

Small-scale spatial variability of selected soil biological properties

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Abstract

A strategy for sampling soil from intact monolith lysimeters was established based on measurements of spatial heterogeneity within the lysimeter area. This was part of an ongoing study to determine relationships between soil microbial diversity and nutrient loss by leaching. The sampling protocol had to allow for collection of soil on a regular basis (as opposed to destructive sampling) and ensure high spatial independence of subsamples. On each of two sites (one developed under organic crop management, the other under conventional crop management), ten 15-cm soil cores (sampling points) were taken from three areas (replicates) of 50-cm-diameter (lysimeter surface area) and separately analysed for biotic (microbial biomass carbon and nitrogen; arginine deaminase activity) and abiotic (total carbon and nitrogen) soil properties. The data was tested for variability, expressed as coefficient of variance (biotic and abiotic), and spatial heterogeneity using geostatistics (biotic properties). The biotic soil properties showed significant differences among sampling points, whereas the abiotic parameters were useful in differentiating on a larger scale, i.e. between sites. For all soil properties tested, the differences among the replicates were smaller than those between sites or among points indicating that, in the main experiment, all treatments can be sampled following the same pattern. Geostatistical analysis and fitting of an exponential model showed that a spatial structure exists in the biotic soil properties and that the samples are independent beyond separation distances of 25-30 cm. A revised sampling pattern consisting of 11 samples per lysimeter is described.

Keywords

Small-scale spatial variability; Intact monolith lysimeters; Sampling pattern; Microbial biomass carbon; Microbial biomass nitrogen; Arginine deaminase activity

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Introduction

It is widely accepted that soil biota play a vital role in the maintenance of soil fertility and productivity. Although we are able to appreciate the significance of microorganisms in the soil we have little information on the role microbial community structure and function play in determining sustainability and functioning of soil ecosystems (Ritz et al., 1994; Insam and Rangger, 1997). Microbial diversity in soil is influenced by a number of factors, including soil properties, environmental conditions and anthropogenic activities. It is, thus, likely that changing management practices can have significant and lasting effects on the functioning of the soil microbial community and the ecosystem as a whole (Waldrop et al., 2000; O'Donnell et al., 2001). Improved understanding of soil microbial diversity and function may enhance our ability to manipulate nutrient cycling and retention *in situ* in order to maintain and improve soil quality and sustainability (Kennedy and Gewin, 1997; Bending et al., 2000).

One way of investigating relationships between microbial diversity (including community composition, activity and function), and how it relates to nutrient cycling and its associated environmental impacts is by using intact monolith lysimeters (e.g. Cameron et al., 1992; Di et al., 1998; Turner and Haygarth, 2000; Riley et al., 2002). Small to medium sized monolith lysimeters (surface area of 0.2 to 0.5 m²) can provide all the benefits of a pot trial, including better control over irrigation, pests and weeds than can be achieved in the field, together with the potential to measure leaching and gaseous losses.

The present study is part of an ongoing experiment that uses intact monolith lysimeters (50 cm diameter; 70 cm deep) of the same soil type taken from adjacent sites under long-term organic and conventional management and involves measurement of nitrogen (N) loss by leaching together with determination of a range of biological and chemical soil properties.

Protocols have been established to take soil from different depths in the lysimeters by destructive sampling (Fraser et al., 1994; Cookson et al., 2001). However, it is not common practice to repeatedly take *in situ* soil samples from undisturbed monolith lysimeters since the type and size of lysimeters used in this study are more commonly maintained under grassland where sampling would create preferential flow pathways (e.g. Bidwell, 2000; Stout et al., 2000; Turner and Haygarth, 2000; Di and Cameron, 2002). A thorough review of the literature revealed no published studies on sampling strategies for intact monolith lysimeters of the size used in this study (c. 0.2 m²). The relatively small surface area and volume of the lysimeters can be seen as the main factors limiting the number of samples that can be taken from the lysimeter at any one time without ultimately compromising the experiment.

The objective of the study presented in this paper was to develop and evaluate a strategy for regular sampling of soil from lysimeters by measuring variance and spatial variability for a range of soil biological parameters and analysing them by means of geostatistics. The results also have implications for field sampling in terms of appropriate distances between sampling points.

Materials and Methods

Context

Two sites, approximately 2 km apart, were identified within the cropping farm at Lincoln University (Canterbury, New Zealand) that had been farmed under contrasting 'organic' and 'conventional' management systems for around 25 years. The organic site (ORG) was established in 1976, while the conventional site (CON) had been maintained under intensive mixed cropping for over 100 years. The soil at both sites was a Wakanui silt loam (free draining to 75 cm) (Mottled Immature Pallic Soil, NZ classification; Udic Ustochrept, USDA).

In November 2001, eight replicate intact monolith lysimeters (50 cm diameter by 70 cm deep) in steel casings were taken from each site, and installed in the lysimeter laboratory according to the protocol established (Cameron et al., 1992; Di and Cameron, 2002). The lysimeters will be maintained under organic and conventional cropping regimes for a minimum of 5 years and soil samples will be taken on a regular basis (bulked) to investigate relationships between soil microbial activity and nutrient loss.

Spatial variability study

The spatial variability study was carried out at ORG and CON on areas of soil equivalent to the lysimeters described above. In April 2002, 10 soil samples were taken within each of three 50-cm-diameter areas (surface area of a lysimeter) in a distinctive pattern (Fig. 1) and a range of soil properties was tested to determine spatial variation within each area. The three sampling areas were placed at distances of approximately 70 to 90 cm from each other (distance from the centre of one area to the centre of the other). This corresponded to the distance between the monoliths when the lysimeters were collected. The soil properties were measured in duplicate in each of 60 soil cores (2.5 cm diameter; 15 cm depth) – two sites, three areas per site (replicates), ten sample points per replicate (positions). The design of the sampling pattern was based on two main constraints: the limited area (c. 0.2 m²) and volume of the lysimeters, and the requirement of a relatively large number of samples and distance comparisons to guarantee meaningful geostatistical analysis. The selected pattern ensured a variety of distances between sampling points resulting in 10 to 40 cm-distance comparisons.

Soil analyses

The samples were passed through a 4-mm mesh and plant material and roots were removed. Sieved samples were stored at 4°C in the dark for up to three weeks until analyses took place. Microbial biomass carbon (C_{mic}) and nitrogen (N_{mic}) were estimated by fumigation extraction following the method of Sparling and West (1988). For conversion of C and N to C_{mic} and N_{mic}, factors of $k_{eC} = 0.35$ (Sparling et al., 1990) and $k_{eN} = 0.54$ (Brookes et al., 1985) were used, respectively. Arginine deaminase activity (or arginine ammonification) (ADA) was measured using a modification of the method of Alef and Kleiner (1987) as described in Alef and Nannipieri (1995). Following extraction with 2 M KCl, ammonium-N (NH₄-N) content in the samples was determined by automated flow injection analysis (Tecator, Sweden). Total carbon (C_{tot}) and nitrogen (N_{tot}) were measured in air-dried and sieved samples (2-mm mesh) on a Leco® CNS-2000 elemental analyser.

Statistical analysis

Variation in biotic and abiotic soil parameters was determined in soils from ORG and CON by calculating coefficients of variance between positions (sampling points within each area), replicates (sampling areas) and the two sites. These different levels of variance were determined for the soil properties C_{mic}, N_{mic}, ADA, C_{tot}, and N_{tot} and some ratios of these (microbial carbon: nitrogen ratio [C_{mic}/N_{mic}]; total carbon: total nitrogen ratio [C/N]; microbial carbon: total carbon ratio [C_{mic}/C_{tot}]).

Geostatistical methods were applied to study spatial variability of the soil properties (Goovaerts, 1998; Legendre and Legendre, 1998) (Eq. 1):

$$\gamma(h) = \frac{1}{2N(h)} \sum_{i=1}^n [z(s_i) - z(s_i + h)]^2 \quad (1)$$

where $\gamma(h)$ = semivariance at a given distance h ;
 $N(h)$ = number of observation pairs for a given distance h ;
 $z(s_i)$ = value of respective soil property at sampling point s_i ;
 $z(s_i+h)$ = value of soil property at a sampling point with the distance h from s_i .

Based on frequency distributions (cf. Section 3.2), C_{mic} , N_{mic} , and ADA data were used to measure autocorrelation of positions and estimate semivariograms. For each soil property, semivariance was calculated for ORG, CON and for the average of the two.

Five distance classes were established, 10 cm (consisting of comparisons where $h = 10$ cm; $N(h)=18$), 15 cm ($h=14.1-15.3$ cm; $N(h)=42$), 20 cm ($h=20$ cm; $N(h)=33$), 30 cm ($h=28-30$ cm; $N(h)=33$) and 40 cm ($h=37-40$ cm; $N(h)=9$). Due to the limitations in sampling area and removable soil volume, a sampling plan consisting of 10 samples was chosen; as a result the rule of thumb (at least 30 observation pairs per distance class (Stein and Ettema, 2003)) could not be fulfilled for all distance classes.

Semivariograms for the three soil properties were fitted with an exponential model (based on Eq. 2), which allowed the spatial dependence for the respective soil properties to be determined (Legendre and Legendre, 1998).

$$y = C_0 + C_1 \left(1 - \exp^{-3x/a}\right) \quad (2)$$

where y = semivariance;
 x = distance between sampling points;
 C_0 = nugget or intercept, i.e. the level of variability at distance $h = 0$ cm;
 C_1 = component related to the spatial pattern of the model;
 a = range, i.e. distance between sampling points at which asymptote is reached.

If spatial dependence exists, it is possible to fit an exponential model and the data will show low levels of variance for short separation distances (in this case 10 cm), followed by an increase until the asymptote is reached at a distance a (range). Beyond the range, the semivariance measured for the data points is independent from the respective separation distance. The sill ($C = C_0 + C_1$ [Eq. 1]) indicates the variance of the asymptote and signifies the total variance of the model.

The decreasing number of pairs of observations with increasing distance between sampling points can result in too few comparisons to achieve sound results. In this case, it was possible to fit the model effectively to data derived from sampling points with separation distances from 0 to 40 cm for ORG and average, while for CON only points from the 30 cm- distance class were included.

The data were analysed using general linear model analysis of variance in Minitab® for Windows Release 13.1 (©2000, Minitab Inc., USA). Regressions were performed in Sigmaplot for Windows Version 5.00 (© 1986-1999, SPSS, Inc.) and R^2 was used to measure goodness of fit of data to the exponential model.

Results

Variance between sites, replicates, and positions

For C_{mic} , N_{mic} and ADA, the variability among the 10 positions within each area, expressed as coefficients of variance of the mean value (CV), was highest ($CV_{position}$: 78.6% [C_{mic}]; 56.1% [N_{mic}]; 50.4% [ADA]) (Fig. 2). For C_{mic} and N_{mic} , this was followed by the variability between the areas

within each site (CV_{area}). ADA showed a similar variability between sites (CV_{site}) as among the 10 positions, while CV_{area} was smaller than the other two ($<5\%$). For $C_{\text{mic}}/N_{\text{mic}}$, CV_{site} was highest (46.6%), while CV_{area} and CV_{position} were both around 27%. For C_{mic} and N_{mic} , CV_{site} was regarded as zero based on the observation that in both cases the mean squares (MSq) for the sites were smaller than for the replicates.

The patterns of variability were similar for C_{tot} and N_{tot} and, consequently $C_{\text{mic}}/C_{\text{tot}}$ and C/N . CV_{position} was larger than CV_{area} (tendency towards zero for C_{tot} and N_{tot}), while the highest degree of variance could be observed for *site* (Fig. 2). For C_{tot} , the error mean square was bigger than the mean square for the effect “area (site)” (replicates within each site). Consequently, the calculated CV was negative and was regarded as zero.

The average values of the biotic soil properties (C_{mic} , N_{mic} , ADA) show that CV_{site} was smaller than CV_{area} and CV_{position} (23.4% as compared to 23.8% and 52.8%, respectively) whereas the average CVs of the abiotic soil properties (C_{tot} , N_{tot}) show the differences between sites were larger (81.5%) than between positions (13.7%) (Fig. 2). For the average of chemical soil properties, CV_{area} was smallest with 4.9%.

This pattern is supported by the results presented in Table 1. The biotic soil properties measured in this experiment (C_{mic} , N_{mic} , ADA and $C_{\text{mic}}/N_{\text{mic}}$) were able to differentiate between the positions in each replicate on a significant level. All but one property (C_{mic}) were able to detect significant differences between the sites, and this effect was highly significant for $C_{\text{mic}}/N_{\text{mic}}$ and $C_{\text{mic}}/C_{\text{tot}}$, as well as for the abiotic soil properties C_{tot} , N_{tot} and C/N ($P<0.001$). This indicated that measuring abiotic rather than biotic soil properties best separated the two sites.

Comparing the mean square values for site and area (site) supports these findings. For the soil properties that showed significant differences between sites, the mean square value for site (MSq_{site}) was bigger than the mean square value for areas within sites ($MSq_{\text{area (site)}}$) (Table 1). On the other hand, $MSq_{\text{area (site)}}$ is expected to be greater than MSq_{site} when differences between replicates are more pronounced than between sites. In this case, variability between sites was regarded as zero (C_{mic} and N_{mic}), as the main purpose of this study was to evaluate the spatial variability within each area (i.e. between positions).

As mentioned above, *site* had a significant effect on all but one property (C_{mic}). The mean values indicated that a significantly higher ADA activity was measured on ORG. Similarly, there were higher contents of N_{mic} , a higher microbial quotient ($C_{\text{mic}}/C_{\text{tot}}$) and higher C/N ratio ($P<0.001$ in both cases). CON showed higher levels of C_{tot} , N_{tot} ($P<0.001$ in both cases) and C_{mic} (not significant) (Table 2).

Looking at the levels of significance of the three effects on average biotic and abiotic soil parameters, respectively, biotic soil characteristics detected highly significant differences among the positions within each area and among areas within each site (Table 3). The abiotic soil properties on the other hand revealed significant differences between sites, while they did not distinguish between the replicates within each site. These findings are supported by the results displayed in Table 1 and Fig. 2.

Spatial pattern determination

Frequency distribution

Across the 60 sampling points at six sampling locations, ADA varied intensely (data range = $1.6 \mu\text{g NH}_4\text{-N g}^{-1} \text{ h}^{-1}$; $CV = 22.5\%$) and displayed a strong positive skew. Tests for normality suggest that the data was randomly distributed; hence, the changes in value seemed to be independent from the respective sampling locations (Table 4).

For C_{mic} , values covered a range of $420 \mu\text{g C g}^{-1}$ soil and showed a coefficient of variance of 0.77%. The frequency distributions for C_{mic} showed a slight positive skew. N_{mic} concentrations ranged over $70 \mu\text{g N g}^{-1}$ with a standard variation of $16.4 \mu\text{g g}^{-1}$. The coefficient of skewness for the frequency distribution was 0.32. Tests for normality showed that in both cases the frequency distributions can be regarded as being normally distributed (Table 4). This indicates that a dependence exists between change in value and increasing sampling point distance.

For C_{tot} and N_{tot} , the frequency distributions were totally random (results not shown) indicating no correlation exists between sampling location and change in value for these parameters within the scale sampled; hence, only C_{mic} , N_{mic} and ADA were tested for semivariance.

Spatial variability

Looking at the development of semivariance in C_{mic} and N_{mic} for ORG, CON and the average of the two with increasing distance between sampling points, it can be noticed that CON showed much lower levels of semivariance than ORG in both, C_{mic} and N_{mic} (Fig. 3). The exponential model, however, suggested a bigger nugget (intercept) for CON than ORG and average. In all cases, the model represented the observed amounts of variance sufficiently well, although CON showed low values for spatial dependence and R^2 (Table 5).

In ADA, the exponential models did not match the data as well as they did for the other two soil properties. For ORG, high value for the sill could be observed compared to the initial value. As shown in Fig. 3, the curve does not reach its asymptote within the displayed range. For ORG, CON and average the estimated range was higher in ADA than in C_{mic} and N_{mic} . And while values for spatial dependence were within the same range as for the other two properties, $R^2 < 0.9$ and $P > 0.05$ indicate that the regression was not significant for ORG, CON and average (Table 5).

Discussion

It is well known that soils vary spatially even over short distances. Spatial variability is understood to have an effect on transport processes in soils, e.g. of organic pollutants (Søvik and Aagaard, 2003), biomass turnover rate (Harden and Joergensen, 2000) and on nutrient cycling processes (de Boer et al., 1996; Corre et al., 2002). Consequently, assessing the spatial structure of a site will help developing better nutrient management strategies, and can serve as a means designing suitable sampling patterns (Ettema and Wardle, 2002; Franklin et al., 2002; Stein and Ettema, 2003). Soil spatial structure has been investigated at different scales (cm, m, km) and with different objectives. Most studies on spatial variability have concentrated on aboveground biota and abiotic soil properties, e.g. soil moisture, hydraulic conductivity, temperature (Bruckner et al., 1999; Al-Kayssi, 2002; Søvik and Aagaard, 2003). Nonetheless, an increasing number of studies have begun to examine heterogeneity of biological soil properties in general and the microbial community in particular (Morris, 1999; Harden and Joergensen, 2000; Corre et al., 2002; Franklin et al., 2002; Nunan et al., 2002; Ettema and Yeates, 2003). Depending on the scale, spatial structure in microbial soil properties is influenced by land use, soil carbon content, topography, plant cover (including form, size, spacing), soil aggregates, fine roots, and substrate hotspots (Ettema and Wardle, 2002). This implies the importance of determining spatial variability even within small areas in order to obtain meaningful bulk samples. However, literature revealed no evidence of the application of spatial variability data to design a soil sampling strategy for within lysimeters.

The overall goal of this study was to design a protocol that allows soil samples to be taken from intact monolith lysimeters on a regular basis. Therefore, the degree of variance was determined on different levels (between sites, among replicates, among positions) and the pattern of spatial heterogeneity within the area of a lysimeter was established.

In general, results suggest that the biotic soil properties studied exhibited spatial dependence within the area of a lysimeter. The existence of spatial patterns in biological soil parameters at this scale is consistent with results shown in previous studies (Brockmann and Murray, 1997; Bruckner et al., 1999; Morris, 1999). ORG and CON showed similar patterns of semivariance. However, spatial dependence was much smaller for CON than for ORG (44-64% as opposed to 82-95%) (Table 5). This outcome is consistent with the expectation that cultivated soils are spatially less variable since cultivation disrupts the spatial pattern (Goovaerts, 1998; Röver and Kaiser, 1999). CON had been cultivated a short time before sampling took place, whereas ORG had been under herb ley for approximately 18 months. This initial difference should not interfere with sampling during the main experiment given that all lysimeters will be cropped and cultivated in the same way. Therefore, an adjustment of the spatial variability in the soils originating from CON is to be expected over time.

Patterns for semivariance were similar in C_{mic} and N_{mic} , both reaching the sill within the sampled scale, whereas in ADA, semivariance displayed a different trend. This suggests that for C_{mic} and N_{mic} , samples taken within the area of the lysimeters (50-cm diameter) are independent from each other when their separation distance exceeds 25 to 30 cm, while, according to the exponential model, for ADA, the samples will not be stochastically independent from each other within the area of the lysimeters. The random distribution of the data points indicates a nugget effect, i.e. the variance is not spatially structured at the scale measured (10-40 cm). However, spatial dependence could exist below this scale.

Statistical analysis can be problematic for autocorrelated samples since autocorrelation reduces variance within the measured bulk sample and increases variability among samples (Franklin et al., 2002). It is, therefore, important to collect a representative bulk sample that consists of a large number of independent samples in order to make valid statistical analysis possible. Regarding the lysimeter sampling design, this stresses the importance of taking a considerable number of independent subsamples (with separation distances >20 cm). In this study, 10 soil cores were sampled per lysimeter (Fig. 1) resulting in 25 pairs with separation distances of 20 cm or more, which was sufficient to achieve meaningful results. It will be difficult to largely increase the number of samples that are more than 30 cm apart given the limited area (c. 0.2 m^2). However, in order to increase the number of pairs in the distance classes 20, 30 and 40 cm, we suggest a sampling regime that consists of 11 soil cores and, consequently, 39 observation pairs per bulk sample (Fig. 4).

It is also important that the samples taken from the different lysimeters are independent to ensure valid statistical analysis. This can be considered to be the case based on the results displayed in Table 5 and Fig. 3, and assuming that the lysimeters have separation distances of more than 50 cm.

The relatively higher values of $CV_{position}$ for C_{mic} and N_{mic} , ADA, and consequently for the average for biotic soil properties, imply that these soil properties can be used to recognise differences on a small spatial scale, i.e. among sampling points up to 40 cm apart, while abiotic soil properties (C_{tot} and N_{tot}) are useful for differentiating on a larger scale, in this case between sites 2 km apart (Fig. 2). The methods showed good reproducibility, there was little variance between the replicates (data not shown) and for all soil parameters the measurable differences among replicates expressed as coefficient of variance were smaller than the differences among positions or sites. Thus, one general sampling strategy can be applied for all treatments and replicates. The results suggest that this is particularly true when assessing biotic soil properties as only minor differences between replicates and sites were detected. The potential of the examined abiotic properties to differentiate between sites will be of advantage after the four different treatments have been imposed on the lysimeters and treatment related effects have to be determined.

In conclusion, the findings of this study showed that

- (a) a spatial structure exists in the biotic soil properties at the scale examined with samples being independent beyond separation distances of 25-30 cm;

- (b) measuring biological soil properties (C_{mic} , N_{mic} , ADA, C_{mic}/N_{mic}) detect differences at a small-scale spatial level (<0.5 m), whereas abiotic soil properties (C_{tot} , N_{tot} , C/N) as well as C_{mic}/C_{tot} can be used to distinguish on a larger scale (between sites, separated by several kilometres);
- (c) the differences between replicates can be disregarded when designing a sampling regime for intact monolith lysimeters. Consequently, all lysimeters can be sampled following the same protocol. However, the sampling design should be adjusted and the number of subsamples increased from 10 to 11 to ensure more distance comparisons of 20 cm and over.

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References

- Alef, K., Kleiner, D., 1987. Applicability of arginine ammonification as indicator of microbial activity in different soils. *Biology & Fertility of Soils* 5, 148-151.
- Alef, K., Nannipieri, P., 1995. *Methods in applied soil microbiology and biochemistry*. Academic Press Limited, London. 576 pp.
- Al-Kayssi, A.W., 2002. Spatial variability of soil temperature under greenhouse conditions. *Renewable Energy* 27, 453-462.
- Bending, G.D., Putland, C., Rayns, F., 2000. Changes in microbial community metabolism and labile organic matter fractions as early indicators of the impact of management on soil biological quality. *Biology & Fertility of Soils* 31, 78-84.
- Bidwell, V.J., 2000. Decision support for protecting groundwater from land treatment of wastewater. *Environmetrics* 11, 553-562.
- Brockmann, F.J., Murray, C.J., 1997. Subsurface microbiological heterogeneity: current knowledge, descriptive approaches and applications. *FEMS Microbiology Reviews* 20, 231-247.
- Brookes, P.C., Landman, A., Pruden, G., Jenkinson, D.S., 1985. Chloroform fumigation and the release of soil nitrogen: a rapid extraction method to measure microbial biomass nitrogen in soil. *Soil Biology & Biochemistry* 17, 837-842.
- Bruckner, A., Kandeler, E., Kampichler, C., 1999. Plot-scale spatial patterns of soil water content, pH, substrate-induced respiration and N mineralization in a temperate coniferous forest. *Geoderma* 93, 207-223.
- Cameron, K.C., Smith, N.P., McLay, C.D.A., Fraser, P.M., McPherson, P.J., Harrison, D.F., Harbottle, P., 1992. Lysimeters without edge-flow: an improved design and sampling procedure. *Soil Science Society of America Journal* 56, 1625-1628.
- Cookson, W.R., Rowarth, J.S., Cameron, K.C., 2001. The fate of autumn-, late winter- and spring-applied nitrogen fertilizer in a perennial ryegrass (*Lolium perenne* L.) seed crop on a silt loam soil in Canterbury, New Zealand. *Agriculture, Ecosystems & Environment* 84, 67-77.

- Corre, M.D., Schnabel, R.R., Stout, W.L., 2002. Spatial and seasonal variation of gross nitrogen transformations and microbial biomass in a Northeastern US grassland. *Soil Biology & Biochemistry* 34, 445-457.
- de Boer, W., Klein Gunnewiek, P.J.A., Parkinson, D., 1996. Variability of N mineralization and nitrification in a simple, simulated microbial forest soil community. *Soil Biology & Biochemistry* 28, 203-211.
- Di, H.J., Cameron, K.C., Moore, S., Smith, N.P., 1998. Nitrate leaching and pasture yields following the application of dairy shed effluent or ammonium fertilizer under spray or flood irrigation: results of a lysimeter study. *Soil Use & Management* 14, 209-214.
- Di, H.J., Cameron, K.C., 2002. The use of nitrification inhibitor, dicyandiamide (DCD), to decrease nitrate leaching and nitrous oxide emissions in a simulated grazed and irrigated grassland. *Soil Use & Management* 18, 395-403.
- Ettema, C.H., Wardle, D.A., 2002. Spatial soil ecology. *Trends in Ecology & Evolution* 17, 177-183.
- Ettema, C.H., Yeates, G.W., 2003. Nested spatial biodiversity patterns of nematode genera in a New Zealand forest and pasture soil. *Soil Biology & Biochemistry* 35, 339-342.
- Franklin, R.B., Blum, L.K., McComb, A.C., Mills, A.L., 2002. A geostatistical analysis of small-scale spatial variability in bacterial abundance and community structure in salt marsh creek bank sediments. *FEMS Microbiology Ecology* 42, 71-80.
- Fraser, P.M., Cameron, K.C., Sherlock, R.R., 1994. Lysimeter study of the fate of nitrogen in animal urine returns to irrigated pasture. *European Journal of Soil Science* 45, 439-447.
- Goovaerts, P., 1998. Geostatistical tools for characterizing the spatial variability of microbiological and physico-chemical soil properties. *Biology & Fertility of Soils* 27, 315-334.
- Harden, T., Joergensen, R.G., 2000. Relationships between simulated spatial variability and some estimates of microbial biomass turnover. *Soil Biology & Biochemistry* 32, 139-142.
- Insam, H., Rangger, A., 1997. *Microbial communities: functional vs. structural approaches*. Springer Verlag, Berlin. 263 pp.
- Kennedy, A.C., Gewin, V.L., 1997. Soil microbial diversity: Present and future considerations. *Soil Science* 162, 607-617.
- Legendre, P., Legendre, L., 1998. *Numerical Ecology*. Elsevier, Amsterdam, The Netherlands. 870 pp.
- Morris, S.J., 1999. Spatial distribution of fungal and bacterial biomass in southern Ohio hardwood forest soils: fine scale variability and microscale patterns. *Soil Biology & Biochemistry* 31, 1375-1386.
- Nunan, N., Wu, K., Young, I.M., Crawford, J.W., Ritz, K., 2002. *In situ* spatial patterns of soil bacterial populations, mapped at multiple scales, in arable soil. *Microbial Ecology* 44, 296-305.
- O'Donnell, A.G., Seasman, M., Macrae, A., Waite, I., Davies, J.T., 2001. Plants and fertilisers as drivers of change in microbial community structure and function in soils. *Plant & Soil* 232, 135-145.
- Riley, N.G., Zhao, F.J., McGrath, S.P., 2002. Leaching losses of sulphur from different forms of sulphur fertilizers: a field lysimeter study. *Soil Use & Management* 18, 120-126.
- Ritz, K., Dighton, J., Giller, K.E., 1994. *Beyond the Biomass: compositional and functional analysis of soil microbial communities*. John Wiley & Sons, Inc., Chichester. 275 pp.

- Röver, M., Kaiser, E.-A., 1999. Spatial heterogeneity within the plough layer: low and moderate variability of soil properties. *Soil Biology & Biochemistry* 31, 175-187.
- Søvik, A.K., Aagaard, P., 2003. Spatial variability of a solid porous framework with regard to chemical and physical properties. *Geoderma* 113, 47-76.
- Sparling, G.P., West, A.W., 1988. Modifications to the fumigation extraction technique to permit simultaneous extraction and estimation of soil microbial C and N. *Communications in Soil Science & Plant Analysis* 19, 327-344.
- Sparling, G.P., Feltham, C.W., Reynolds, J., West, A.W., Singleton, P., 1990. Estimation of soil microbial C by a fumigation-extraction method: use on soils of high organic matter content, and a reassessment of the k_{ec} -factor. *Soil Biology & Biochemistry* 22, 301-307.
- Stein, A., Ettema, C.H., 2003. An overview of spatial sampling procedures and experimental design of spatial studies for ecosystem comparisons. *Agriculture, Ecosystems & Environment* 94, 31-47.
- Stout, W.L., Fales, S.L., Muller, L.D., Schnabel, R.R., Weaver, S.R., 2000. Water quality implications of nitrate leaching from intensively grazed pasture swards in the northeast US. *Agriculture, Ecosystems & Environment* 77, 203-210.
- Turner, B.L., Haygarth, P.M., 2000. Phosphorus forms and concentration in leachate under four grassland soil types. *Soil Science of America Journal* 64, 1090-1099.
- Waldrop, M.P., Balsler, T.C., Firestone, M.K., 2000. Linking microbial community composition to function in a tropical soil. *Soil Biology & Biochemistry* 32, 1837-1846.

Table 1

Results of ANOVA based on levels of eight selected parameters and their ratios. Displayed are levels of significance for the effects of position (area), area (site) and site, as well as comparisons of mean square values (MSq).

Soil property	Position (area)	Area (site)	Site	MSq
C_{mic} ($\mu\text{g C g}^{-1}$ soil)	***	**	NS	area (site)>site
N_{mic} ($\mu\text{g C g}^{-1}$ soil)	***	***	*	area (site)>site
ADA ($\mu\text{g NH}_4\text{-N g}^{-1} \text{ h}^{-1}$)	***	NS	**	site>area (site)
C_{mic}/N_{mic}	***	***	***	site>area (site)
C_{mic}/C_{tot} (%)	ND	***	***	site>area (site)
C_{tot} (%)	ND	NS	***	site>area (site)
N_{tot} (%)	ND	NS	***	site>area (site)
C/N	ND	***	***	site>area (site)

Position (area): positions within each area; Area (site): areas within each site; ***, $P<0.001$; **, $P<0.01$; *, $P<0.05$; NS, not significant; ND, not determined; N=60 for area (site) and site; N=120 for position (area) (For C_{tot} , N_{tot} , C/N and C_{mic}/C_{tot} N=60 for position (area) which makes statistical analysis impossible.).

Table 2

Levels of selected soil properties in ORG and CON and significance of the effect *site*.

Soil property	ORG	CON	Site effect
C_{mic} ($\mu\text{g C g}^{-1}$ soil)	460.8 (16.1)	502.6 (16.1)	NS
N_{mic} ($\mu\text{g C g}^{-1}$ soil)	62.7 (2.36)	54.6 (2.36)	*
ADA ($\mu\text{g NH}_4\text{-N g}^{-1} \text{ h}^{-1}$)	0.96 (0.047)	0.74 (0.047)	**
C_{mic}/N_{mic}	7.77 (0.30)	9.61 (0.30)	***
C_{mic}/C_{tot} (%)	2.02 (0.043)	1.77 (0.043)	***
C_{tot} (%)	2.27 (0.042)	2.84 (0.042)	***
N_{tot} (%)	0.19 (0.004)	0.24 (0.004)	***
C/N	12.3 (0.048)	11.5 (0.048)	***

Values are mean (standard error of mean in brackets); ***, $P<0.001$; **, $P<0.01$; *, $P<0.05$; NS, not significant; N=60.

Table 3

Significance of the effects of position (area), area (site), and site on average log-transformed values of biotic (C_{mic} , N_{mic} , ADA, C_{mic}/N_{mic}) and abiotic (C_{tot} , N_{tot} , C/N) soil properties.

	biotic	abiotic
position (area)	***	ND
area (site)	**	NS
site	NS	***

position (area): positions within each area; area (site): areas within each site.

NS, not significant; ND, not determined.

Table 4

Descriptive statistics for frequency distribution of C_{mic} , N_{mic} , ADA. (A^2 , P: Anderson-Darling Normality Test).

	C_{mic} ($\mu\text{g C g}^{-1}$ soil)	N_{mic} ($\mu\text{g N g}^{-1}$ soil)	ADA ($\mu\text{g NH}_4\text{-N g}^{-1} \text{ h}^{-1}$)
A^2	0.320	0.394	1.14
P	0.524	0.365	0.005
Mean	481.7	58.6	0.851
StDev	97.4	16.4	0.284
Skewness	0.218	0.323	1.49
Median	480.6	59.0	0.799
SE	12.6	2.1	0.037
CV %	0.736	2.52	2.5

Table 5

Parameters of autocorrelation in C_{mic} , N_{mic} and ADA based on best-fit lines derived from exponential model (Eq. 1)

		Nugget (C_0)	Sill ($C=C_0+C_1$)	Spatial dependence	R^2	P	range (cm)
C_{mic}	ORG	824	16292	0.95	0.99	0.001	43
	CON	1110	3075	0.64	0.98	0.018	31
	average	958	8788	0.89	0.98	0.003	13
N_{mic}	ORG	21	333	0.94	0.99	0.002	31
	CON	27	61	0.56	0.96	0.04	25
	average	23	184	0.88	0.99	<0.001	22
ADA	ORG	0.023	0.126	0.82	0.81	0.082	179
	CON	0.016	0.028	0.44	0.67	0.327	48
	average	0.019	0.047	0.60	0.81	0.081	36

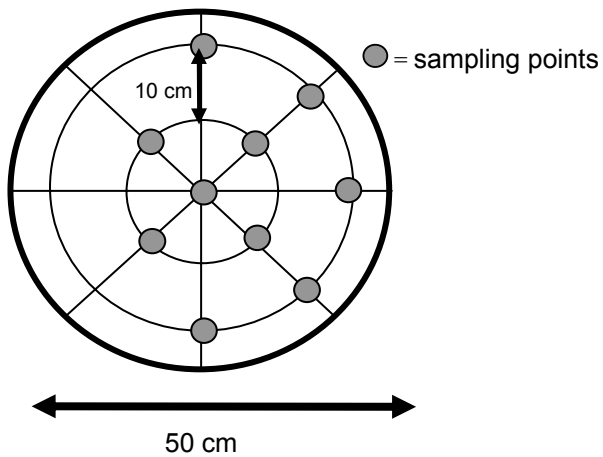


Fig. 1.
Sampling regime for spatial pattern determination.

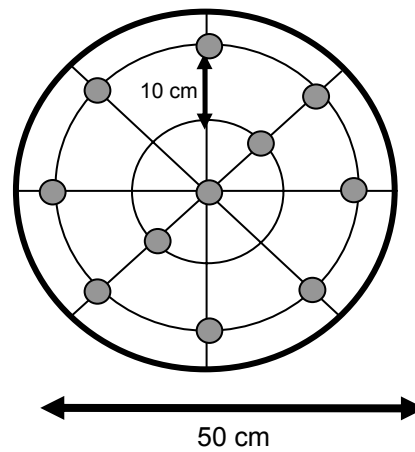


Fig. 4.
Revised strategy for regular soil sampling from lysimeters.

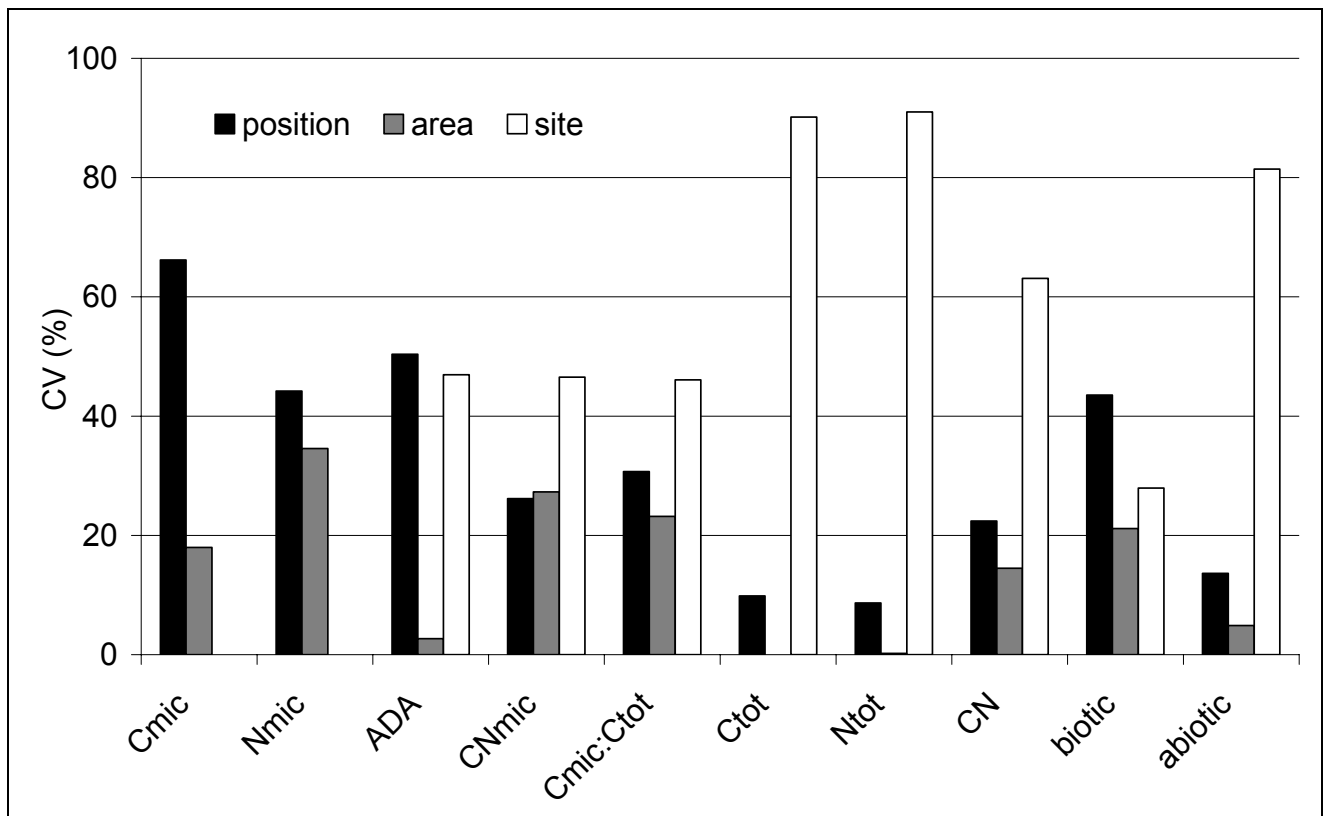


Fig. 2.

Coefficients of variance at different levels of variability for eight soil properties and their averages (position = variance between ten positions within each area; area = variance among three replicates within each site; site = variance between the sites ORG and CON): Cmic = microbial C; Nmic = microbial N; ADA = arginine deaminase activity; Cmic/Nmic = microbial carbon: microbial nitrogen ratio; Cmic/Ctot = microbial quotient; Ctot = total C; Ntot = total N; C/N = carbon: nitrogen ratio; moisture = soil moisture content; biotic = average of biological soil properties (Cmic; Nmic; ADA; Cmic/Nmic); abiotic = average of chemical soil properties (Ctot, Ntot, C/N).

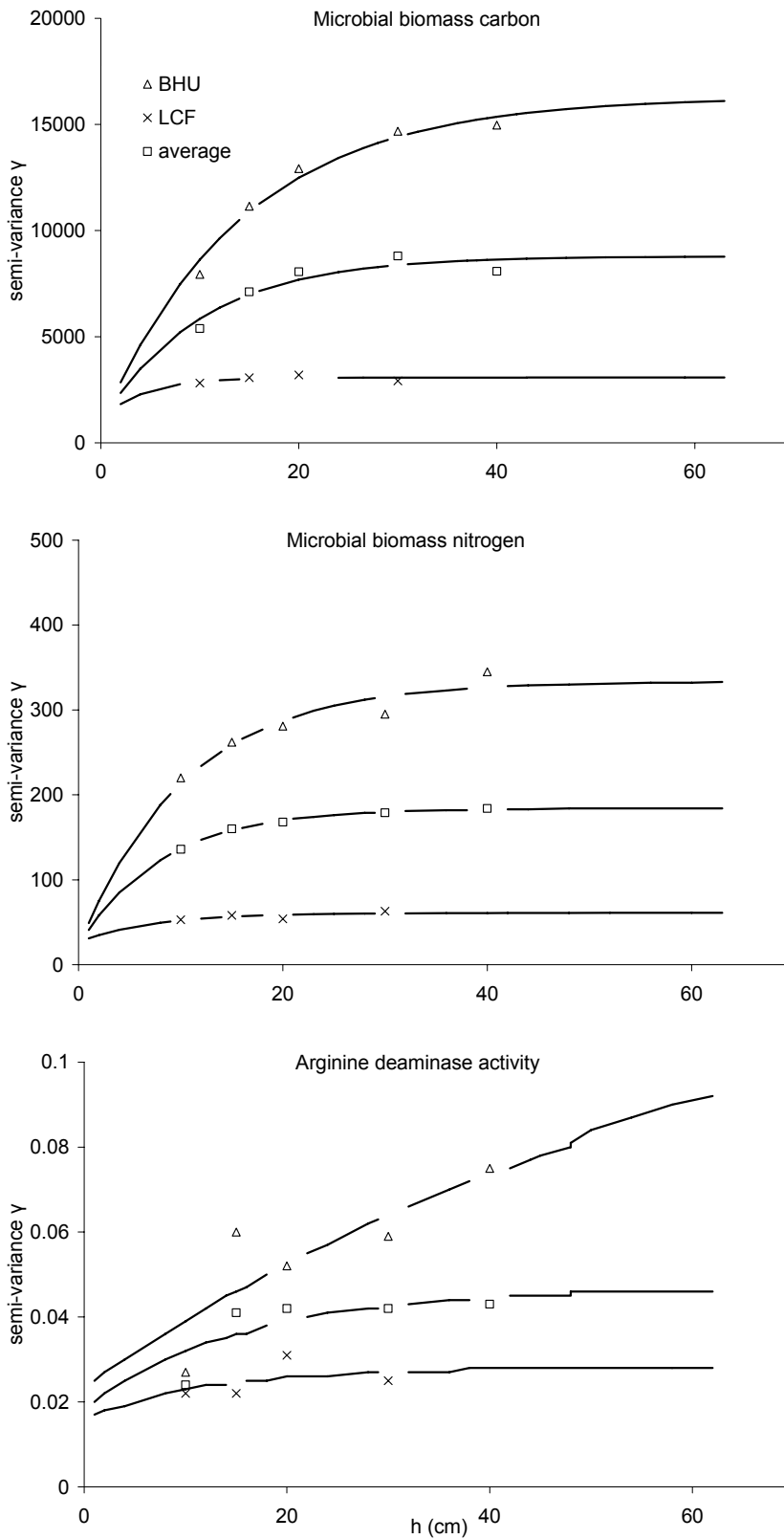


Fig. 3.

Patterns of semivariance (γ) in C_{mic} and N_{mic} and in ADA with increasing distance (h) between sampling points for ORG (distance classes 0-40 cm), CON (0-30 cm) and average (0-40 cm). Best-fit lines (based on exponential model) are shown for ORG, CON and average. (ORG, CON = average of three replicates, average = overall average).