Amorim, P.R.S., 2014. Time required to stabilize thermographic images at rest. *Infrared Phys*
15 *Technol* 65, 30-35.

16 Mercer, J.B., Ring, E.F.J., 2009. Fever screening and infrared thermal imaging: concerns and guidelines.
17 *Thermol Int* 19, 67-69.

18 Moher, D., Jones, A., Lepage, L., Group, C., 2001. Use of the CONSORT statement and quality of reports of
19 randomized trials: a comparative before-and-after evaluation. *JAMA* 285, 1992-1995.

20 Moseley, A.M., Herbert, R.D., Maher, C.G., Sherrington, C., Elkins, M.R., 2011. Reported quality of
21 randomized controlled trials of physiotherapy interventions has improved over time. *J Clin Epidemiol* 64, 594-
22 601.

23 Priego-Quesada, J.I., Carpes, F.P., Bini, R.R., Palmer, R.S., Pérez-Soriano, P., de Anda, R.M.C.O., 2015a.
24 Relationship between skin temperature and muscle activation during incremental cycle exercise. *J Therm Biol*
25 48, 28-35.

1 Priego-Quesada, J.I., Guillamón, N.M., de Anda, R.M.C.O., Psikuta, A., Annaheim, S., Rossi, R.M., Salvador,
2 J.M.C., Pérez-Soriano, P., Palmer, R.S., 2015b. Effect of perspiration on skin temperature measurements by
3 infrared thermography and contact thermometry during aerobic cycling. *Infrared Phys Technol* 72, 68-76.

4 Ring, E., Ammer, K., Wiecek, B., Plassmann, P., Jones, C., Jung, A., Murawski, P., 2007a. Quality assurance
5 for thermal imaging systems in medicine. *Thermol Int* 17, 103-106.

6 Ring, E.F., Ammer, K., 2012. Infrared thermal imaging in medicine. *Physiol Meas* 33, R33-46.

7 Ring, E.F.J., Ammer, K., 2000. The Technique of Infra red Imaging in Medicine. *Thermol Int* 10, 7-14.

8 Ring, F.J., Ammer, K., Wiecek, B., Plassmann, P., Jones, C.D., Jung, A., Murawski, P., 2007b. Quality
9 assurance of thermal imaging systems in medicine. *Thermol Int* 16, 10-15.

10 Robinson, L.J., Law, J.M., Symonds, M.E., Budge, H., 2016. Brown adipose tissue activation as measured by
11 infrared thermography by mild anticipatory psychological stress in lean healthy females. *Exp Physiol* 101,
12 549–557.

13 Sanchez-Marin, F.J., Calixto-Carrera, S., Villaseñor-Mora, C., 2009. Novel approach to assess the emissivity
14 of the human skin. *J Biomed Opt* 14, 024006.

15 Savastano, D.M., Gorbach, A.M., Eden, H.S., Brady, S.M., Reynolds, J.C., Yanovski, J.A., 2009. Adiposity
16 and human regional body temperature. *Am J Clin Nutr* 90, 1124-1131.

17 Schulz, K.F., Altman, D.G., Moher, D., 2010. CONSORT 2010 statement: updated guidelines for reporting
18 parallel group randomised trials. *BMC medicine* 340, c332.

19 Schwartz, R., Elliott, R., Goldberg, G., 2006. Guidelines for neuromusculoskeletal thermography. *Thermol*
20 *Int* 16, 5-9.

21 Schwartz, R.G., O'Young, B., Getson, P., Govindan, S., Uricchio, J., Bernton, T., Brioschi, M., Zhang, H.-
22 Y., 2015. Guidelines for neuromusculoskeletal infrared thermography sympathetic skin response (ssr) studies.
23 *Pan Am J Med Thermol* 2, 35-43.

24 Seixas, A., Gonjo, T., Vardasca, R., Gabriel, J., Fernandes, R., Vilas-Boas, J., 2014. A preliminary study on
25 the relationship between energy expenditure and skin temperature in swimming, 12th International Conference
26 on Quantitative InfraRed Thermography, Bordeaux, France, pp. 90-97.

1 Selfe, J., Alexander, J., Costello, J.T., May, K., Garratt, N., Atkins, S., Dillon, S., Hurst, H., Davison, M.,
2 Przybyla, D., 2014. The effect of three different (-135 C) whole body cryotherapy exposure durations on elite
3 rugby league players. *PLoS One* 9, e86420.

4 Selfe, J., Hardaker, N., Thewlis, D., Karki, A., 2006. An accurate and reliable method of thermal data analysis
5 in thermal imaging of the anterior knee for use in cryotherapy research. *Arch Phys Med Rehabil* 87, 1630-
6 1635.

7 Selfe, J., Sutton, C., Hardaker, N., Greenhalgh, S., Karki, A., Dey, P., 2010. Cold Females, A Distinct Group
8 Of Patellofemoral Pain Syndrome Patients? *J Orthop Sports Physical* 40, A42.

9 Silva, Y.A., Santos, B.H., Andrade, P.R., Santos, H.H., Moreira, D.G., Sillero-Quintana, M., Ferreira, J.J.A.,
10 2017. Skin temperature changes after exercise and cold water immersion. *Sport Sci Health* 13, 195-202.

11 Snyder, K.R., Evans, T.A., Neibert, P.J., 2014. Developing a Framework for Ankle Function: A Delphi Study.
12 *J Athl Train* 49, 747-757.

13 Steketee, J., 1973. Spectral emissivity of skin and pericardium. *Phys Med Biol* 18, 686-694.

14 Steurer, J., 2011. The Delphi method: an efficient procedure to generate knowledge. *Skeletal Radiology* 40,
15 959-961.

16 Tanda, G., 2016. Skin temperature measurements by infrared thermography during running exercise. *Exp*
17 *Therm Fluid Sci* 71, 103-113.

18 Taylor, N.A., Tipton, M.J., Kenny, G.P., 2014. Considerations for the measurement of core, skin and mean
19 body temperatures. *J Therm Biol* 46, 72-101.

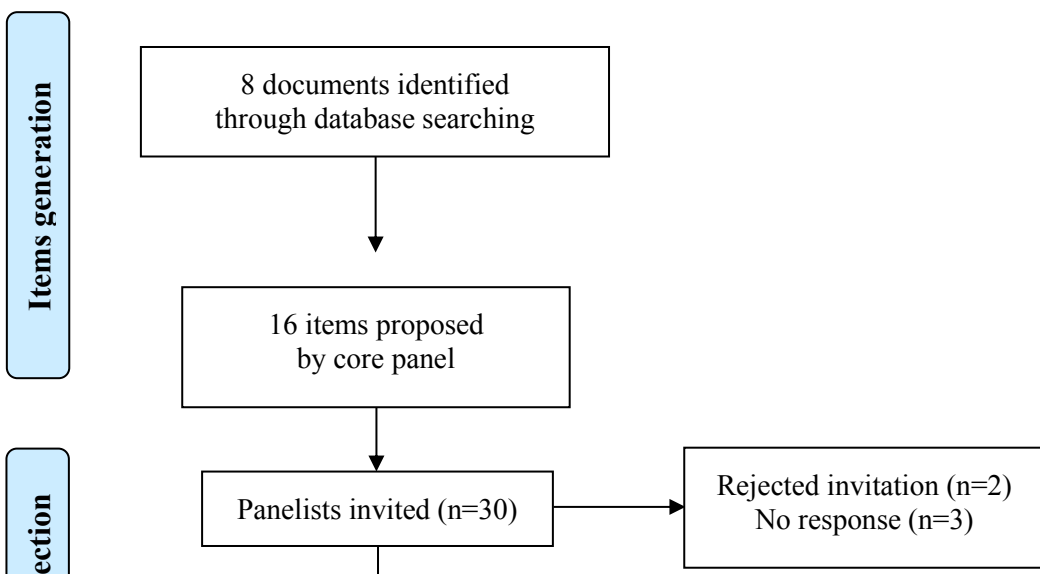
20 Tkacova, M., Hudak, R., Foffova, P., Zivcak, J., 2010. An importance of camera - subject distance and angle
21 in musculoskeletal application of medical thermography. *Acta Electrotechnica et Informatica* 10, 57-60.

22 Vainer, B.G., 2001. Treated-skin temperature regularities revealed by IR thermography, *Aerospace/Defense*
23 *Sensing, Simulation, and Controls*. International Society for Optics and Photonics, pp. 470-481.

24 Vainer, B.G., 2005. FPA-based infrared thermography as applied to the study of cutaneous perspiration and
25 stimulated vascular response in humans. *Phys Med Biol* 50, R63-94.

26 Vellard, M., Arfaoui, A., 2016. Detection by Infrared Thermography of the Effect of Local Cryotherapy
27 Exposure on Thermal Spreadin Skin. *J Imaging* 2 (2), 1-9.

1 Verhagen, A.P., de Vet, H.C., de Bie, R.A., Kessels, A.G., Boers, M., Bouter, L.M., Knipschild, P.G., 1998.
2 The Delphi list: a criteria list for quality assessment of randomized clinical trials for conducting systematic
3 reviews developed by Delphi consensus. *J Clin Epidemiol* 51, 1235-1241.
4 Whiting, P., Rutjes, A.W., Reitsma, J.B., Bossuyt, P.M., Kleijnen, J., 2003. The development of QUADAS: a
5 tool for the quality assessment of studies of diagnostic accuracy included in systematic reviews. *BMC Med*
6 *Res Methodol* 3, 3-25.



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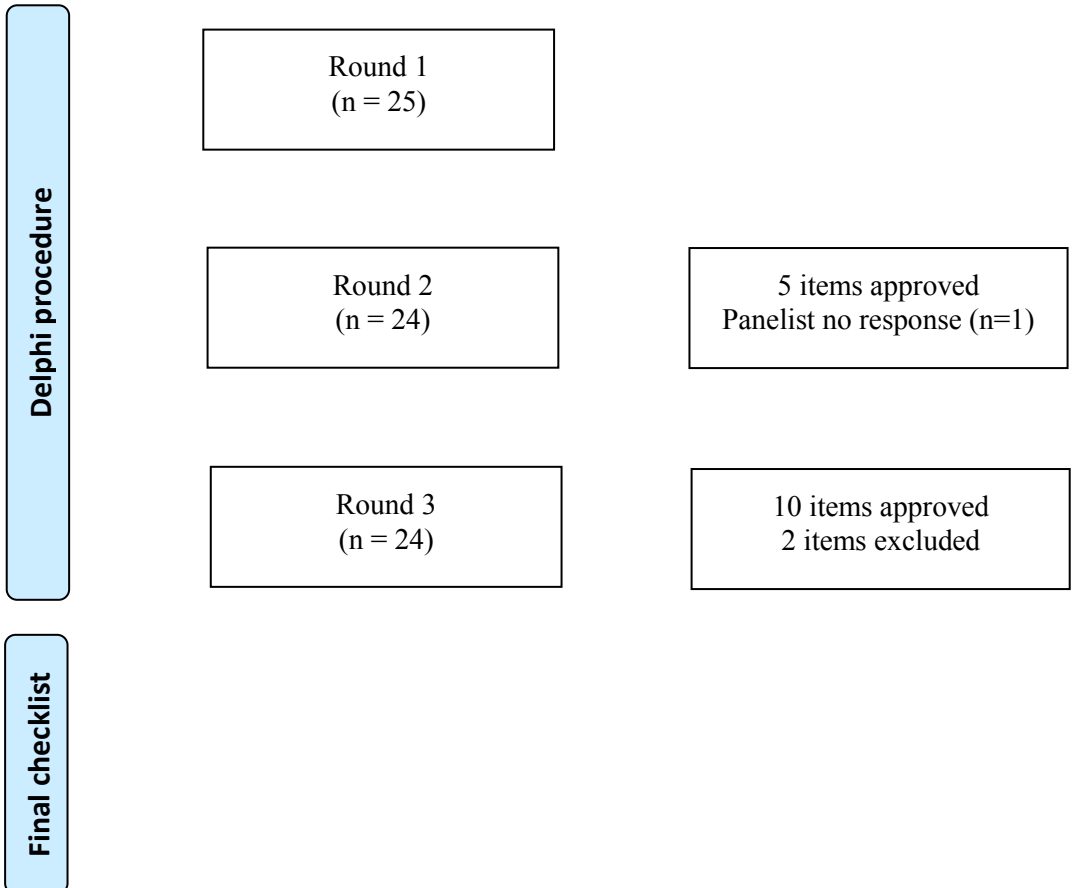


Figure 1. Flowchart of the development process (n = number of panelists).

Table 1. Panelists' responses through the rounds with core panel decisions for each item.

| Item | Round 1 (n=25) | | | | | | Round 2 (n=24) | | | | | | Round 3 (n=24) | | | | | |
|---|----------------|---------|--------|--------|--------|--------------------|----------------|--------|--------|--------|--------|--------------------|----------------|--------|--------|--------|-------|-------------------|
| | SA | MA | N | MD | SD | SA+MA: Decision | SA | MA | N | MD | SD | SA+MA: Decision | SA | MA | N | MD | SD | SA+MA: Decision |
| Individual data ¹ | 17 (68) | 4 (16) | 2 (8) | 2 (8) | 0 (0) | 21 (84): modify | 19 (79) | 4 (17) | 1 (4) | 0 (0) | 0 (0) | 23 (96): modify | 20 (83) | 4 (17) | 0 (0) | 0 (0) | 0 (0) | 24 (100): approve |
| Previous instructions ² | 19 (76) | 4 (16) | 1 (4) | 1 (4) | 0 (0) | 23 (92): modify | 22 (92) | 2 (8) | 0 (0) | 0 (0) | 0 (0) | 24 (100): rephrase | 19 (79) | 4 (17) | 1 (4) | 0 (0) | 0 (0) | 23 (96): approve |
| Extrinsic factors ³ | 22 (88) | 2 (8) | 1 (4) | 0 (0) | 0 (0) | 24 (96): rephrase | 22 (92) | 1 (4) | 1 (4) | 0 (0) | 0 (0) | 23 (96): approve | a | a | a | a | a | a |
| Environmental condition ⁴ | 12 (48) | 4 (16) | 2 (8) | 4 (16) | 3 (12) | 16 (64): modify | 20 (83) | 3 (13) | 1 (4) | 0 (0) | 0 (0) | 23 (96): approve | a | a | a | a | a | a |
| Environmental setup ⁵ | 18 (72) | 7 (28) | 0 (0) | 0 (0) | 0 (0) | 25 (100): rephrase | 19 (79) | 4 (17) | 0 (0) | 1 (4) | 0 (0) | 23 (96): rephrase | 18 (75) | 2 (8) | 1 (4) | 3 (13) | 0 (0) | 20 (83): approve |
| Equipment ⁶ | 13 (52) | 11 (44) | 0 (0) | 1 (4) | 0 (0) | 24 (96): rephrase | 19 (79) | 5 (21) | 0 (0) | 0 (0) | 0 (0) | 24 (100): rephrase | 15 (63) | 7 (29) | 0 (0) | 2 (8) | 0 (0) | 22 (92): approve |
| Image background | 11 (44) | 7 (28) | 3 (12) | 3 (12) | 1 (4) | 18 (72): modify | 17 (71) | 2 (8) | 4 (17) | 1 (4) | 0 (0) | 19 (79): modify | 13 (54) | 3 (13) | 6 (25) | 2 (8) | 0 (0) | 16 (67): exclude |
| Aclimation ⁷ | 12 (48) | 8 (32) | 0 (0) | 3 (12) | 2 (8) | 20 (80): modify | 16 (67) | 5 (21) | 1 (4) | 2 (8) | 0 (0) | 21 (88): rephrase | 16 (67) | 4 (17) | 1 (4) | 3 (13) | 0 (0) | 20 (83): approve |
| Camera preparation ⁸ | 15 (60) | 6 (24) | 1 (4) | 2 (8) | 1 (4) | 21 (84): modify | 19 (79) | 3 (13) | 1 (4) | 1 (4) | 0 (0) | 22 (92): rephrase | 17 (71) | 4 (17) | 0 (0) | 3 (13) | 0 (0) | 21 (88): approve |
| Image recording ⁹ | 13 (52) | 8 (32) | 2 (8) | 2 (8) | 0 (0) | 21 (84): rephrase | 19 (79) | 3 (13) | 0 (0) | 2 (8) | 0 (0) | 22 (92): modify | 16 (67) | 6 (25) | 0 (0) | 2 (8) | 0 (0) | 22 (92): approve |
| Camera position ¹⁰ | 22 (88) | 2 (8) | 1 (4) | 0 (0) | 0 (0) | 24 (96): rephrase | 23 (96) | 1 (4) | 0 (0) | 0 (0) | 0 (0) | 24 (100): approve | a | a | a | a | a | a |
| Emissivity ¹¹ | 17 (68) | 5 (20) | 2 (8) | 1 (4) | 0 (0) | 22 (88): rephrase | 23 (96) | 1 (4) | 0 (0) | 0 (0) | 0 (0) | 24 (100): approve | a | a | a | a | a | a |
| Assessment time ¹² | 12 (48) | 8 (32) | 1 (4) | 3 (12) | 1 (4) | 20 (80): exclude# | 9 (38) | 5 (21) | 3 (13) | 4 (17) | 3 (13) | 14 (58): divide | 17 (71) | 4 (17) | 2 (8) | 1 (4) | 0 (0) | 21 (88): approve |
| Assessment operators* | - | - | - | - | - | - | - | - | - | - | - | - | 12 (50) | 5 (21) | 6 (25) | 0 (0) | 1 (4) | 17 (71): exclude |
| Body position ¹³ | 15 (60) | 6 (24) | 2 (8) | 1 (4) | 1 (4) | 21 (84): rephrase | 18 (75) | 5 (21) | 0 (0) | 1 (4) | 0 (0) | 23 (96): rephrase | 17 (71) | 6 (25) | 0 (0) | 1 (4) | 0 (0) | 23 (96): approve |
| Method of drying the skin ¹⁴ | 16 (64) | 5 (20) | 2 (8) | 1 (4) | 1 (4) | 21 (84): exclude# | 13 (54) | 4 (17) | 5 (21) | 1 (4) | 1 (4) | 17 (71): modify | 19 (79) | 3 (13) | 2 (8) | 0 (0) | 0 (0) | 22 (92): approve |
| Image evaluation ¹⁵ | 22 (88) | 1 (4) | 2 (8) | 0 (0) | 0 (0) | 23 (92): rephrase | 24 (100) | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 24 (100): approve | a | a | a | a | a | a |

Data expressed as number (percentage of responses); Superscript numbers refer to final checklist (see table 2); a = item approved in the previous round; * The item was proposed by the division of item “assessment time” in the third round; # The item was presented again in a different approach; SA = Strongly agree; MA = Moderately agree; N = Neutral; MD = Moderately disagree; SD = Strongly disagree; SA+MD = sum of the responses of strongly agree and moderately agree.

Table 2. Thermographic imaging in sports and exercise medicine (TISEM).

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| <p>1) The relevant individual data of the participants must be provided. Note: These could include, but are not limited to, age, sex, body mass, height, body mass index, ethnicity and whether they are smokers or not. An indication of physical activity profile (e.g. frequency, duration, intensity, and activity description) should be reported.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>2) Participants should be instructed to avoid alcohol beverages, smoking, caffeine, large meals, ointments, cosmetics and showering for four hours before the assessment. Also, sunbathing (e.g. UV sessions or direct sun without protection) should be avoided before the assessment. Note: This should be confirmed verbally before the assessment. The use of any medicinal treatments or drugs should be recorded. Any condition that could not be avoided should be reported.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>3) Extrinsic factors affecting skin temperature (e.g. physical activity prior to the assessment, massage, electrotherapy, ultrasound, heat or cold exposure, cryotherapy) should be clearly described.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>4) Ambient temperature and relative humidity of the location where the assessment took place must be recorded and reported as mean \pm standard deviation.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>5) The assessment should be completed away from any source of infrared radiation (e.g. electronic devices, lightning) or airflow (e.g. under an air conditioning unit). Note: Any condition that could not be controlled should be reported.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>6) The manufacturer, model and accuracy of the camera used should be provided. Note: When available it is recommended to provide the maintenance information of the equipment (e.g. when and where it was completed the last calibration).</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>7) An acclimation period in the examination room should be completed. Note: This item is only applicable for initial baseline measurements or basal analysis.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>8) If necessary the camera should be turned on for some time prior to the test to allow sensor stabilization following the manufacturer's guidelines.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>9) Conditions of image recording such as mean distance between object and camera, percentage of the region of interest within the image should be detailed.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>10) The camera should be positioned perpendicular to the region of interest.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>11) Emissivity settings of the camera must be reported. Note: 0.98 of emissivity is suggested for a dry clean skin surface.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>12) The time of day at which the images were taken should be reported.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>13) The standard body position of the subject and the regions of interest must be well described and appropriately selected. A visual example (with temperature scale presented and scale of colors properly configured) is recommended.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>14) If the skin is dried (e.g. to remove surface water), the drying method should be clearly described.</p> <p><input type="checkbox"/> Yes <input type="checkbox"/> No <input type="checkbox"/> Unclear</p> |
| <p>15) The evaluation of thermograms and collection of temperature from the software should be clearly described.</p> |

Yes No Unclear