1	The effect of surface properties of polycrystalline, single phase metal coatings on bacterial
2	retention
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#### 26 ABSTRACT

27 In the food industry microbial contamination of surfaces can result in product spoilage which 28 may lead to potential health problems of the consumer. Surface properties can have a 29 substantial effect on microbial retention. The surface characteristics of chemically different 30 coatings (Cu, Ti, Mo, Ag, Fe) were defined using white light profilometry (micro-topography 31 and surface features), atomic force microscopy (nano-topography) and physicochemical 32 measurements. The Ag coating had the greatest topography measurements and Fe and Mo the 33 least. Mo was the most hydrophobic coating (lowest  $\gamma_{AB}$ ,  $\gamma^+$ ,  $\gamma^-$ ) whilst Ag was the most hydrophilic (greatest  $\gamma_{AB}$ ,  $\gamma^+$ ,  $\gamma^-$ ). The physicochemical results for the Fe, Ti and Cu coatings 34 35 was found to lie between those of the Ag and Mo coatings. Microbiological retention assays 36 were carried out using Listeria monocytogenes, Escherichia coli and Staphylococcus aureus in 37 order to determine how surface properties influenced microbial retention. It was found that 38 surface chemistry had an effect on microbial retention, whereas the shape of the surface 39 features and nano-topography did not. L. monocytogenes and S. aureus retention to the surfaces 40 were mostly affected by surface micro-topography, whereas retention of E. coli to the coatings was mostly affected by the coating physicochemistry. There was no trends observed between 41 42 the bacterial cell surface physicochemistry and the coating physicochemistry.

This work highlights that different surface properties may be linked to factors affecting microbial retention hence, the use of surface chemistry, topography or physicochemical factors alone to describe microbial retention to a surface is no longer adequate. Moreover, the effects of surface parameters on microbial retention should be considered individually for each bacterial genus.

*Keywords:* Food; Metals; Surface topography; Bacterial retention; Physicochemistry;
Chemistry

## 51 1. Introduction

52 A major concern in the food processing industry is to ensure that food produce does not become spoiled or cause disease in humans as a result of bacterial contamination. Bacteria naturally 53 54 occur in raw materials, but may (re)contaminate food products due to their residing in food 55 processing equipment (Carrasco et al., 2012; Skovager et al., 2012). Stainless steel, a Fe alloy 56 usually containing Cr in excess of 11% but less than 30% (Adams, 1983) is regularly used in 57 the manufacturer of food processing equipment since it is relatively easy to clean, mechanically 58 strong, is relatively easy to fabricate and is corrosion resistant. However, it has been shown 59 that even with good manufacturing processes that include stringent cleaning and sanitation 60 procedures, microorganisms can remain in a viable state on surfaces (Marouani-Gadri et al., 61 2010), or once established, remain as persistent strains within the industrial workplace 62 (Carpentier and Cerf, 2011).

63 Bacterial retention has been shown to be affected by surface roughness and topography 64 (Whitehead et al., 2011; Wickens et al., 2012; 2014). The influence of surface topography on 65 bacterial adhesion is an important issue, and it has been suggested that surface roughness must 66 not exceed  $R_a$  values of 0.8  $\mu$ m (Flint et al., 2000). Alterations in chemical composition (Ma et 67 al., 2008) and/or surface physicochemistry, may affect bacterial attachment and retention onto a surface (Abban et al., 2012; Skovager et al., 2012). Thus, the study of microbial retention on 68 69 surfaces is important to enable informed decisions to be made regarding the design and use of 70 materials in the food industry.

Magnetron sputtering, a physical vapour deposition process, can be used to deposit thin metal films or alloys onto chosen substrates (Whitehead et al., 2004). The metals used in this study (Ti, Ag, Cu, Fe, Mo) were chosen for either their mechanical properties and/or their potential antimicrobial properties. Ti has good mechanical properties and corrosion resistance (Zaffe et al., 2003). Fe is the key component of stainless steel. Cu has been shown to inhibit growth of bacteria (Champagne et al., 2013). Ag has long been known for its antibacterial properties
(Skovager et al., 2013). It is also lubricious and enables coatings to be self-lubricating (Kelly
et al., 2009). Mo forms hard, stable carbides and is sometimes used in high strength steel alloys
and it has also been suggested to have antimicrobial properties (Tetault et al., 2012; Zollfrank
et al., 2012).

Three microorganisms where used in this study, *Listeria monocytogenes, Escherichia coli* and *Staphylococcus aureus* since they have a potential disease burden associated with their contamination of food contact surfaces. The aim of this work was to determine the effect of surface properties of single phase, metal coatings to determine if a single surface parameter (chemistry, nano- or micro-topography, surface features or physicochemistry) had the greatest influence on bacterial retention.

## 87 2. Methods and Materials

## 88 2.1. Coating production and characterisation

89 In order to produce the substrata, prior to deposition, silicon substrates (10 mm x 10 mm 90 samples) were cleaned with methanol (BDH). The pure metal coatings were deposited using a 91 99.9 % Ag target, a 99.5% Ti target, a 99.5% Cu target, a 99.95% Mo target, and a 99.95% Fe 92 target all of 150 mm diameter (Teer Coatings, Worcestershire, UK). Ag, Ti, Cu and Mo 93 coatings were deposited using DC mode (Advanced Energy MDX) magnetron sputtering. An 94 average power of 500 W was applied to the Ag, Ti, Cu and Mo targets at an operating pressure 95 of 0.36 Pa with an Ar flow of 5 standard cubic cm per min (sccm). Being ferromagnetic, Fe 96 can be a difficult material to deposit using a magnetron. Thus, Fe coatings were deposited by 97 magnetron sputtering in DC mode (Advanced Energy MDX) with additional magnets placed 98 behind the substrate to link the field lines from the magnetron. An average target power of 200 99 W was used at an operating pressure between 2.27 Pa and 2.93 Pa with an Argon flow of 8 sccm. Due to the different sputtering rates of each metal, the deposition time was varied
between 3 min (Ag), 5 min (Cu), 15 min (Ti), 10 min (Mo) and 40 min (Fe).

102 Analysis measurements of coating micro-topography ( $S_a$ ) was achieved using a white light 103 profilometer (Whitehead et al., 2010) and topography ( $R_a$ ) measurements were obtained using 104 atomic force microscopy (Skovager et al., 2013). Physicochemistry of the coatings was 105 determined as carried out by Whitehead et al., (2009). Surface tension parameters for polar and 106 apolar liquids were used to calculate physicochemical parameters (van Oss et al., 1990; van 107 Oss 1995; van Oss et al. 1986) with modifications as described in Whitehead et al., (2009).

108 2.2. Microbiology

109 Three potentially pathogenic food borne microorganisms were used in this work. L. 110 monocytogenes EGDe was kindly provided by Prof. Lone Gram (DTU, Denmark). E. coli 111 CCL410 was a kind gift from Dr. Brigitte Carpentier (AFSSA, France). This strain was recovered from a heifers faecal samples by the laboratory of Dr C. Vernozy-Rozand (Unité de 112 Microbiologie alimentaire et prévisionnelle, Ecole vétérinaire de Lyon, France). This strain 113 114 was selected since it is a non-pathogenic E. coli O157:H7 wild type strain that does not the 115 carry stx1 and stx2 genes. S. aureus (NCIMB 9518) was kindly supplied by Campden BRI 116 (UK). All stock cultures were stored at -80 °C, until needed for use and were recovered as 117 described in Caballero et al., (2009).

118 Cultures were stored at 4 °C on agar for four weeks for ease of access. They were then replaced 119 by fresh cultures taken from the freezer mix. In preparation for retention assays *E. coli* was 120 inoculated onto Brain Heart Infusion Agar (BHIA) and incubated at 37 °C overnight. Ten 121 millilitres of BHIB was inoculated with a single colony of *E. coli* and incubated at 37 °C 122 overnight. One hundred microlitres of this culture was used to inoculate 100 ml BHIB, which 123 was incubated at 37 °C for 18 h with shaking (200 rpm). Following incubation, cells were 124 harvested at 567 × g for 10 min and washed once, by re-suspension in sterile distilled water, 125 vortexing for 30 sec, and then centrifugation at  $567 \times g$  for 10 min. In preparation for retention 126 assays *L. monocytogenes* was treated as the *E. coli* except cells were incubated in TSB at 30 127 °C and *S. aureus* was grown in nutrient broth at 37°C. *L. monocytogenes* were grown 30 °C 128 instead of 37 °C so that they would retain their peritrichous flagella, thus allowing the cells 129 motive activity. All cells were re-suspended to an OD of 1.0 at 540 nm in sterile distilled water 130 corresponding to  $10^8$  cfu/ml.

131 The microbial affinity to hydrocarbons (MATS) assay was followed according to an adapted 132 method described by Bellon-Fontaine et al., (1996). Retention assays were carried out 133 according to Whitehead and Verran (2007).

134 *2.3. Statistics* 

The standard deviation of the mean is shown on the graphs using error bars. *p* values werecalculated at the 95% confidence level using ANOVA and t-tests.

137 **3. Results** 

#### 138 *3.1 Surface characterisation*

139 The Ag coating demonstrated the greatest micro-topography, determined by the  $S_a$  value (21.4) 140 nm), followed by the Cu (15.1 nm), Ti (12. 9 nm), Fe (10.8 nm) and Mo (10.2 nm) (Table 2). 141 Significant differences were observed between the  $S_a$  values for the Ag and Fe, Mo or Ti coatings (p < 0.05). Differences were noted between the surface features of the individual 142 143 metals. The Ti coating had randomly spaced, sharp surface protrusions (100 nm - 300 nm wide, 144 33.2 nm  $\pm$  11.1 nm height) (Figure 1a and 2a), whereas the Ag coating demonstrated larger  $(100 \text{ nm} - 750 \text{ nm wide}, 43.4 \text{ nm} \pm 8.58 \text{ nm height})$ , closely packed, randomly distributed 145 146 rounded surface structures (Figure 1b and 2b). The Cu coating had a dense, nano textured form 147 with some linear features apparent (50 nm - 300 nm wide, 41.7 nm  $\pm$  13.4 nm height) (Figure 1c and 2c). The Fe coating had sharp, periodic protrusions (50 nm - 200 nm wide, 14.9 nm  $\pm$ 148 149 6.36 nm height) (Figure 1d and 2d). The Mo coating demonstrated randomly spaced, densly packed, linear shaped round surface protrusions (100 nm - 400 nm wide, 10.5 nm  $\pm$  4.65 nm height) (Figure 1e and 2e). The height of the coating features for all the coatings was in the nano range.

153 In terms of nano topographies (R values), generally, Ti demonstrated the highest R values 154 followed by Ag, Cu Fe and Mo (Table 2). There was a significant difference between the R155 values for the Ag, Fe and Mo surfaces, with the exception that there was no significant 156 difference observed for the  $R_{\nu}$  value between the Fe and Mo surfaces.

The Mo coating was found to be the most hydrophobic surface (-48.90 mJ/m<sup>2</sup>) (Figure 3). The Ti, Ag and Cu coatings were similar and partially hydrophilic (range -13.03 mJ/m<sup>2</sup> to – 11.72 mJ/m<sup>2</sup>), whilst the Ag coating was hydrophilic (-0.08 mJ/m<sup>2</sup>). For the apolar component, Mo demonstrated the most apolar surface (36.87 mJ/m<sup>2</sup>), with Cu demonstrating the least apolar surface (8.38 mJ/m<sup>2</sup>). For the polar, electron accepting  $\gamma_s^+$  and electron donating  $\gamma_s^-$  surface components the Ag coating demonstrated the highest values whilst the Mo coating demonstrated the lowest.

164 *3.2 Microbiology* 

*L. monocytogenes* was demonstrated to have a strongly hydrophobic cell surface, with *S. aureus* and *E. coli* being strongly hydrophilic; *E. coli* slightly more than *S. aureus*.

The different bacteria were not retained in the same percentage coverage to the same coatings. *L. monocytogenes* demonstrated the greatest percentage coverage on the Ag coating (4.27 x 10<sup>4</sup> cells/cm<sup>2</sup>) and the lowest on the Fe coating (1.74 x 10<sup>4</sup> cells/cm<sup>2</sup>) (Figure 5a). *E. coli* retained the greatest percentage coverage on the Mo coating (1.65 x 10<sup>4</sup> cells/cm<sup>2</sup>) and the least on the Fe coating (3.39 x 10<sup>2</sup> cells/cm<sup>2</sup>) (Figure 5b). *S. aureus* was retained in the greatest percentage coverage on the Fe metal coating (1.48 x 10<sup>4</sup> cells/cm<sup>2</sup> (Figure 5c) and the least the Ag coating (6.35 x 10<sup>3</sup> cells/cm<sup>2</sup>). There was a significant difference for the *L*. *monocytogenes* cells/cm<sup>2</sup> retained between the Ag and Fe coatings, for the *E. coli* between the
Mo and Fe coatings and for the *S. aureus* between the Fe and Ag coatings.

## 176 **4. Discussion**

The presence of potential food borne pathogens in the food processing industry is of increasing
concern. It is known that surface properties such as physicochemistry (Skovanger et al., 2013),
chemistry (Ma et al., 2008; Whitehead and Verran 2007) and/or topography (Palmer et al., 2007;
Whitehead et al., 2005) may affect cell retention.

181 The materials deposited were of different chemistries, and presented differently shaped 182 topographies. Surface chemistry was found to affect the numbers of bacteria retained, with the 183 greatest differences being observed on the Mo, Ag and Fe surfaces. It was found that there was 184 no trend in the shape of the surface features and bacterial retention. Although all the coating 185 topographies were well below the  $R_a$  values of 0.8 µm previously determined to be hygienic 186 for the food industry (Flint et al., 2000) it was demonstrated that there was a significant 187 difference on the effect of surface micro-topography but not nano-topography on bacterial 188 retention. Results from the *L. monocytogenes* retention assays suggest that these bacteria were 189 most influenced by surface roughness since they were retained in the greatest numbers on the 190 rough hydrophilic surfaces and least on the smooth hydrophilic surfaces. E. coli was most 191 influenced by the physicochemical status of the surface since it was retained in greatest 192 numbers on the smooth hydrophobic Mo, with the lowest  $\gamma_{AB}$ ,  $\gamma^+$  and  $\gamma^-$  values and lowest on 193 the partially hydrophilic, smooth Fe coating. S. aureus was retained in the greatest numbers on 194 the smooth, partially hydrophilic Fe coating and in the lowest numbers on the roughest, 195 hydrophilic Ag coating which suggests that surface topography had the greatest influence on 196 S. aureus retention to surfaces. It is important to note that the viability of the attached cells was 197 unknown.

There was no trend found between the hydrophobicity of the cells and retention and the physicochemistry of the surfaces. It might be that the MATS assay may be better used to determine trends between bacterial attachment and adhesion to substrata, since cell physicochemical factors are prevalent in initial cell attachment to a surface, rather than in the effects of cell retention.

The Cu and Ti coating displayed similar mid-range results for their topographies and physicochemistries, and thus did not have a significant effect on microbial retention when compared to the other coatings.

## 206 Conclusion

The results demonstrated that the different coatings exhibited a range of nano-topographies and physicochemistries. These results suggest that the effect of surface properties on cell retention is genus specific. It also highlights that different aspects and measurements used to describe the surface properties may be linked to factors affecting microbial retention hence, the use of surface roughness or physicochemistry alone is no longer adequate.

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## 216 **References**

- 217 Abban, S., Jakobsen, M., Jespersen, L. 2012. Attachment behaviour of *Escherichia coli* K12
- and *Salmonella typhimurium* P6 on food contact surfaces for food transportation. Food
  Microbiology 31, 139-147.

220

Adams, R.O. 1983. A review of stainless steel structure. Journal of Vacuum Science and
Technology A-Vacuum Surfaces and Films 1, 12-18.

223

Anonymous, 2010. BS 1134 Assessment of surface texture. Guidance and general information.

225 British Standards Institute, UK.

226

Bellon-Fontaine, M.-N., Rault, C.J., van Oss, C.J. 1996. Microbial adhesion to solvents: a novel
method to determine the electron-donor/electron-acceptor or Lewis acid–base properties of
microbial cells. . Colloids and Surfaces. B, Biointerfaces 7, 47–53.

230

231 Caballero, L., Whitehead, K.A., Allen, N.S., Verran, J. 2009. Inactivation of Escherichia coli

on immobilized TiO2 using fluorescent light. Journal of Photochemistry and Photobiology A:

233 Chemistry 202, 92-98.

234

Carpentier, B., Cerf, O. 2011. Review - Persistence of *Listeria monocytogenes* in food industry
equipment and premises. International Journal of Food Microbiology 145, 1-8.

237

Carrasco, E., Morales-Rueda, A., Maria Garcia-Gimeno, R. 2012. Cross-contamination and
recontamination by Salmonella in foods: A review. Food Research International 45, 545-556.

- Champagne, V.K., Helfritch, D.J. 2013. A demonstration of the antimicrobial effectiveness of
  various copper surfaces. Journal of Biological Engineering 7, 1-7.
- 243

Flint, S.H., Brooks, J.D., Bremer, P.J. 2000. Properties of the stainless steel substrate,
influencing the adhesion of thermo-resistant streptococci. Journal of Food Engineering 43,
235-242.

- 247
- Kelly, P., Whitehead, K.A., Li, H., Verran, J., Arnell, R.D. 2009. The influence of silver
  content on the tribological and antimicrobial properties of ZrN/Ag nanocomposite coatings.
  Journal of Nanoscience and Nanotechnology 11, 1-5.
- 251

Ma, H., Winslow, C.J., Logan, B.E. 2008. Spectral force analysis using atomic force microscopy reveals the importance of surface heterogeneity in bacterial and colloid adhesion to engineered surfaces. Colloids and Surfaces B-Biointerfaces 62, 232-237.

- 255
- Marouani-Gadri, N., Augier, G., Carpentier, B. 2009. Characterization of bacterial strains
  isolated from a beef-processing plant following cleaning and disinfection Influence of isolated
  strains on biofilm formation by Sakai and EDL 933 *E. coli* O157:H7. International Journal of
  Food Microbiology 133, 62-67.
- 260
- Palmer, J., Flint, S., Brooks, J. 2007. Bacterial cell attachment, the beginning of a biofilm.
  Journal of Industrial Microbiology and Biotechnology 34, 577-588.
- 263
- Poudelet, E. 2012. 2011 E. coli O104:H4 Outbreak in Germany and in France: Lessons learnt.
- 265 Bulletin De L Academie Veterinaire De France 165, 347-354.

Skovager, A., Whitehead, K., Siegumfeldt, H., Ingmer, H., Verran, J., Arneborg, N. 2012.
Influence of flow direction and flow rate on the initial adhesion of seven *Listeria monocytogenes* strains to fine polished stainless steel. International Journal of Food
Microbiology 157, 174-181.

271

Skovager, A., Larsen, M.H., Castro-Mejia, J.L., Hecker, M., Albrecht, D., Gerth, U., Arneborg,
N., Ingmer, H. 2013. Initial adhesion of *Listeria monocytogenes* to fine polished stainless steel
under flow conditions is determined by prior growth conditions. International Journal of Food
Microbiology 165, 35-42.

276

Tetault, N., Gbaguidi-Haore, H., Bertrand, X., Quentin, R., van der Mee-Marquet, N. 2012.
Biocidal activity of metalloacid-coated surfaces against multidrug-resistant microorganisms.
Antimicrobial resistance and infection control 1, 35-35.

280

Van Oss, C.J., Giese, R.F., Costanzo, P.M. 1990. DLVO and non-DLVO interactions in
hectorite Clays Clay Miner 38, 151.

- Van Oss, C. 1995. Hydrophobicity of biosurfaces- origin, quantitative determination and
  interaction energies. Colloids and Surfaces B: Biointerfaces 5, 91 110.
- 286
- Van Oss, C.J., Good, R.J., Chaudhury, M.K. 1986. The role of van der Waals forces and
  hydrogen bonds in 'hydrophobic interactions' between biopolymers and low energy surfaces.
- 289 Journal of Colloid and Interface Science 111, 378–390.
- 290

- Vernozy-Rozand, C., Roze, S. 2003. In: Roze, C.V.-R.a.S., (Ed.), Bilan des connaissances
  relatives aux *Escherichia coli* producteurs de Shiga-toxines (STEC). French Food Safety
  Agency, Maisons-Alfort, France.
- 294
- Whitehead, K.A., Colligon, J.S., Verran, J. 2004. The production of surfaces of defined topography and chemistry for microbial retention studies, using ion beam sputtering technology. International Biodeterioration and Biodegradation 54, 143-151.
- 298
- Whitehead, K.A., Colligon, J., Verran, J. 2005. Retention of microbial cells in substratum
  surface features of micrometer and sub-micrometer dimensions. Colloids and Surfaces BBiointerfaces 41, 129-138.
- 302
- Whitehead, K.A., Verran, J. 2007. The effect of surface properties and application method on
  the retention of *Pseudomonas aeruginosa* on uncoated and titanium-coated stainless steel.
  International Biodeterioration and Biodegradation 60, 74-80.
- 306
- Whitehead, K.A., Benson, P., Smith, L.A., Verran, J. 2009. The use of physicochemical
  methods to detect organic food soils on stainless steel surfaces. Biofouling 25, 749-756.
- 309
- Whitehead, K. A., Kelly, P. J., Li, H., Verran, J. 2010. Surface topography and
  physicochemistry of silver containing titanium nitride nanocomposite coatings Journal of
  Vacuum Science and Technology B 28 (1), 180-187.
- 313

314	Whitehead, K.A., Li, H., Kelly, P.J., Verran, J. 2011. The antimicrobial properties of titanium
315	nitride/silver nanocomposite coatings. Journal of Adhesion Science and Technology 25, 2299-
316	2315.

Wickens, D.J., West, G., Kelly, P.J., Verran, J., Lynch, S., Whitehead, K.A. 2012.
Antimicrobial activity of nanocomposite zirconium nitride/silver coatings to combat external
bone fixation pin infections. International Journal of Artificial Organs 35, 817-825.

321

Wickens, D., Lynch, S., West, G., Kelly, P., Verran, J., Whitehead, K.A. 2014. Quantifying the pattern of microbial cell dispersion, density and clustering on surfaces of differing chemistries and topographies using multifractal analysis. Journal of Microbiological Methods 104, 101-108.

326

327 Zaffe, D., Bertoldi, C., Consolo, U. 2003. Element release from titanium devices used in oral
328 and maxillofacial surgery. Biomaterials 24, 1093-1099.

329

330 Zollfrank, C., Gutbrod, K., Wechsler, P., Guggenbichler, J.P. 2012. Antimicrobial activity of

331 transition metal acid MoO<sub>3</sub> prevents microbial growth on material surfaces. Materials Science

and Engineering C-Materials for Biological Applications 32, 47-54.

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225	Table 1 Descrip	ntions of surface	measurements of	the single t	nhace metals (	Anonymous	-2010
555		prioris of surface	measurements of	uic single p	mase metals (	Anonymous,	, 2010)

Roughness parameter	Description
Roughness	The irregularities in the surface texture which are inherent in the production process but excluding waviness and errors of form
$R_a$ and $S_a$	Average absolute deviation of the roughness irregularities from the mean line over one sampling length or from the average absolute deviation of the surface respectively
$R_p$	The maximum height of the profile above the mean line within the assessment length
$R_{\nu}$	The maximum profile valley depth above the mean line within the assessment length
$R_z$	The difference in height between the average of the five highest peaks, and the five lowest valleys along the assessment length of the profile

**Table 2** Surface topography measurements using *S* and *R* values of the polycrystalline, single

340 phase metal coatings demonstraing that Ag was the roughest surface with the greatest

	Ti	Ag	Cu	Fe	Mo
$S_a$ (nm)	12.9 ± 2.70	21.4 ± 1.75	15.1 ± 4.15	10.8 ± 2.73	10.2 ± 1.82
$R_a$ (nm)	7.51 ± 0.59	6.96 ± 0.53	4.39 ± 0.75	1.46 ± 0.27	1.45 ± 0.17
$R_p(nm)$	50.4 ± 16.2	48.1 ± 12.6	49.4 ± 19.6	14.6 ± 5.73	9.86 ± 2.00
$R_v(\mathrm{nm})$	15.90± 6.05	38.6 ± 4.55	33.9 ± 7.14	$\begin{array}{c} 15.2 \pm \\ 6.98 \end{array}$	11.2 ± 1.29
$R_z(nm)$	$7.51 \pm$	9.09 ±	$6.09 \pm$	$1.85 \pm$	$1.87 \pm$

341 topographical values, whilst Fe and Mo were the smoothest

342



Fig. 1. AFM two-dimensional profiles of surface topographies of the single phase metals a)
Ti b) Ag c) Cu d) Fe and e) Mo demonstrating the differences in the shape and scale of the
surface features across one X axis. Z heights are not on the same scale in order to allow
visualisation of surface features



**Fig. 2.** AFM images of surface nano-topographies of the single phase metals a) Ti b) Ag c)

Cu d) Fe and e) Mo demonstrating the surface features. Z heights are not on the same scale in

353 order to allow visualisation of surface features



**Fig. 3.** Physicochemical measurements of single phase metals surfaces demonstrating that the



358 molybdenum the most hydrophobic





361 **Fig. 4.** Cell affinity to solvents following the MATS assay demonstrating that each bacterial

362 species demonstrates unique cell surface properties. As demonstrated by % cell affinity to

363 hexadecane, *L. monocytogenes* was the most hydrophobic, whilst *E. coli* was the least



Fig. 5. Number cells retained on the surface following retention assays a) Lm = L. 

monocytogenes b) Ec = E. coli and c) Sa = S. aureus 

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