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Holocene paleoecology and paleoceanography of the southwestern Black Sea shelf revealed by ostracod assemblages

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Abstract

The Holocene replacement of Ponto-Caspian ostracod assemblages by Mediterranean species is studied in two long composite cores, M02-45 (a composite of cores M02-45P, M02-45T and M05-03P) and M05-50 (a composite of cores M05-50P and M05-51G), acquired at sites -69 m and -91 m deep on the southwestern Black Sea shelf. Composite core M02-45 was collected from the middle shelf and composite core M05-50 was acquired on the distal fringe of the eastern levée of a saline underflow channel emanating from the Strait of Bosphorus. Sixteen radiocarbon dates in M02-45 and nine in M05-50 are used to construct age models, which show recovery of sediments as old as 12915 cal yr BP (M02-45 site) and 12010 cal yr BP (M05-50 site). A total of 45 ostracod species are identified in the two cores. From ~12000 to ~7425 cal yr BP, the ostracod assemblage is dominated by Ponto-Caspian species, mainly *Loxococoncha sublepada*, *L. lepida* and *Tyrrhenocythere amnicola donetziensis*. From ~7425 to ~6315 cal yr BP the assemblage consists of nearly equal abundances of Mediterranean species (*Cytheroma variabilis* in M02-45; *Sagmatocythere littoralis* in M05-50) and the Ponto-Caspian species. After ~6315 cal yr BP to the tops of the cores, the assemblage is dominated by Mediterranean species, including *Palmoconcha agilis*, *Carinocythereis carinata*, *Hiltermannicythere rubra* and *Pterygocythereis jonesii*. Cluster analysis further subdivides the stratigraphic succession into six bioecozones with different ostracod assemblages. The changes in the ostracod assemblages from one bioecozone to the next indicate that progressive environmental changes took place on the southwestern Black Sea shelf from at least 7500 cal yr BP to the present. The first hint of changing conditions at ~7500 cal yr BP lags the initial reconnection to the Mediterranean Sea through the Strait of Bosphorus by ~2000 yr, demonstrating that Black Sea salinity increased slowly and took that long to reach values tolerable to marine ostracod immigrants. Widespread

colonization by Mediterranean species took even longer, ~3000 years from the time of the initial reconnection.

Key words: Ostracods, Black Sea, salination, environmental change, Holocene paleoecology, ecological watermasses

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1. Introduction

The Black Sea is the largest permanently anoxic basin in the world (Murray and İzdar, 1989), situated between the Pontic Mountains of northern Turkey to the south, the Caucasus and Crimea Mountains of Russia and Ukraine to the north and northeast, and the Danube, Dneister, Dneiper and Bug alluvial plains to the west and northwest (Fig. 1). It is connected to the Aegean Sea and the eastern Mediterranean Sea through the Strait of Bosphorus, the landlocked Marmara Sea and the Strait of Dardanelles, collectively referred to as the Marmara Sea Gateway. A two-way flow exists between the Mediterranean Sea, Aegean Sea and Black Sea via the Marmara Sea Gateway (Latif et al., 1992). The Black Sea watermass consists of a low-salinity (17–20 practical salinity units (psu), hereafter unitless) 25–100 m-thick surface layer which flows at a velocity of 10–30 cm s⁻¹ into the northern Aegean Sea. Higher salinity (38–39) warm (15–20°C) Mediterranean water flows north and east across the gateway eventually entering the Black Sea where it contributes to a bottom watermass below the low-salinity, low-density surface water layer (Özsoy et al., 1995). At the northern Bosphorus exit, the underflow salinity is ~35 and at the shelf edge it is reduced to ~31 by entrainment of surface water (Özsoy et al., 2001). A prominent submarine channel begins at the Bosphorus exit and extends to the shelf edge northwest of the Bosphorus Canyon, accommodating the inflow of Mediterranean bottom water into the Black Sea (Fig. 2; Flood et al., 2009; Hiscott et al., 2013).

The Black Sea water column exhibits strong vertical stratification and there is little mixing between the bottom water layer and the upper oxygen-rich layer (i.e., a meromictic water body) because the intervening 50–200 m-deep halocline is deeper than the storm wave base. The warmer, higher-salinity, higher-density bottom water layer below a depth of 150–200 m is

permanently anoxic and enriched in hydrogen sulfide (Panin and Strehie, 2006; Eckert et al., 2013).

The Holocene connection history of the Black Sea to the eastern Mediterranean Sea has been vigorously debated, as summarized in Ryan et al. (2003), Hiscott et al. (2007a), Yanko-Hombach et al. (2007) and Yanchilina et al. (2017). Remaining disagreements can be understood by comparing Yanchilina et al. (2017), who advocate an abrupt incursion of saline water into a lowstand Black Sea taking a few decades to raise its level from ~ -120 m relative to modern sea level (rmsl) to ~ -30 m rmsl, to studies by Lister et al. (2015), Aksu et al. (2016) and Hiscott et al. (2017) which assert that the Black Sea was already approaching a highstand and was flowing out into the Marmara Sea and beyond by the earliest Holocene, so that the first input of saline water advanced northward as a salt wedge and underflow through the Strait of Bosphorus, beneath the outflow, leading to a progressive and slow salination.

2. Methods

Several piston and gravity cores were collected from the southwestern Black Sea shelf using the RV *Koca Piri Reis* of Dokuz Eylül University (Figs. 1, 2). The piston-coring equipment consisted of a Mooring Systems Inc.TM Kullenburg-type piston corer (original BenthosTM design) with split piston option, 1000 kg head, and ~ 4.5 m free-fall. Five cores were selected for further detailed studies: piston cores M02-45P, M05-03P, M05-50P and gravity cores M02-45T and M05-51G. Two of these cores (M05-50P and M05-51G) are located on the outer fringe of the eastern levée of a saline underflow channel (Flood et al., 2009; Hiscott et al., 2013), while three cores (M02-45P, M02-45T and M05-03P) are located on the middle shelf, ~ 66 km northwest of the saline channel near the Turkish–Bulgarian border. Cores M02-45T and M02-45P are trigger-weight and piston cores acquired <1 m apart during the same deployment.

Cores M02-45P/T and M05-03P penetrated the seabed ~500 m apart, while cores M05-50P and M05-51G penetrated the seabed ~310 m apart (Figs. 3, 4). On the basis of visual core descriptions and carbon and sulfur elemental and isotopic data, a 50 cm core-top loss is calculated for M05-50P relative to M05-51G (Linegar, 2012). A composite core for this site (hereafter designated as M05-50) was created by adding data for the upper 50 cm of core M05-51G to the top of core M05-50P. A similar exercise showed a 110 cm core-top loss for M02-45P relative to the co-located M02-45T (Hiscott et al., 2007b). During a later campaign, piston core M05-03P was acquired in an effort to extend recovery downward to the α unconformity (Fig. 3). Precise matching of elemental and isotopic trends for M02-45P/T with those for M05-03P, and radiocarbon dates, allows correlation of all three cores to within <5 cm uncertainty (Fig. 5; Reynolds, 2012). This correlation indicates that M05-03P recovered an estimated 78 cm of sediments older than the base of core M02-45P. In this paper, data from the basal 78 cm of core M05-03P are added to data from the M02-45P/T pair to form what is hereafter referred to as the M02-45 composite.

Approximately 20–25 cm³ of sediment were taken from cores M05-50P and M05-51G at 10 cm intervals to complete the M05-50 composite. Cores M02-45P and M02-45T were sampled at less regular intervals to fill the gaps in an earlier study of the same cores by Evans (2004; discussed later). Finally, the lowermost 70 cm of M05-03P was sampled at 10 cm intervals to complete the M02-45 composite. The samples were first oven-dried at ~25°C and the dry weights were recorded. They were then put into small plastic containers and treated with ~70 ml of 1% Calgon solution and left for several days, gently shaken occasionally to disaggregate the sediments. Each sample was wet-sieved using a 63 μ m sieve. To prevent contamination, the sieve was carefully washed, dried front and back with pressurized air, and

visually inspected to remove adhering grains between successive samples. The $>63\ \mu\text{m}$ fractions were collected, dried in an oven at $\sim 25^\circ\text{C}$, then passed through a stack of four sieves ($355\ \mu\text{m}$, $180\ \mu\text{m}$, $125\ \mu\text{m}$ and $63\ \mu\text{m}$) to separate the sample into smaller more manageable portions for subsequent hand-picking of ostracod valves and carapaces.

Each sample was examined under a stereoscopic microscope. Ostracod valves were hand-picked using a size 000 paint brush wetted with water. All valves that could be found in the samples were collected. Fragments were also collected; however, fragments that were less than $\sim 30\%$ of the original valve, or fragments inadequate for identification, were not picked. All instar stages (i.e., adults and juveniles) of the ostracods were also picked.

The ostracod valves are well-preserved throughout the cores. Because no internal soft-part preservation was observed, the ostracods were identified based on gross morphology of the valves. Shell shape and size, surface ornamentation, types of marginal pore canals, and internal features including the hinge, muscle imprints, inner lamella, and vestibule are keys to identifying ostracod taxa. Examination of valve morphology was done mainly under a stereoscopic microscope. Transmitted light microscopy was also used to observe characteristics not discernible with a stereoscopic microscope, mainly marginal pore canals. Ostracods have nine growth stages: eight juvenile stages and one adult stage (Athersuch et al., 1989). Thus, one individual ostracod may contribute up to 18 valves to the sediment over its lifetime. It is impossible then to really know how many individuals are actually represented in a sample because it will usually contain disarticulated valves as well as articulated carapaces and probably various juvenile stages. In this study, one valve (i.e., half of a full carapace) is counted as 1, while a full carapace is counted as 2 (Boomer et al., 2003). The ecological affinities of the ostracods present in this study can be differentiated into freshwater, brackish and marine (Puri et

al., 1964; Schornikov, 1964, 1966, 1969; Bonaduce et al., 1975; Athersuch et al., 1989; Stancheva, 1989; Boomer et al., 1996, 2005, 2010; Opreanu, 2008; Zenina et al., 2017).

3. Stratigraphy and age models

3.1 Core Descriptions

Composite core M05-50 consists mostly of Holocene sediments on the eastern bank of a submarine channel north of the Bosphorus exit (Figs. 2, 4). The original core descriptions were published by Flood et al. (2009). The composite core is 787 cm long (Fig. 6). Bioturbation is only moderate in core M05-50 and is not believed to affect ostracod distribution at the 10 cm vertical sample spacing used for this study. Three lithostratigraphic units are defined. Unit 3 extends from the base of the composite core to an unconformity at 695 cm depth which corresponds to the α_1 reflector in high-resolution seismic data (Figs. 4, 6). This unit is comprised of interbedded silty mud and graded beds of coarse silt to mainly very fine sand. The brackish water bivalve *Dreissena* is abundant in some of the graded beds (Flood et al., 2009). Unit 2 extends from 695 cm to 95 cm and consists of moderately bioturbated silty mud ($\leq 10\%$ sand) with scattered marine mollusc shells above a composite depth of 513 cm. Unit 1 extends from 95 cm to the top of core M05-50 and is distinguished from Unit 2 by its color banding. The texture is silty mud.

Composite core M02-45 consists of Holocene sediments on the middle shelf ~66 km northwest of the M05-50 site (Figs. 1, 3, 6). Water depth at the site is -69 m. Except for the lower 78 cm, the original core descriptions were published by Hiscott et al. (2007b). The composite core M02-45 is 1028 cm long. Four lithologic units are recognized (Fig. 6); these are given alphabetical identifiers to distinguish them from the units in composite core M05-50. The oldest Unit D extends from the base of the core to 970 cm composite depth. The base of the unit

correlates with the seismic reflector α (Figs. 3, 6). The lowermost 10 cm of Unit D consists of mud containing scattered pebbles and cobbles, and is succeeded by alternating horizons of sandy mud, shell hash and very coarse sand and gravel. Unit C extends from 970 cm to a composite depth of 480 cm. Its top therefore correlates to seismic reflector α_1 (Fig. 3). Unit C consists of color-banded mud with graded laminae and beds of silt to very fine sand (Fig. 6), and scattered shells of *Truncatella subcylindrica*, *Monodacna pontica*, *Didacna* spp., *Dreissena polymorpha* and *D. rostriformis* Hiscott et al. (2010). It was divided by Hiscott et al. (2007b) into subunit C1 below 615 cm and subunit C2 above that depth based on geochemistry. Unit B extends upward from 480 cm to 270 cm, the latter just below a <5 cm-thick shelly horizon which correlates in the seismic data to unconformity α_2 . Unit B consists of alternating horizons of mud and shelly mud (Fig. 6). The abundance of bioclastic sand locally exceeds 20%. The mollusc assemblage includes *T. subcylindrica*, *Mytilus galloprovincialis*, *Parvicardium exiguum*, *Rissoa* spp. and *Modiolula phaseolina* (Hiscott et al., 2007b). Unit A represents the youngest sediments in the composite core. It consists of color-mottled/banded, burrowed mud with silt laminae and scattered shells of several immigrant Mediterranean molluscs: *Bittium reticulatum*, *Spisula subtruncata*, *Acanthocardia paucicostata*, *Abra alba*, *M. galloprovincialis*, *T. subcylindrica*, and *Turritella communis*.

3.2 Radiocarbon dates and age model

A total of 33 radiocarbon ages were obtained from composite cores M05-50 (13 dates) and M02-45 (20 dates) (Table 1). Eight and thirteen of the uncalibrated dates from M05-50 and M02-45 were originally published by Flood et al. (2009) and Hiscott et al. (2007b), respectively. Transposing the stratigraphic record into a chronological framework requires the conversion of the raw radiocarbon dates (reported here with the units ^{14}C yr BP) to the astronomical timescale

(with units of calendar years BP, or cal yr BP). For this task, we used the Marine13 calibration curve (Reimer et al., 2013) and estimates of the age of the contemporary inorganic carbon reservoir from which biogenic carbonate was precipitated. The Marine13 curve incorporates a modelled, time-varying reservoir age (R) of ~ 400 yr, but adjustments (ΔR) are required based on the composition and history of the local watermass (controlled in part by residence times of carbonate and bicarbonate ions in deeper waters, and any hard-water effect through supply of old carbon by rivers). Reservoir age is defined (Stuiver and Braziunas, 1993) as the difference in radiocarbon age between (a) marine precipitates (e.g., shells) or organic matter and (b) contemporaneous terrestrial organic material (e.g., wood) or precipitates (e.g., speleothems) that formed in contact with the atmosphere. The reservoir age of the modern Black Sea is ~ 415 yr (Siani et al., 2000), but appropriate reservoir ages prior to reconnection with the global ocean are controversial.

Kwiecien et al. (2008) and Soulet et al. (2011) advocated applying a time-varying reservoir age in the range 0–300 ^{14}C yr to pre-reconnection radiocarbon dates of Black Sea shells back to ~ 12.0 ^{14}C ka (Fig. 7). Mudie et al. (2014) warned that the pre-reconnection reservoir age might have been, at times, >1120 ^{14}C yr, but could not be firm on this assessment because of the possibility that a *D. polymorpha* shell in their study was reworked. Yanchilina et al. (2017) departed in a significant way from these earlier studies by arguing that the time-varying record of the Black Sea reservoir age could be determined by tuning (a) the $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ stable isotopic records for radiocarbon-dated shells secreted in the Black Sea to (b) the U/Th-dated stable isotopic records from stalagmites of the Sofular Cave, located immediately inland of the southern Black Sea coast. This novel approach is based on an expectation that carbon and oxygen incorporated into the cave dripstones would inherit their isotopic signatures from regional

vegetation patterns (distribution of C₃ versus C₄ plants) and from water vapour in equilibrium with the surface waters of the adjacent sea. Yanchilina et al. (2017) provide supplementary data and graphs to support their preferred best match between the Black Sea and the Sofular Cave $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ records. The graphs (their supplementary file mmc5.pdf) employ scales of uncorrected (raw) ^{14}C ages for shells and calendar ages for the cave deposits, making it difficult to judge how application of time-varying reservoir ages might compensate for mismatches of key features in the graphs.

The validity of the Yanchilina et al. (2017) tuning is critical to the development of any late Quaternary age model (e.g., for core M05-50) because the rapid changes in reservoir age which they advocate effectively stretch or compress the resultant time scale. The large excursions and rapid changes in reservoir age proposed by those authors were independently assessed by replotting their supplementary data for carbon and oxygen isotopes (Fig. 8). The U/Th age for each Sofular Cave data point was converted to an equivalent atmospheric radiocarbon date using the definition table for the IntCal13 calibration curve (Reimer et al., 2013). The isotopic values for the Sofular Cave stalagmites were plotted against these atmospheric ^{14}C ages. Isotopic values for Black Sea molluscs were then superimposed, first as plots against uncorrected ^{14}C dates for each shell (Fig. 8 a,b), second after subtracting Yanchilina et al. (2017) *R* values (Fig. 8 c,d) and third after subtracting *R* values similar to those used by authors AEA and RNH in earlier work (e.g., Bradley et al., 2012; Mertens et al., 2012) (Fig. 8 e,f). For ^{14}C dates of ~9500–12000 ^{14}C yr BP, reservoir ages preferred here (Fig. 7) are similar to those proposed by Yanchilina et al. (2017) because of the need to match the prominent trough in $\delta^{13}\text{C}$ values in the Bølling/Allerød warm interval. However, in the critical period surrounding the reconnection of the Black Sea to the global ocean, reservoir ages proposed by Yanchilina et

al. (2017) do not provide a better match to the Sofular Cave data than author-preferred values, largely because the considerable scatter in both data sets does not allow unambiguous ties between (a) peaks, troughs or inflections in the marine data set and (b) features in the cave records. In the absence of truly compelling justification for the large swings in reservoir age proposed by Yanchilina et al. (2017), we have elected to undertake calibration of radiocarbon dates using a gradual increase in R from its pre-reconnection minimum of ~ 50 ^{14}C yr at ~ 10200 ^{14}C yr BP to reach 19th–20th century values (Siani et al., 2000) by the time euryhaline molluscs populated the Black Sea shelves (~ 7100 ^{14}C yr BP according to Ryan et al., 2003; Hiscott et al., 2007b). R values for dates in the range 8400–9400 ^{14}C yr BP are lower than those employed by the authors in earlier calibrations (Bradley et al., 2012; Mertens et al., 2012). After 7100 ^{14}C yr BP, we have used the default Marine13 calibration with a fixed ΔR value of +10 years.

Nine calibrated ages from cores M05-50P and M05-51G were used to construct an age model for the composite core M05-50 (Fig. 6). The age at the top of the core is assumed to be zero. The ages assigned to depths of 695 and 696 cm were extrapolated using constant sedimentation rates between dated intervals to either side of the $\alpha 1$ unconformity. The $\alpha 1$ surface occurs between 695 and 696 cm; the conformable surface $\alpha 2$ occurs at ~ 595 cm depth in the composite core (Fig. 6). Ignoring the $\alpha 1$ hiatus, the average sedimentation rate determined from the age model is 82 cm ka^{-1} . For samples spaced each 10 cm, temporal resolution is ~ 240 yr since 2000 cal yr BP, ~ 60 – 200 yr from 2000–4500 cal yr BP, then ~ 125 – 140 yr for the older parts of the cored succession. The $\alpha 1$ hiatus has a duration of 2200–2700 years depending on how the rather large \pm uncertainties for deeper ages (Table 1) are combined to guide the upward extrapolation to 696 cm composite depth. It is noteworthy that tracing of seismic reflections just below $\alpha 1$ in the vicinity of the M05-50 site indicates that as much as ~ 1.3 m more section is

present a short distance away, as the unconformity rises upward. The temporal resolution of 125–140 yr translates to an accumulation rate below α_1 of ~ 75 cm/10³ yr. Therefore, 1.3 m of sediment might indicate that α_1 developed in this area after ~ 9050 cal yr BP, rather than after 10755 cal yr BP as estimated at the core site itself (Table 1).

Sixteen calibrated ages from composite core M02-45 were used to construct an age model (Fig. 6). The ages assigned to depths 270 and 271 cm (straddling unconformity α_2) were extrapolated using constant sedimentation rates between dated intervals above and below that level, leading to the conclusion that the α_2 hiatus has a duration of ~ 3390 years. The conformable surface α_1 occurs at ~ 480 cm depth in the composite core (Fig. 6) where the interpolated age is 7260 cal yr BP. For samples spaced each 10 cm, temporal resolution is ~ 120 yr since ~ 9100 cal yr BP, ~ 30 – 50 yr from 10000–9100 cal yr BP, then ~ 500 yr for the older parts of the cored succession. The last value implies a very reduced accumulation rate close to the basal α unconformity.

3.3 Implications of the α , α_1 and α_2 unconformities

The α unconformity truncates a wide range of older strata, including Eocene–Pliocene tilted and deformed strata (Aksu et al., 2002b). It is a relatively flat surface with strong reflectivity in seismic profiles. Where exposed (Flood et al., 2009, their figure 16) or cored, there is a layer of gravel and/or coquina. Sediments below α on the southwestern shelf are generally older than 30,000 ¹⁴C yr BP (Fig. 9). Therefore, α is interpreted as a lowstand subaerial unconformity modified by the last post-glacial transgression. But, α_2 has an entirely different interpretation. At the M02-45 site it overlies sediment as young as 5445 cal yr BP when the Black Sea was fully connected to the world ocean and stood at ~ -5 m relative to modern sea level (rmsl; Lambeck et al., 2007). Hence, water depth at the M02-45 site was little

different to today and $\alpha 2$ has to be a submarine erosional surface, probably created by strong along-shelf currents associated with the Rim Current in the Black Sea (Flood et al., 2009). The $\alpha 1$ surface is believed to have a similar origin to $\alpha 2$. It becomes a conformable surface (or imperceptible disconformity) near the M02-45 site and is underlain by sediment as young as 7260 cal yr BP. As explained above, sediment underneath $\alpha 1$ might be as young as 9050 cal yr BP near the M05-50 site. The level of the Black Sea was ~ -10 m rmsl at 7260 cal yr BP, and ~ -23 m rmsl at 9050 cal yr BP (Lambeck et al., 2007), the latter several hundred years after entry of the first Mediterranean water into the Black Sea (Yanchilina et al., 2017). Hence almost the entire southwestern shelf was apparently under tens of meters of water during $\alpha 1$ development and it must also be a subaqueous unconformity (to local conformity). Furthermore, there is no evidence in split cores of subaerial exposure; e.g., rooting, desiccation cracks, caliche nodules, other pedogenic textures, or changing water content.

4. Ostracod results

Ostracods are abundant and well-preserved throughout composite cores M05-50 and M02-45, with a total of 45 ostracod species. In composite cores M05-50 and M02-45 all ostracods found in the samples were handpicked and counted (Fig. 10). The ostracod assemblages in the composite cores are described using key ostracod species (Table 2; Figs. 11, 12). Key species are considered to be those which are consistently abundant, occur in most samples through particular core intervals and are represented by both adults and several stages of juveniles (see below). They can therefore be assumed to have lived and died at the core site, as opposed to having been reworked or transported to the site, and are therefore more reliable for reconstructing ecological changes in the study area (De Deckker, 2002; Boomer et al., 2003). Taphonomic assessment using population age diagrams (Fig. 13) is a useful precursor to using

ostracods to interpret paleoenvironments (De Deckker, 2002; Boomer et al., 2003). By plotting how many valves of a particular species are present, from adults down to the smallest juveniles, an assessment of the taphonomic history of the assemblage can be made (Boomer et al., 2003). The population age diagrams for core M05-50 show that all assemblages are autochthonous (Fig. 13). The presence of various stages of juveniles in most samples indicates a low to moderate energy system.

4.1 Composite core M05-50

Three temporally distinctive ostracod assemblages are delineated in core M05-50: Ponto-Caspian assemblage, transitional assemblage and Mediterranean assemblage.

Ponto-Caspian (brackish) assemblage (780–630 cm)

Samples from 780–630 cm depth in core M05-50 contain on average 96.9% specimens commonly found in brackish waters of the Ponto-Caspian basins (Fig. 14; e.g., Schornikov, 1969; Opreanu, 2008; Boomer et al., 2010). The brackish assemblage is overwhelmingly dominated by *L. sublepada* and *L. lepida* (Fig. 11, 12), in that these two species make up an average of 63.5% of each sample, with a maximum abundance of 81.5% at 660 cm. Rare occurrences of *S. littoralis* are observed with abundances of < 2%. The fact that these "flickering" occurrences are present for at least 1.5 m below the overlying *S. littoralis* peak rules out bioturbational mixing from higher levels, as does the lack of an upward increase in abundance that would be expected with bioturbation. If these rare occurrences were a coring artifact because of smearing of younger sediment along the core liners, then one should expect to see flickering occurrences of, for example, *C. carinata* and *H. rubra*, which are not recorded.

Other significant species in the Ponto-Caspian assemblage are *T. amnicola donetziensis* (averaging 11.2%), *Amnicythere olivia* (5.4%), *Graviacypris elongate* (4.9%) and *A.*

quinquetuberculata (4.3%; Fig. 14). Species sporadically appearing in the brackish assemblage in low abundances of <2% are *Euxinocythere bacuana*, *Amnicythere pediformis*, *Amnicythere* sp. aff. *subcaspia*, *Amnicythere striatocostata*, *Amnicythere cymbula*, *Euxinocythere* sp. aff. *relicta*, and *Loxoconchissa* (*Loxocaspia*) *immodulata* (Williams, 2012). The marine species *Palmoconcha agilis* (which is closely similar to the "granulata" form of *P. guttata* from the NW European coasts: the two might be considered synonymous) is present at most depths in the brackish section, but in extremely low abundances, typically only one or two valves per sample. Given the high abundances of this species in upper parts of the core (Fig. 14), contamination from mud smear along the core liner cannot be ruled out.

The $\alpha 1$ unconformity occurs within the brackish section at a depth of 695 cm in composite core M05-50 (Figs. 4, 14). It represents an ~2500-year hiatus (averaging from 10725 to 8420 cal yr BP). The ostracod assemblage before and after this hiatus changes very little, but some differences are noteworthy: for example, *L. sublepidata* and *L. lepidata* increase from 55.6% to 70.0% from 730–700 cm depth (~200 years preceding the hiatus) to 690–670 cm depth (~250 years after the hiatus; Fig. 14). Other changes over the same interval involve *T. amnicola donetziensis* which decreases from 18.9 to 5.6% and *A. olivia* which decreases from 7.4 to 2.6%.

Transitional assemblage (620–540 cm)

Samples from 620–540 cm depth show co-occurrence of brackish and marine ostracod species (Fig. 14). From 630 cm to 620 cm depth in the core the Mediterranean species *S. littoralis* (Fig. 9) dramatically increases in abundance from 1.7% to 40.6%, and within the transitional assemblage it maintains an average abundance of 41.6 %, making it the dominant species.

Brackish water species remain abundant in the transitional section (Fig. 14). *Loxococoncha sublepada* and *L. lepida*, which had been the dominant species in the brackish assemblage, become the second most abundant species in the transitional assemblage with an average abundance of 33.3%. Likewise, *T. amnicola donetziensis*, which had been the second most abundant species in the brackish section, is still present in the transitional assemblage but its average abundance decreases from 11.2% to 5.8% as a result of dilution by *S. littoralis*, making *T. amnicola donetziensis* the third most abundant species.

The number of brackish species is still greater than the number of marine species. However, given the high number of specimens of the Mediterranean species *S. littoralis*, the average ratio of marine to brackish specimens is calculated to be 60:40 in the transitional section. In the earliest stages of the transitional section the Mediterranean species *C. carinata*, *H. rubra* and *C. variabilis* first appear albeit only a few individuals in core M05-50. The marine species *P. agilis* maintains a low percentage (2.7%) in the transitional assemblage.

Mediterranean (marine) assemblage (530–0 cm)

At 530 cm depth in core M05-50, the Mediterranean species *S. littoralis* decreases dramatically from 55.4% abundance at 540 cm depth to only 2.5% at 530 cm depth. Between the same two sample depths, the Ponto-Caspian species *L. sublepada* and *L. lepida* together decrease from 28.4% to 3.1%. Above this depth, all Ponto-Caspian species are supplanted by the Mediterranean ostracods *P. agilis*, *H. rubra*, and *C. carinata* together constituting 89.4 % of the 530 cm sample (Fig. 14). Together with other marine ostracods, these marine species dominate (at 99.2%) the ostracod assemblage from that point upward to the top of the core. *Sagmatocythere littoralis* all but disappears in the marine assemblage, occurring only a few times represented by a valve or two. *Loxococoncha sublepada* and *L. lepida* do occur often in the marine

assemblage but in extremely low abundances of < 1 %, and mainly below a depth of 250 cm (Fig. 14). These could be reworked. *Tyrrhenocythere amnicola donetziensis* also occurs in the marine assemblage in abundances typically 1% or less and most frequently between the depths 440 cm and 320 cm. By far the most abundant taxon in the entire marine assemblage is the Mediterranean species *P. agilis* (Fig. 14). Valves of this species make up an average of 56.6 % of each sample from 530 cm upward. The other significant species present in the marine assemblage of core M05-50 are *Costa edwardsi* and *P. jonesii*, with smaller abundances of *Paracypris polita*, *Leptocythere multipunctata*, *Callistocythere diffusa*, *C. variabilis*, *Bythocythere* sp. and *Xestoleberis* sp. aff. *cornelii* (Fig. 14).

4.2 Composite core M02-45

There are several fundamental similarities and some differences between the assemblages in core M02-45 when compared to M05-50. A portion of core M02-45 was initially studied at 20 cm sampling intervals by Evans (2004) with preliminary results briefly summarized in Hiscott et al. (2007b). Although most of the ostracod species identified in core M05-50 are also present in core M02-45, some notable differences exist: for example *C. edwardsi*, *P. jonesii*, and *P. polita* are three Mediterranean species in the marine Mediterranean assemblage in core M05-50 which are not recorded in core M02-45. Furthermore, *S. littoralis* occurs in core M02-45 in only very low abundances in three widely spaced samples (Evans, 2004). In core M05-50, *S. littoralis* has a much more significant role essentially dominating the entire transitional assemblage. Despite these differences, three temporally distinctive ostracod assemblages similar in their implications to those identified in core M05-50 are present in core M02-45: a Ponto-Caspian assemblage, transitional assemblage and Mediterranean assemblage.

Ponto-Caspian (brackish) assemblage (1012–490 cm)

Samples from 1012–490 cm depth in core M02-45 contain on average 95.5% brackish-water Ponto-Caspian species (Fig. 15). In the lowermost 30–35 cm of the composite core (~975–1012 cm; lithologic Unit D) the brackish water assemblage is overwhelmingly dominated by *T. amnicola donetziensis*, averaging 48.1%, with notable contributions from *L. (Loxocaspia) immodulata*, *L. sublepada* and *L. lepida*, and *A. olivia* (Fig. 15). The middle portion of the assemblage between 975 and 700 cm depth in the core has a diverse Ponto-Caspian assemblage dominated by *G. elongata*, *L. sublepada* and *L. lepida* with notable contributions from *A. olivia*, *A. quinquetuberculata*, *E. bacuana* and smaller influxes of *A. striatocostata*, *Amnicythere* sp. aff. *subcaspia*, *Euxinocythere* sp. aff. *relicta* and *L. (Loxocaspia) immodulata* (Fig. 15). The upper portion of the assemblage from ~700 cm to 490 cm depth in the core is overwhelmingly dominated by *L. sublepada* and *L. lepida* and *G. elongata*, with minor contributions from *A. olivia*, *A. quinquetuberculata* and *E. bacuana* (Fig. 15).

Transitional assemblage (490–450 cm)

In the five samples from 490–450 cm depth, brackish and marine ostracod species co-occur (Fig. 15). The fauna are characterized by large abundances of marine species of *C. variabilis* (~60%) with decreasing yet notable abundances of Ponto-Caspian species *L. sublepada* and *L. lepida* and minor occurrences of *A. olivia*, *A. quinquetuberculata* (Fig. 15). *Palmoconcha agilis* and *H. rubra* also occur within the zone, with progressively upward increasing abundances. The most striking characteristic of the assemblage is the co-occurrence of a few key Mediterranean and Ponto-Caspian species. The transitional assemblage is observed across a narrow depth zone of only 40 cm, possibly raising the question: is this zone real or is it created by contamination? The high percentage of *C. variabilis* in sediment with ~10% *L. sublepada* and

L. lepida cannot be explained by downward smearing of *C. variabilis* from higher levels. It is possible that *L. sublepada* and *L. lepida* could be reworked or burrow-mixed into younger sediment from depths of >490 cm, but in that case why not other brackish ostracod species, such as *A. quinquetuberculata* and *G. elongata* to the same extent? Coring-induced contamination and bioturbational mixing can be ruled out because of the sharp downward disappearance of several marine species with effectively no "flickering" occurrences below, strongly suggesting that the transitional zone is indeed real.

Mediterranean (marine) assemblage (450–0 cm)

Above 450 cm depth in core M02-45, a notable faunal turnover takes place where the brackish-water Ponto-Caspian ostracod assemblage is completely replaced by the Mediterranean ostracod assemblage (Fig. 15). *Palmoconcha agilis* and *H. rubra* show dramatic increases upcore with abundances collectively exceeding 80–85%. The Ponto-Caspian species *L. sublepada* and *L. lepida* together decrease from ~10% to <1%. By far the most abundant taxon in the marine assemblage is the Mediterranean species *P. agilis* with significant contributions from *L. multipunctata*, *C. diffusa*, *C. variabilis*, *C. carinata* and *X. sp. aff. cornelii* (Fig. 15). In comparison to the marine Mediterranean assemblage seen in core M05-50, there are several species which are notably absent, including *C. edwardsi*, *P. jonesii*, *P. polita* and *S. littoralis* (Fig. 14). This issue is discussed below.

5. Cluster Analysis and bioecozones

CONISS (Constrained Incremental Sums of Squares) cluster analysis (Grimm, 1987) was run on the age-converted ostracod data from composite cores M02-45 and M05-50 using Tilia software (Grimm, 2011). In each core, the results show six well-defined clusters (Figs. 16, 17; Table 3). These are called "bioecozones", consistent with the definition of Poag and Valentine

(1976) that a bioecozone is "a body of strata characterized by a certain paleoenvironment that is inferred from its fossil content and that can be distinguished from the paleoenvironment of adjacent strata". A bioecozone may be time-transgressive.

Bioecozone 1 – brackish water environment

Bioecozone 1 is part of the lower brackish assemblage which occurs from the base of both composite cores M05-50 and M02-45 (Figs. 16, 17; Table 3). In cores M05-50 and M02-45 it is dated from 11910–10755 cal yr BP (truncated by unconformity α_1), and 11780–9350 cal yr BP, respectively. In both cores the most abundant species over these time intervals are *L. sublepada* and *L. lepida*, *T. amnicola donetziensis*, *G. elongata*, and *A. quinquetuberculata* (Figs. 16, 17; Table 3). These species and other less abundant ones such as *E. bacuana*, *A. pediformis*, *A. cymbula* and *L. (Loxocaspia) immodulata* (in core M02-45) are currently found living in the Caspian Sea which has a salinity range of 1–13. *Graviacypris elongata* prefers salinities of up to 8 (Aladin, 1993) and *A. quinquetuberculata* prefers salinities of up to 5 (Opreanu, 2008). *Loxoconcha lepida* is also found living in deltas around the Azov Sea where the maximum water depth is 14 m and salinity is very low, certainly far less than 10 at the coast. *Amnicothera cymbula* is found today in lagoons and estuaries in the Ponto-Caspian basins (Opreanu, 2008). On the basis of the upper salinity preferences of *G. elongata*, *A. quinquetuberculata* and *L. lepida*, Bioecozone 1 is interpreted to represent a low salinity environment, with salinities of <10, possibly as low as 5–8.

Bioecozone 2 – brackish water environment

In composite core M05-50 the base of Bioecozone 2 cannot be assigned an age because sedimentation after the α_1 hiatus did not begin until ~8390 cal yr BP. After the hiatus, Bioecozone 2 extends to 7450 cal yr BP (Figs. 16, 17; Table 3). The α_1 surface is conformable

in composite core M02-45 (Hiscott et al., 2007b). There, Bioecozone 2 extends from 9350–7410 cal yr BP and is marked by notable upward changes in the ostracod fauna compared with Bioecozone 1. For example, *T. amnicola donetziensis* completely disappears, and Bioecozone 2 is dominated by *G. elongata*, *L. lepida*, and *L. sublepida*, with notable occurrences of other Ponto-Caspian species, including *E. bacuana*, *A. olivia* and *A. quinquetuberculata* (Fig. 17; Table 3). In the composite core M05-50 there are only small changes in this bioecozone relative to Bioecozone 1. *Loxoconcha lepida* together with *L. sublepida* together increase in abundance and *T. amnicola donetziensis* and *A. olivia* decrease in abundance (Table 3). The ostracod data suggest that the brackish environment persisted on the southwestern Black Sea shelf until ~7425 cal yr BP.

Bioecozone 3 – transitional environment

In both cores, Bioecozone 3 marks a major change in the ostracod assemblage (Figs. 16, 17; Table 3). In the composite core M02-45, this interval extends from 7410–6310 cal yr BP and is notably characterized by the influx of marine species *C. variabilis* with significant contributions from *P. agilis* and *H. rubra* and minor showings of *L. multipunctata* and *X. sp. aff. cornelii* (Figs. 17, 18; Table 3). In core M05-50 this interval begins with the first significant occurrence of marine ostracods (*S. littoralis*) at ~7425 cal yr BP and extends upward to the end of the co-occurrence of marine and brackish species at ~6320 cal yr BP (Figs. 17, 18). The change in the assemblage at the base of the bioecozone happens rather abruptly in geological terms, considering that it occurs within 10 cm of sediment representing ~125–140 years. If the species *S. littoralis* is truly stenohaline, as asserted by Ruggieri (1964) in studies near Naples, Italy, then it stands to reason that when the water reached precise conditions favored by *S. littoralis* this species was able to rapidly expand its population from only one or two specimens

per sample in Bioecozone 1 to ~40 % in Bioecozone 3. Another possibility is that *S. littoralis* is simply an opportunistic species and was the fastest to colonize the new environment at the northern exit of the Bosphorus Strait, and took advantage of the lack of competition from others. Ivanova et al. (2007) regard this species as preferring salinities up to 18 so this might be an upper limit for the salinity around the core site toward the end of the Bioecozone 3 interval at ~6320 cal yr BP. However, even until this late date the brackish species *L. sublepada*, *L. lepida* and *T. amnicola donetziensis* also inhabited the site. The coexistence of brackish and marine species suggests that salinity was rising but remained toward the low end of the range preferred by the marine immigrants. Other marine ostracods, particularly *P. agilis*, were present in the later stages of the brackish interval along with *S. littoralis* (Fig. 16; Table 3). However, when the opportunity afforded by increasing salinity developed, *S. littoralis* was the first of the marine species to take advantage and colonize the area. It is possible that the salinity at that time was in the exact range suitable for *S. littoralis* to thrive. However, that particular salinity range was only sustained for ~1000 years after which other Mediterranean species began to replace *S. littoralis*.

Although the relative abundance of *S. littoralis* is lower at core site M02-45, the transitional interval is marked by Mediterranean species co-existing with lingering Ponto-Caspian brackish species for ~1000 years (Figs. 16–18). This observation supports the idea that the changes in water-mass characteristics and ecological parameters were protracted.

Bioecozone 4 – earliest marine environment

In composite core M05-50, Bioecozone 4 starts at ~6320 cal yr BP and marks the time when the brackish assemblage was completely displaced by marine species immigrating through the Strait of Bosphorus (Fig. 16; Table 3). The pioneering *S. littoralis* was rapidly replaced at

this time by new Mediterranean species and the ostracod assemblage in the core became fully marine. The first ostracods to dominate the new marine assemblage in Bioecozone 4 were *P. agilis*, *C. carinata* and *H. rubra* (Fig. 16; Table 3). These species are found living today around the Mediterranean and adjoining seas where the salinities are 37–39. *Palmoconcha agilis* is found in the Adriatic Sea (Bonaduce et al., 1975) and it is abundant today around the shallow (10–100 m) shoreline of the Black Sea (Schornikov, 1967). *Carinocythereis carinata* can be found today all around the Atlantic Ocean and Mediterranean Sea and prefers depths of ~60–80 m (Keen, 1982; Athersuch et al., 1989). *Hiltermannicythere rubra* lives on the shallow banks around Ischia, Procida, Naples and the Sorrento Peninsula at depths up to 100 m (Puri et al., 1964).

In composite core M02-45, Bioecozone 4 begins at ~6310 cal yr BP, and is marked by a near complete disappearance of Ponto-Caspian brackish water species, and dominance by Mediterranean species (Fig. 17; Table 3). *Cytheroma variabilis* which dominated the transitional Bioecozone 3 is notably reduced in abundances while *P. agilis* becomes the dominant species with significant contributions from *H. rubra* and to a lesser extent *C. diffusa*. Lingering occurrence of *G. elongata* is found in one sample.

Bioecozone 4 is the earliest marine stage of the Holocene during which there was increasingly saline bottom water on the southwestern Black Sea shelf due to steady Mediterranean inflow. Salinity had risen to a point where it was unfavorable for brackish Ponto-Caspian species and they soon all but disappeared as Mediterranean ostracods began to diversify and flourish.

Bioecozone 5 – middle Holocene marine environment

Bioecozone 5 is marked by a further increase in salinity indicated by the introduction of two new marine species in core M05-50 (Fig. 16). At 4310 cal yr BP, *C. edwardsi* and *P. jonesii* both appear for the first time in core M05-50 and persist as two of the more common species in the marine assemblage. *Pterygocythereis jonesii* prefers salinities of 26–35 (Neale, 1988). *Costa edwardsi* and *P. jonesii* are both sublittoral species (Athersuch et al., 1989) and *P. jonesii* commonly occurs in water well over 80–100 m deep (Bonaduce et al., 1975; Penny, 1993). These two Mediterranean species are notably absent in composite core M02-45 (Fig. 17), where the fauna are dominated by *P. agilis*, but also show sustained occurrences of *H. rubra*, *C. diffusa*, *C. variabilis*, *L. multipunctata*, *Leptocythere spp.*, and the first introduction of *C. carinata* (Fig. 17; Table 3). The base of Bioecozone 5 at the M02-45 site is younger than at the M05-50 site because of missing section at the α_2 unconformity and so begins at ~2055 cal yr BP rather than at 4310 cal yr BP.

The Mediterranean species *C. edwardsi*, *P. jonesii* and *P. polita* prefer high salinities as indicated by their late arrival at core site M05-50 (Fig. 16). As with *S. littoralis*, these species are present at core site M05-50 but not at core site M02-45 simply due to the close proximity of the M05-50 site to the Strait of Bosphorus. A slightly lower salinity at core site M02-45 is not likely to be a contributing factor because these species were present at site M05-50 since ~4300 cal yr BP (Fig. 17) when the salinity on the Black Sea shelf was lower than today (Mertens et al., 2012).

Bioecozone 6 – late Holocene marine environment

Bioecozone 6 started at ~1590 cal yr BP at core site M02-45 and ~2390 cal yr BP at core site M05-50 (Figs. 16–15; Table 3), so its base is more diachronous than for other bioecozones.

Bottom water salinities across the southwestern Black Sea were probably very similar to modern values during Bioecozone 6 (see also Soulet et al., 2010). This bioecozone is defined by a decline in the marine species *P. agilis* and *H. rubra* which are common to Bioecozones 4 and 5. *Palmoconcha agilis* has an average abundance of 63.1% in Bioecozones 3 and 4 and in Bioecozone 6 it decreases to 42.6%. There is also a major decline in *L. multipunctata*, *X. sp. aff. cornelii* and *C. variabilis* whose abundances all drop well below 1% in Bioecozone 6. In core M05-50, *P. jonesii* and *C. edwardsi* become more abundant than previously and *P. polita* becomes much more common (Fig. 16; Table 3).

6. Discussion

The ostracod data obtained from composite cores M05-50 and M02-45 show that a distinct faunal turnover occurred from a brackish to a marine assemblage beginning at ~7400–7450 cal yr BP (Figs. 16–18). In both cores, the suddenness of the onset of this turnover, occurring in ~150 years, and its magnitude strongly suggest that it must have been driven by fundamental ecological changes at the core sites associated with the increasing salinity of the Black Sea following the reconnection to the Mediterranean Sea. The high faunal diversity (Evans, 2004; Williams 2012) excludes changes in oxygen availability as a cause of this turnover. Environmental preferences of the various taxa implicate climbing salinity as the primary control. It should be remembered that the turnover occurred almost 2000 yr after strontium-isotopic evidence for the initial reconnection of the Black Sea with the Mediterranean Sea (Yanchilina et al., 2017), and when global sea level and Black Sea level were in tandem at ~-12 m rmsl. Hence this rapid ecological event has nothing to do with the purported catastrophic flooding through the Strait of Bosphorus, but instead signals the attainment of a threshold beyond which marine ostracods could flourish and brackish-water ostracods became vulnerable.

Other studies have demonstrated a significant lag in the salination of Black Sea surface waters relative to the time of first reconnection (e.g., Major et al., 2006; Hiscott et al., 2007b; Marret et al., 2009; Flood et al., 2009; Yanchilina et al., 2017). Hiscott et al. (2007b) and Marret et al. (2009) advocated that the first arrival of saline water was as short pulse(s) of Mediterranean inflow facilitated by a temporary weakening of Black Sea outflow and not the beginning of persistent two-way flow across the Strait of Bosphorus.

At the M05-50 site, sediments above the unconformity α_1 provide evidence for the pace of post-glacial Mediterranean inflow into the Black Sea. Based on dinocyst data from core M02-45, Hiscott et al. (2007b) suggested that after the initiation of persistent Mediterranean inflow, and the associated establishment of two-way exchanges across the Strait of Bosphorus, the salinity increased to >10–12. Such salinities would have been tolerable for the brackish-water ostracods in Bioecozone 2. Subsequently, the start of sulfate reduction signifying a good input of normal marine, saline water to the M02-45 site occurred at ~8000 cal yr BP (7500 ^{14}C ka recalibrated from Hiscott et al., 2007b; also Mertens et al., 2012). This implies a lag of at least 500 years between the beginning of persistent marine inflow into the Black Sea at ~8000 cal yr BP and the pioneer colonization of the area by the first Mediterranean ostracod *S. littoralis* at ~7450–7425 cal yr BP (Figs. 16–18). This lag is interpreted as the time needed for enough Mediterranean water to enter the Black Sea to raise the salinity to a level suitable for marine ostracods and to form a breeding population. This is in agreement with Major et al. (2006) who reinterpreted the date of ~7570 cal yr BP which Ryan et al. (1997) had initially thought marked initial Mediterranean inflow to instead mark the time when the salinity became suitable for Mediterranean molluscan fauna to begin colonizing the Black Sea. This lag in the progress of salination cannot be used to discriminate between a catastrophic versus a gradual initial

reconnection, because modelling by Soulet et al. (2010) showed that even a very rapid rise in the level of the Black Sea from \sim -90 m to \sim -40 m due to the addition of \sim 28000 km³ of 38 Mediterranean water would only increase the average salinity of a well mixed Black Sea by \sim 2.

Ostracod species found toward the base of cores M05-50 and M02-45 show that conditions in the Black Sea from \sim 11910 to \sim 7425 cal yr BP were brackish rather than fresh. Apart from the presence of the “Caspi-brackish” assemblage, the notable absence of typical freshwater lacustrine species (that are common and well-known in the European Holocene) is significant. A freshwater Black Sea lake has been postulated by Ryan et al. (1997), Soulet et al. (2010) and Yanchilina et al. (2017), but brackish conditions in the early Holocene Black Sea have been confirmed by a number of authors (Giunta et al., 2007; Mudie et al., 2007; Marret et al., 2009; Mertens et al., 2012).

After the establishment at \sim 8000 cal yr BP of fully developed two-way flow through the Strait of Bosphorus, salinity values reached 13–14 during Bioecozone 3 time, followed by a 14–16.5 climb through Bioecozone 4 time (Mertens et al., 2012). The 13–14 salinity was apparently tolerable for both marine and brackish water ostracods in the transitional Bioecozone 3. The M05-50 site is immediately adjacent to the saline underflow channel which crosses the shelf from the Strait of Bosphorus, so it is probable that the bottom waters after reconnection were more saline than 14, even more conducive to marine ostracods.

The establishment of Bioecozone 4 at \sim 6310–6320 cal yr BP (Figs. 16–18) coincides approximately with the weakening of Black Sea outflow at \sim 6400 cal yr BP proposed by Hiscott et al. (2002), facilitating the entry of more marine water and organisms. The floor of the Strait of Bosphorus would have been continuously bathed with Mediterranean water; juvenile ostracods would have been more effectively transported by stronger northward-flowing bottom waters.

This timing also coincides with the maximum input of organic carbon from marine sources at site M02-45 (Hiscott et al., 2007b) indicating a well-established reconnection.

At the start of Bioecozone 5 time, Mertens et al. (2012) indicate a sharp salinity rise to ~17 which then gradually climbs to ~18 after ~2300 cal yr BP. This is in agreement with the ostracod data from core M05-50 which show that salinity conditions reached near-modern levels in Bioecozone 6 starting at ~2390 cal yr BP, which is ~5000 years after the beginning of the Holocene faunal turnover at ~7425 cal yr BP.

The results of this study closely resemble the findings of Zenina et al. (2017), who studied Holocene environmental changes in the Black Sea using ostracod assemblages in two cores from its northeastern outer shelf. Those authors distinguished three distinctive faunal assemblages. Their assemblage I is characterized by a Caspian-type ostracod fauna with common occurrences of *L. lepida* and *G. elongata* and lesser quantities of *L. sublepida* (Zenina et al., 2017). This assemblage dominated the early Holocene up to ~7.4 cal ka BP. It is very similar both in composition and timing to bioecozones 1 and 2 identified at sites M02-45 and M05-50. The Zenina et al. (2017) assemblage II is characterized by a mixed fauna of Caspian-type species (e.g., *L. lepida*, *L. sublepida* and *G. elongate*) and Mediterranean-type species (e.g., *H. rubra*, *L. multipunctata* and *P. agilis*), co-existing from ~7.4 cal ka BP to 6.7 cal ka BP. Assemblage II is nearly identical in faunal characteristics and age to the transitional bioecozone 3 described in this study. Finally, the Zenina et al. (2017) assemblage III is characterized by the progressive dominance of Mediterranean species (particularly *P. agilis*, *Bythocythere* sp., and minor occurrences of *C. carinata* and *C. diffusa*) from 6.8 cal ka BP to the present.

The results of this study are also similar to the findings of Giunta et al. (2007) who noted three calcareous nannoplankton “ecozones” in sediment cores from the western Black Sea. They

interpreted a gradual increase in salinity in the Black Sea after ~7600 cal yr BP. Based on changes in the calcareous nannoplankton *Emiliana huxleyi* and *Braarudosphaera bigelowii* they interpreted surface-water salinities of less than 11–17 prior to ~7600 cal yr BP and >17 afterward until the onset of near-modern conditions by ~3100 cal yr BP. Giunta et al. (2007) also noted that lithological boundaries always preceded biostratigraphic boundaries “probably because colonization of the new habitat by calcareous nannoplankton is gradual”. We would generalize this statement to include the ostracod species examined in this study, as well as molluscs considered by other workers (e.g., Major et al., 2006).

The ostracod fauna show the presence of a diverse and thriving brackish-water ostracod community dominated by *L. sublepada* and *L. lepida* until ~7425 cal yr BP. Subsequently, the first Mediterranean species *S. littoralis* began to rapidly colonize the southwestern Black Sea shelf north of the Bosphorus Strait. For ~900 years the original Ponto-Caspian ostracod species and the new Mediterranean immigrant species lived at the M05-50 and M02-45 core sites in more-or-less equal abundances. After ~6310 cal yr BP the Mediterranean species began to diversify and dominate the core sites on the southwestern Black Sea shelf. The Ponto-Caspian ostracod species and the Mediterranean species *S. littoralis* were replaced by new Mediterranean ostracod species, the most abundant being *P. agilis*.

Two-way flow between the Black Sea and the Mediterranean Sea was initiated at ~8000 cal yr BP, approximately 1500 years after strontium-isotopic evidence for first contact between the two watermasses. There was a further a lag of ~500 years between the first persistent entry of Mediterranean water into the Black Sea and the discernible colonization of the area by Mediterranean ostracod species. This lag is interpreted as the amount of time necessary for the water at the M05-50 core site to reach salinity levels favorable to ostracods migrating from the

more saline environment of the Mediterranean Sea. Statistical analyses of the ostracod data show six bioecozones with distinct ostracod assemblages; changes in those assemblages indicate a gradual and sequential salination of the Black Sea bottom water as a result of steady inflow of Mediterranean waters. The uppermost Bioecozone 6 contains ostracods which prefer marine salinities, some of which can inhabit depths of 100 m or more, suggesting that sealevel and salinity in the Black Sea were near modern values by ~2300 cal yr BP.

7. Conclusions

Ostracods are used to delineate the paleoceanographic and paleoecological evolution of the southwestern Black Sea in two composite cores (M02-45 and M05-50). Sixteen radiocarbon dates in M02-45 and nine in M05-50 constrain age models extending from ~12900–12000 cal yr BP to the present. A local unconformity known as $\alpha 1$ occurs at 695 cm depth in core M05-50, corresponding to a hiatus of ~2200–2700 years. This level is conformable in M02-45. Another local unconformity known as $\alpha 2$ occurs at 265 cm depth in core M02-45, corresponding to a hiatus of ~3390 years. This level is conformable in M05-50. The following are the salient results of the ostracod data:

- Ostracod valves collected from composite cores M05-50 (M05-50P & M05-51G) and M02-45 (M02-45P/T & M05-03P) reveal that from 11910–7425 cal yr BP the ostracod assemblage was dominated by brackish water Ponto-Caspian species. From 7425–6310 cal yr BP the Mediterranean species co-existed with brackish-water species in more-or-less equal abundances. From 6310 cal yr BP to the present the Ponto-Caspian species were replaced by exclusively Mediterranean species.
- There was a time lag of ~500 years between the initiation of persistent inflow from the Mediterranean Sea at ~8000 cal yr BP (as part of a two-way flow system) and the

colonization of the Black Sea shelf by the first Mediterranean ostracod species *S. littoralis*. This lag is interpreted as the time needed for enough water to enter the Black Sea to mix with the bottom water and raise the salinity to levels favourable to Mediterranean ostracod species.

- CONISS cluster analyses divided the cored succession into six bioecozones. Bioecozone 1 constitutes a lower, brackish assemblage dominated by *L. sublepada*, *L. lepida*, *T. amnicola donetziensis*, *G. elongata*, *A. olivia* and *A. quinquetuberculata*. Based on the modern occurrences of these species, the salinity of the Black Sea from 11910–7425 cal yr BP is estimated to be ~5–10. Bioecozone 2 represents a transitional assemblage with salinities high enough for the Mediterranean species *S. littoralis* (and later others) to rapidly move into the area. *S. littoralis* might indicate that the salinity of the Black Sea bottom water from ~7425–6310 cal yr BP was ~13–15, with an upper limit of ~18 to account for the continued presence of several brackish-water species. Bioecozones 3, 4 and 5 are composed of marine assemblages.

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Figure captions

Figure 1. Location map showing the Black Sea and its connection to the Aegean Sea via the Marmara Sea Gateway. M02-45/M05-03 and M05-50/51 locate cores M02-45T/P, M05-03P and M05-50P, M05-51G, respectively (described in text). Distances between individual cores and their penetrations are shown in Figs. 3 and 4. The last glacial maximum (LGM) shoreline is drawn at the ~125 m isobath (IOC, 1981). At that time, the water body labelled as the Black Sea was instead the Neoeuxine Lake. Topography and bathymetry compiled using GeoMapApp (Ryan et al., 2009) in Global Mapper. Coastline and rivers are from NOAA National Geophysical Data Center (<http://www.ngdc.noaa.gov/mgg/shorelines/shorelines.html>).

Figure 2. Sun-illuminated multibeam bathymetry of the southwestern Black Sea immediately north of the Strait of Bosphorus, showing the prominent saline channel (created using Caris Base Editor 4.1) and the locations of the cores collected during the MAR05 cruise. Cores M05-50P and M05-51G are described in text. The multibeam data were collected during the M05, M08 and M11 cruises of the R/V Koca Piri Reis of the Dokuz Eylül University, and are fully described and interpreted in Flood et al. (2009) and Hiscott et al. (2013). Coastline is digitized in Google Earth. Isobaths are created in Global Mapper. The 50 and 100 m isobaths outside the detailed multibeam area are from the International Bathymetric Charts of the Mediterranean (IOC, 1981).

Figure 3. Huntect DTS boomer profile collected during the MAR2005 cruise showing the acoustic character of the Holocene sedimentary successions across the southwestern Black Sea and the along-strike projections of core sites M02-45P/T and M05-03P to this line. Vertical resolution is ~25 cm. Reflectors α and α_2 are unconformities described

and interpreted in Aksu et al. (2002) and overlapped by younger strata. Reflector α_1 appears to show mild local truncation of underlying reflectors so might be a disconformity around the core sites, although no evidence for this is present in the age model or visual descriptions. Elsewhere across the southwestern Black Sea shelf, α_1 is interpreted by Hiscott et al. (2007b) and Flood et al. (2009) as a subaqueous erosion surface. Considering projection distances, the extent of basal onlap between the M02-45P/T and M05-03P sites cannot be estimated from this graphic. Water depth and sediment thicknesses are calculated using an acoustic velocity of 1500 m s^{-1} . Vertical exaggeration $VE = 27x$; f 926.5–f 927.5 are fix numbers assigned during the survey. Location is shown in the inset.

Figure 4. Hunttec DTS boomer profile collected during the MAR2005 cruise showing the acoustic character of the Holocene sedimentary successions across the southwestern Black Sea and the along-strike projections of core sites MAR05-50P and MAR05-51G to this line. This profile is part of a 200 line-km grid of Hunttec data around the core sites. Vertical resolution is $\sim 25 \text{ cm}$. Reflectors α , α_1 and α_2 are unconformities (the latter only locally in this area described and interpreted in Aksu et al. (2002), Hiscott et al. (2007b) and Flood et al. (2009); all are overlapped by younger strata. Water depth and sediment thicknesses are calculated using an acoustic velocity of 1500 m s^{-1} . Vertical exaggeration $VE = 27x$; f 1063–f 1064 are fix numbers assigned during the survey. Location is shown in the inset.

Figure 5. Paired downcore plots of total organic carbon (TOC), total sulphur (TS) and $\delta^{34}\text{S}$ for (a) gravity and piston cores M02-45T and M02-45P (correlated as justified by Hiscott et al., 2007b, with 110 cm core-top loss in M02-45P) and (b) piston core M05-03P. M05-

03P apparently had its trip wire set incorrectly during coring so that core-top loss is 302 cm. For analytical methods, see Hiscott et al. (2007b). Several key peaks, troughs, and abrupt steps in the plots lead to an unambiguous correlation, and to the conclusion that M05-03P extends into older sediment than M02-45P. Its lowermost facies also are consistent with intersection of gravelly lag deposits along the α unconformity. This facies is absent in M02-45P.

Figure 6. Simplified lithostratigraphy of the composite cores M02-45 (**left**) and M05-50 (**right**) showing the positions of the lithological units (discussed in text) and the radiocarbon dates (shown as calendar years before present, cal yr BP; see Table 1 for raw ^{14}C ages). In the case of M02-45, the splice with core M05-03P (adding 78 cm to the base of the composite) used three geochemical markers found in both piston cores at that site, and marked in Fig. 5. The age versus depth plots show the 2305 years hiatus at the $\alpha 1$ unconformity in core M05-50 and the 3025 years hiatus at the $\alpha 2$ unconformity in core M02-45. Note that the $\alpha 2$ reflector which is a local unconformity at core site M02-45 is apparently a conformable surface at core site M05-50. Conversely, the $\alpha 1$ reflector which is a local unconformity at core site M05-50 is apparently a conformable surface at core site M02-45. Location is shown in Figure 1.

Figure 7. Summary of interpreted reservoir ages for shallow waters of the latest Quaternary Black Sea. Yanchilina et al. (2017) proposed but did not explain the reason for an abrupt rise in R several centuries before reconnection to the world ocean. Dashed lines and question marks denote uncertainty noted in the original articles. The default Marine13 modelled reservoir age from $\sim 450\text{--}7100$ ^{14}C yr BP was calculated by determining the difference between the Marine13 and IntCal13 defining curves (Reimer et al., 2013), the

latter smoothed with a 9-point moving average to mimic the natural smoothing of the marine record (Stuiver and Braziunas, 1993). In this paper, calibrations of mollusc ages at the M02-45 core site use R values picked from the blue trend.

Figure 8. $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ trends for stalagmites from the Sofular Cave, Turkey, presented as raw data (thin blue trends) and 5-point moving averages (bold gray trends) plotted against atmospheric ^{14}C ages converted from original U/Th dates on stalagmite samples using the IntCal13 data set. Superimposed are scatterplots of $\delta^{13}\text{C}$ and $\delta^{18}\text{O}$ data from Black Sea molluscs (Yanchilina et al., 2017, their supplementary file mmc6.xlsx). Red scales relate to red data points for molluscs, and black scales relate to Sofular Cave trendlines. Parts a and b plot the mollusc results using uncorrected ^{14}C dates; parts c and d incorporate reservoir corrections proposed by Yanchilina et al. (2017; consistent with their curve in Fig. 7); parts e and f use reservoir ages preferred by the authors (labelled as "this paper R " in Fig. 7). Reservoir age is an intrinsic property of the dissolved inorganic carbon (DIC) in the seawater of the day, so must be applied identically to each pair of isotopic plots. BA = Bølling/Allerød 'trough' in $\delta^{13}\text{C}$ that effectively dictates the choice of appropriate reservoir ages before ~ 11000 ^{14}C yr BP. Circled 1 = cluster of $\delta^{13}\text{C}$ data points (2–3‰) which cannot be adjusted to fit the slightly older peak in the Sofular Cave trend because reservoir offsets can only shift marine radiocarbon ages downward. Circled 2 = swath of time (~ 8200 – 10000 ^{14}C yr BP) devoid of mollusc results when ages are corrected using Yanchilina et al. (2017) parameters, because high R values squeeze raw ^{14}C dates in this interval to younger ages (compare parts a and c). Circled 3 = cluster of $\delta^{18}\text{O}$ data points conspicuously shifted away from the Sofular Cave trend by Yanchilina et al. (2017) procedures, but not by author calibrations (compare parts d and

f). Circled 4 = cluster of $\delta^{18}\text{O}$ data points which poorly fit the Sofular Cave trend, but which cannot be moved by increasing the reservoir age because this would destroy the fine match between $\delta^{13}\text{C}$ mollusc data and the Bølling/Allerød "trough", which cannot be abandoned because it is the most compelling match in the $\delta^{13}\text{C}$ plots.

Figure 9. Plot of uncalibrated radiocarbon age versus the depositional elevation (relative to modern sea level = rmsl) of 123 dated shells extracted from Memorial University of Newfoundland cores raised from the modern southwestern Black Sea shelf. The authors agree with Yanchilina et al. (2017) that the shelf was fully flooded to an elevation of -40 m rmsl (or above) since ~ 8750 ^{14}C yr BP; the senior authors have proposed (Aksu et al., 2016) that this elevation was reached and maintained earlier, by ~ 11.1 cal ka (~ 10.1 ^{14}C ka). Even the younger estimate of Yanchilina et al. (2017) requires that both the $\alpha 1$ and $\alpha 2$ levels at site M02-45 developed under tens of meters of water as submarine conformities/unconformities because sediments beneath these horizons are younger than ~ 7.0 ^{14}C ka and ~ 5.1 ^{14}C ka, respectively (annotated arrows to right side). Twenty dated shells (white symbols) come from modern depths which Yanchilina et al. (2017) propose to have been subaerial; instead, these *D. polymorpha*, *D. rostriformis*, *Theodoxus* spp. and *Turricaspia spica* shells confirm subaqueous conditions on the earliest Holocene southwestern shelf. The lack of data from water depths less than about -60 m attests to the steep inclination of the coastal profile in this area so that most of the shelf is deeper than ~ -50 m (Aksu et al., 2002, their figure 2).

Figure 10. Number of ostracod valves counted in composite cores M02-45 and M05-50. All valves were picked in each sample, so for samples of approximately equal volume, number of counts provides an indication of the richness of the samples. Notice that in

M02-45, off-scale peaks are located very close to unconformities (α , α_2) or intervals of slow accumulation (α_1 , Fig. 5).

Figure 11. Scanning electron microscope images of key Ponto-Caspian ostracod species. (1) *Amniccythere cymbula* (left valve, LV), M05-50 690 cm (2) *Amniccythere olivia* (LV) M05-50 580 cm (3) *Amniccythere quinquetuberculata* (LV), M05-50 770 cm (4) *Amniccythere* sp. aff. *subcaspia* (LV), M05-50 740 cm (5) *Graviacypris elongate* (right valve, RV), M05-50 640 cm (6) *Euxinocythere bacuana* (LV), M05-50 680 cm (7) *Tyrrhenocythere amnicola donetziensis* (RV), M05-50 780 cm (8) *Loxoconchissa* (*Loxocaspia*) *immodulata* (LV), M05-50 660 cm (9) *Loxoconcha lepida* (female, LV), M05-50 720 cm (10) *Loxoconcha lepida* (male, RV), M05-50 720 cm (11) *Loxoconcha sublepida* (LV), M05-50 720 cm (12) *Euxinocythere* sp. aff. *relicta* (RV), M05-50 760 cm. Scale bars = 100 μ m.

Figure 12. Scanning electron microscope images of key Mediterranean ostracod species. (1) *Amniccythere propinqua* (LV), M05-50 120 cm (2) *Callistocythere diffusa* (LV), M05-50 120 cm (3) *Carinocythereis carinata* (female, LV), M05-50 350 cm (4) *Carinocythereis carinata* (male, LV), M05-50 350 cm (5) *Costa edwardsi* (LV), M05-50 50 cm (6) *Cytheroma variabilis* (LV), M05-50 120 cm (7) *Hiltermannicythere rubra* (LV), M05-50 330 cm (8) *Leptocythere multipunctata* (LV), M05-50 350 cm (9) *Sagmatocythere littoralis* (LV), M05-50 580 cm (10) *Pterygocytheris jonesii* (LV) M05-50 370 cm (11) *Palmoconcha agilis* (female, LV), M05-50 430 cm (12) *Palmoconcha agilis* (male, LV), M05-50 430 cm (13) *Xestoleberis* sp. aff. *cornelii* (RV), M05-50 390 cm, (14), *Paracypris polita* (RV), M05-50 20 cm. Scale bars = 100 μ m.

Figure 13. Population age diagrams for key species from irregularly spaced depths throughout composite core MAR05-50. The presence of adults (A) and various juveniles (A-1 through A-6) confirms an autochthonous assemblage on which an environmental interpretation can be based. Gray and green bars indicate brackish and marine species, respectively.

Figure 14. Downcore variations of key Mediterranean (blue line–green fill) and Ponto-Caspian (red line–gray fill) ostracod species in composite core MAR05-50, shown in percent abundances of total fauna in depth domain. Lithostratigraphic Units 1–3 and $\alpha 1$ surface are explained in text. Core location is shown in Figures 1 and 2.

Figure 15. Downcore variations of key Mediterranean (blue line–green fill) and Ponto-Caspian (red line–gray fill) ostracod species in composite MAR02-45, shown in percent abundances of total fauna in depth domain (partial data from Evans, 2004). Lithostratigraphic Units A–D, α and $\alpha 2$ unconformities, and the $\alpha 1$ surface are explained in text. Core location is shown in Figures 1 and 3.

Figure 16. CONISS cluster analysis results show that core MAR05-50 can be divided into six bioecozones (1–6) based on distinct assemblages of key ostracod species, reflecting step-by-step ecological changes that took place in the Black Sea driven by gradual salination following the post-glacial reconnection to the Mediterranean Sea. Hiatus at $\alpha 1$ is described in text.

Figure 17. CONISS cluster analysis results show that core MAR02-45 can be divided into six bioecozones (1–6) based on distinct assemblages of key ostracod species, reflecting step-by-step ecological changes that took place in the Black Sea driven by gradual salination

following the post-glacial reconnection to the Mediterranean Sea. Hiatus at $\alpha 2$ is described in text.

Figure 18. Synthesis of CONISS results and the associated Bioecozone across the southwestern Black Sea shelf. Purple lines with italicized purple numbers are bioecozone boundaries and their ages in cal yr BP. Pink highlight depicts the estimated lateral extents of the hiatuses at $\alpha 1$ and $\alpha 2$ (described in text). Salinity estimates in brackets for each bioecozone are explained in text. Downcore plots of the total Mediterranean versus total Ponto-Caspian assemblages (at far right and far left) show the co-occurrence of Mediterranean and Ponto-Caspian species associated with the dramatic faunal turnover during Bioecozone 3.

Table 1. Radiocarbon ages for cores M02-45T, M02-45P, M05-03P, M05-50P and M05-51G reported as conventional ^{14}C dates (half-life of 5568 years; errors represent 68.3% confidence limits), calibrated to calendar years using the Marine13 data set and reservoir ages from Fig. 7. Bold dates are not used in the age models for this paper. Calibrations of raw dates younger than 7.0 ^{14}C ka are effectively identical to those of Mertens et al. (2012). TO = IsoTrace Radiocarbon Laboratory, Accelerator Mass Spectrometry Facility, University of Toronto. UC = UCIAMS = Radiocarbon Dating Laboratory, Université Laval, in collaboration with KECK Carbon Cycle AMS Facility, University of California, Irvine. BETA = BETA Analytic, Miami. T = trigger-weight core; P = piston core.

Core	Depth (cm)	Composite depth (cm)	Material dated	Raw date (^{14}C yr BP)	ΔR (^{14}C yr)	Calendar Age (cal yr BP)	Lab number
M02-45T	92	92	<i>Spisula subtruncata</i>	730±50	+10	365±50	TO-11433
M02-45P	33	143	<i>Spisula subtruncata</i>	730±40	+10	365±45	TO-11435
M02-45T	145	145	<i>Spisula subtruncata</i>	770±50	+10	395±55	TO-11434
M02-45P	158	268	<i>Mytilus galloprovincialis</i>	2400±60	+10	2025±80	TO-11006
M02-45P	160	270	downward extrapolation			2055±~85	$\alpha 2$ level
M02-45P	161	271	upward extrapolation			5445±~55	$\alpha 2$ level
M02-45P	174	284	<i>Mytilus galloprovincialis</i>	5115±20	+10	5480±40	UC-85907
M02-45P	220	330	<i>Mytilus galloprovincialis</i>	5190±50	+10	5535±55	TO-11436
M02	302	412	<i>Mytilus</i>	5900±60	+10	6310±65	TO-11437

-45P			<i>galloprovincialis</i>				
M05 -03P	183	485	<i>Cardium edule</i>	6810±25	+10	7330±35	UC-85908
M02 -45P	406	516	<i>Monodacna pontica</i>	7560±60	-12	8030±70	TO-11438
M05 -03P	226	528	<i>Adacna</i> <i>?laeviuscula</i>	7785±25	-42	8305±45	UC-85911
M02 -45P	495	605	<i>Truncatella</i> <i>subcylindrica</i>	8380±70	- 145	9150±100	TO-11142
M05 -03P	342	644	<i>Monodacna caspia</i>	8340±25	- 135	9070±55	UC-85910
M02 -45P	569	679	<i>Didacna</i> <i>?praetrigonides</i>	8570±70	- 141	9380±80	TO-11439
M02 -45P	639	749	<i>Didacna</i> spp.	8620±70	- 143	9440±70	TO-11440
M02 -45P	754	864	<i>Dreissena</i> <i>rostriformis</i>	8840±70	- 184	9700±120	TO-11441
M02 -45P	810	920	<i>Dreissena</i> <i>rostriformis</i>	9370±70	- 234	10480±100	TO-11007
M02 -45P	822	932	<i>Dreissena</i> <i>polymorpha</i>	9340±70	- 238	10450±100	TO-11442
M02 -45P	835	945	<i>Theodoxus</i> spp.	9070±70	- 127	10010±115	TO-11443
M05 -03P	678	980	<i>Dreissena</i> spp.	9510±25	- 191	10600±45	UC-85909
M05 -03P	712	1014	<i>Dreissena</i> spp.	10475±3 0	- 306	12240±120	UC-85912
M05 -51G	145	145	<i>Mytilus</i> <i>galloprovincialis</i>	3280±60	+10	3095±95	TO 13101
M05	44	94	<i>Mytilus</i>	2590±90	+10	2255±120	TO 13095

-50P			<i>galloprovincialis</i>				
M05	180	230	<i>Mytilus</i>	3240±50	+10	3045±75	TO 13096
-50P			<i>galloprovincialis</i>				
M05	200	250	<i>Mytilus</i>	3590±15	+10	3470±35	UCIAMS-
-50P			<i>galloprovincialis</i>				96128
M05	279	329	<i>Mytilus</i>	3250±70	+10	3055±105	TO 13097
-50P			<i>galloprovincialis</i>				
M05	310	360	<i>Mytilus</i>	4130±20	+10	4175±45	UCIAMS-
-50P			<i>galloprovincialis</i>				96127
M05	340	390	<i>Mytilus</i>	4320±60	+10	4435±90	TO 13098
-50P			<i>galloprovincialis</i>				
M05	435	485	<i>Mytilus</i>	5330±70	+10	5695±80	TO 13099
-50P			<i>galloprovincialis</i>				
M05	550	600	foraminifera/ostraco	7710±40	-50	8235±55	BETA30592
-50P			d				0
M05	620	670	foraminifera/ostraco	8540±50	-	9320±70	BETA30592
-50P			d		110		1
M05	625	675	foraminifera/ostraco	7570±40	-12	8040±60	BETA30798
-50P			d				1
M05	645	695	downward			8285±~60	α1 level
-50P			extrapolation				
M05	646	696	upward			10755±~10	α1 level
-50P			extrapolation			0	
M05	670	720	<i>Dreissena</i>	9880±11	-	11095±135	TO 13100
-50P			<i>polymorpha</i>	0	231		
M05	737	787	<i>Dreissena</i> spp.		-	12010±205	TO 12915
-50P				10270±9	385		
				0			

Table 2. List of most common ostracod species found in the SW Black Sea cores.

Genus and species	Original Author	Figure
<i>Amnicythere cymbula</i>	(LIVENTAL, 1929)	Fig.11, #1
<i>Amnicythere olivia</i>	(LIVENTAL, 1938)	Fig.11, #2
<i>Amnicythere propinqua</i>	(LIVENTAL, 1929)	Fig.12, #1
<i>Amnicythere quinetuberculata</i>	(SCHWEYER, 1949)	Fig.11, #3
<i>Amnicythere</i> sp. aff. <i>subcaspia</i>	(LIVENTAL, 1929)	Fig.11, #4
<i>Amnicythere striatocostata</i>	(SCHWEYER, 1949)	Not shown
<i>Bythocythere</i> sp.	SARS, 1866	Not shown
<i>Callistocythere diffusa</i>	(MÜLLER, 1894)	Fig.12, #2
<i>Carinocythereis carinata</i>	(ROEMER, 1838)	Fig.12, #3, 4
<i>Costa edwardsi</i>	(ROEMER, 1838)	Fig.12, #5
<i>Cytheroma variabilis</i>	MÜLLER, 1894	Fig.12, #6
<i>Euxinocythere bacuana</i>	(LIVENTAL, 1938)	Fig.11, #6
<i>Euxinocythere</i> sp. aff. <i>relicta</i>	(SCHWEYER, 1949)	Fig.11, #12
<i>Graviacypris elongata</i>	(SCHORNIKOV, 1964)	Fig.11, #5
<i>Hiltermannicythere rubra</i>	(MÜLLER, 1894)	Fig.12, #7
<i>Leptocythere multipunctata</i>	(SEGUENZA, 1884)	Fig.12, #8
<i>Loxoconchissa (Loxocaspia) immodulata</i>	STEPANAITYS, 1962	Fig.11, #8
<i>Loxoconcha lepida</i>	STEPANAITYS, 1962	Fig.11, #9, 10
<i>Loxoconcha sublepida</i>	STANCHEVA, 1989	Fig.11, #11
<i>Palmoconcha agilis</i>	(RUGGIERI, 1967)	Fig.12, #11, 12
<i>Paracypris polita</i>	SARS, 1866	Fig.12, #14
<i>Pterygocythereis jonesii</i>	(BAIRD, 1850)	Fig.12, #10
<i>Sagmatocythere littoralis</i>	(MÜLLER, 1894)	Fig.12, #9
<i>Tyrrhenocythere amnicola donetziensis</i>	(DUBOWSKY, 1939)	Fig.11, #7
<i>Xestoleberis</i> sp. aff. <i>cornelii</i>	CARAION, 1963	Fig.12, #13

Table 3. Predominant ostracod species in Bioecozones 1–6 delineated by CONISS in composite cores M02-45 and M05-50, and their ecological significance for the southwestern Black Sea.

Species with greatest abundance are set in bold.

Bioecozones	Core M02-45 composite	Core M05-50 composite	Ecological significance
Bioecozone 1	<i>L. sublepada</i> , <i>L. lepida</i> , <i>T. amnicola</i> <i>donetziensis</i> , <i>A. quinquetuberculata</i> , <i>A. olivia</i> , <i>G. elongata</i>	<i>L. sublepada</i> , <i>L. lepida</i> , <i>T. amnicola</i> <i>donetziensis</i> , <i>A. quinquetuberculata</i> , <i>A. olivia</i> , <i>G. elongata</i>	Low salinity, possibly 5–8 psu but not fresh water
Bioecozone 2	<i>L. sublepada</i> , <i>L. lepida</i> , <i>A. quinquetuberculata</i> , <i>A. olivia</i> , <i>G. elongata</i> , <i>A. bacuana</i>	<i>L. sublepada</i> , <i>L. lepida</i> , <i>T. amnicola</i> <i>donetziensis</i> , <i>A. quinquetuberculata</i> , <i>A. olivia</i> , <i>G. elongata</i>	Brackish, possibly slight salinity increase, possible inflow of Mediterranean water
Bioecozone 3	<i>Cytheroma</i> , <i>C. variabilis</i> , <i>L. lepida</i> , <i>L. sublepada</i> , <i>A. bacuana</i> , <i>A. olivia</i> , <i>G. elongata</i>	<i>S. littoralis</i> , <i>L. sublepada</i> , <i>L. lepida</i> , <i>T. amnicola</i> <i>donetziensis</i> , <i>A. quinquetuberculata</i> , <i>A. olivia</i> , <i>G. elongata</i>	Post-reconnection transitional phase, rising salinity suitable for co-occurrence of brackish and marine ostracods, possibly up to 18 psu
Bioecozone 4	<i>P. agilis</i> , <i>H. rubra</i> , <i>L. multipunctata</i> , <i>X. sp. aff. cornelli</i>	<i>P. agilis</i> , <i>C. carinata</i> , <i>H. rubra</i> , <i>L. multipunctata</i> , <i>C. variabilis</i>	Salinity increased allowing new Mediterranean species to dominate – salinity 18–25 psu
Bioecozone 5	<i>P. agilis</i> , <i>C. carinata</i> ,	<i>P. agilis</i> , <i>C. carinata</i> ,	Introduction of new

	<i>H. rubra</i> , <i>L. multipunctata</i> , <i>C. variabilis</i> , <i>Leptocythere</i> , <i>Cytheroma</i>	<i>H. rubra</i> , <i>L. multipunctata</i> , <i>C. variabilis</i> , <i>P. jonesii</i> , <i>C. edwardsi</i>	Mediterranean sublittoral species, possible sealevel rise, salinity perhaps ≥ 25 psu
Bioecozone 6	<i>P. agilis</i> , <i>C. carinata</i> , <i>H. rubra</i> , <i>L. multipunctata</i> , <i>C. variabilis</i> , <i>Leptocythere</i> , <i>Cytheroma</i>	<i>P. agilis</i> , <i>C. carinata</i> , <i>H. rubra</i> , <i>P. jonesii</i> , <i>C. edwardsi</i> , <i>P. polita</i>	New Mediterranean species, salinity similar to today

Highlights

- Ostracods in two cores from SW Black Sea show a slow, time-lagged salination
- Between 12000–7425 cal yr BP ostracods are dominated by Ponto-Caspian species
- From 7425–6315 cal yr BP ostracods consist of Mediterranean and Ponto-Caspian species
- After ~6315 cal yr BP the assemblage consists exclusively of Mediterranean species

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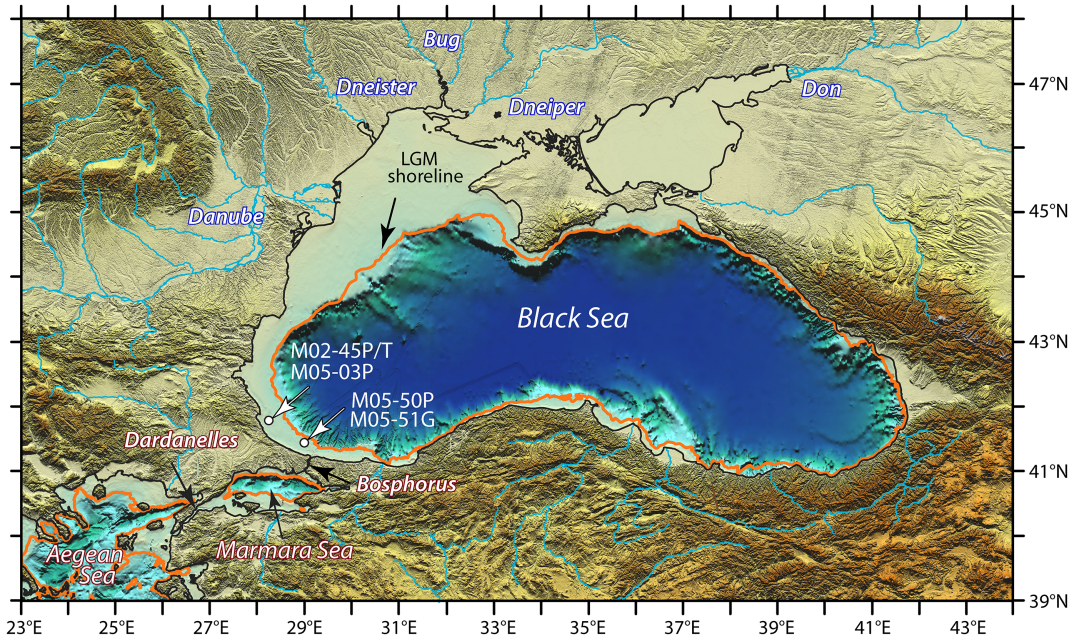


Figure 1

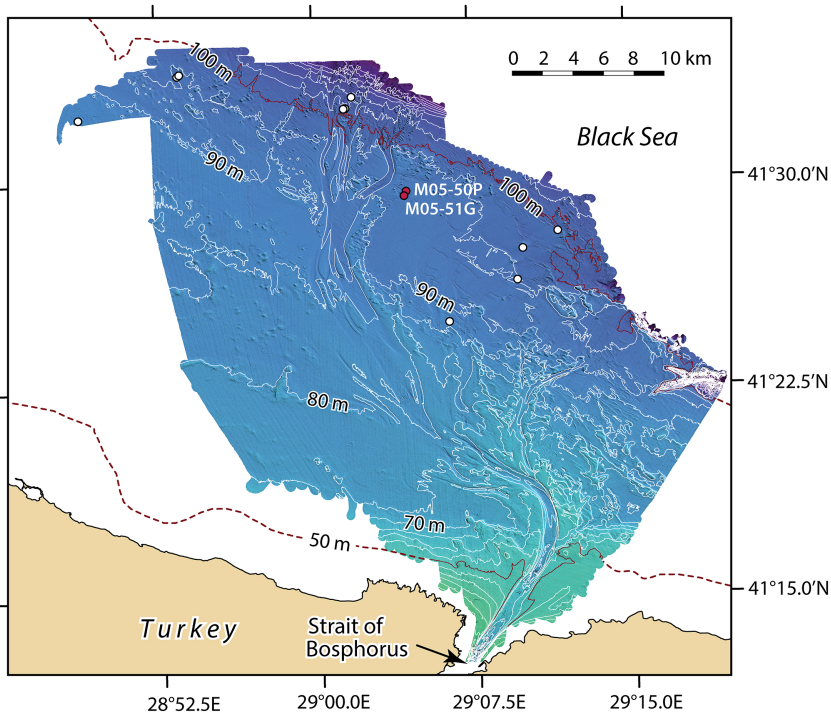


Figure 2

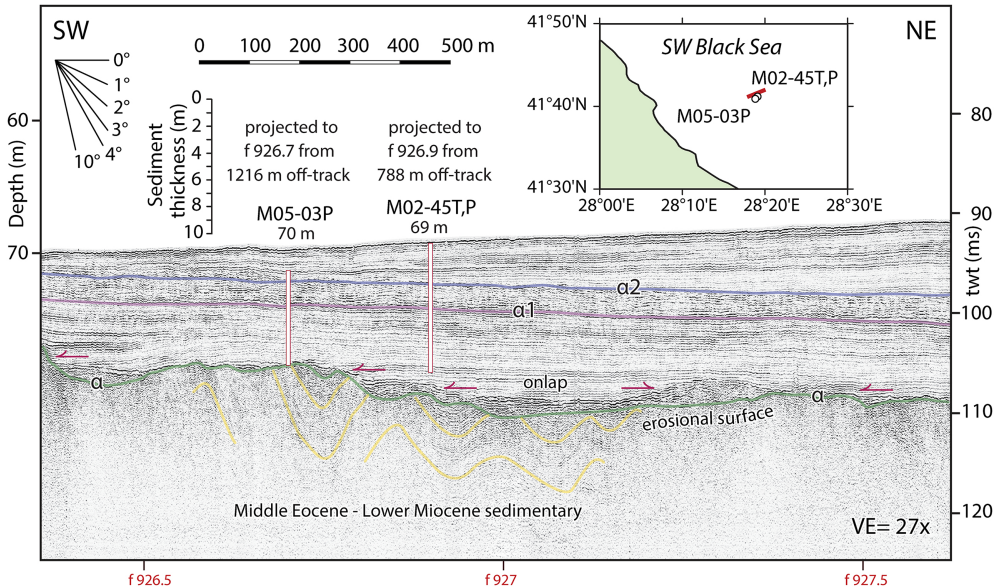


Figure 3

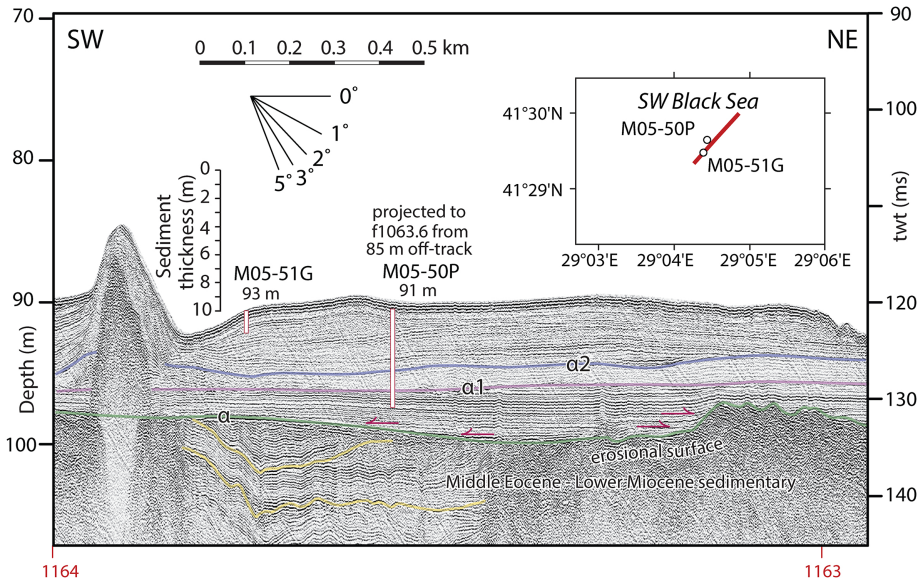


Figure 4

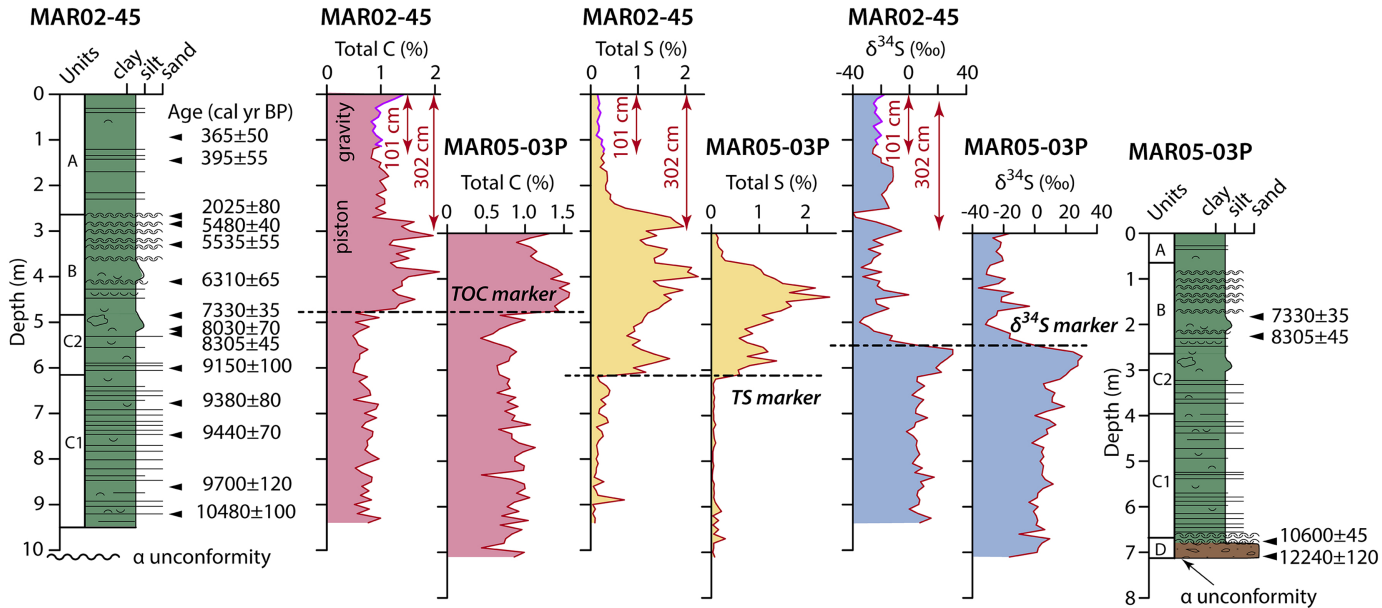


Figure 5

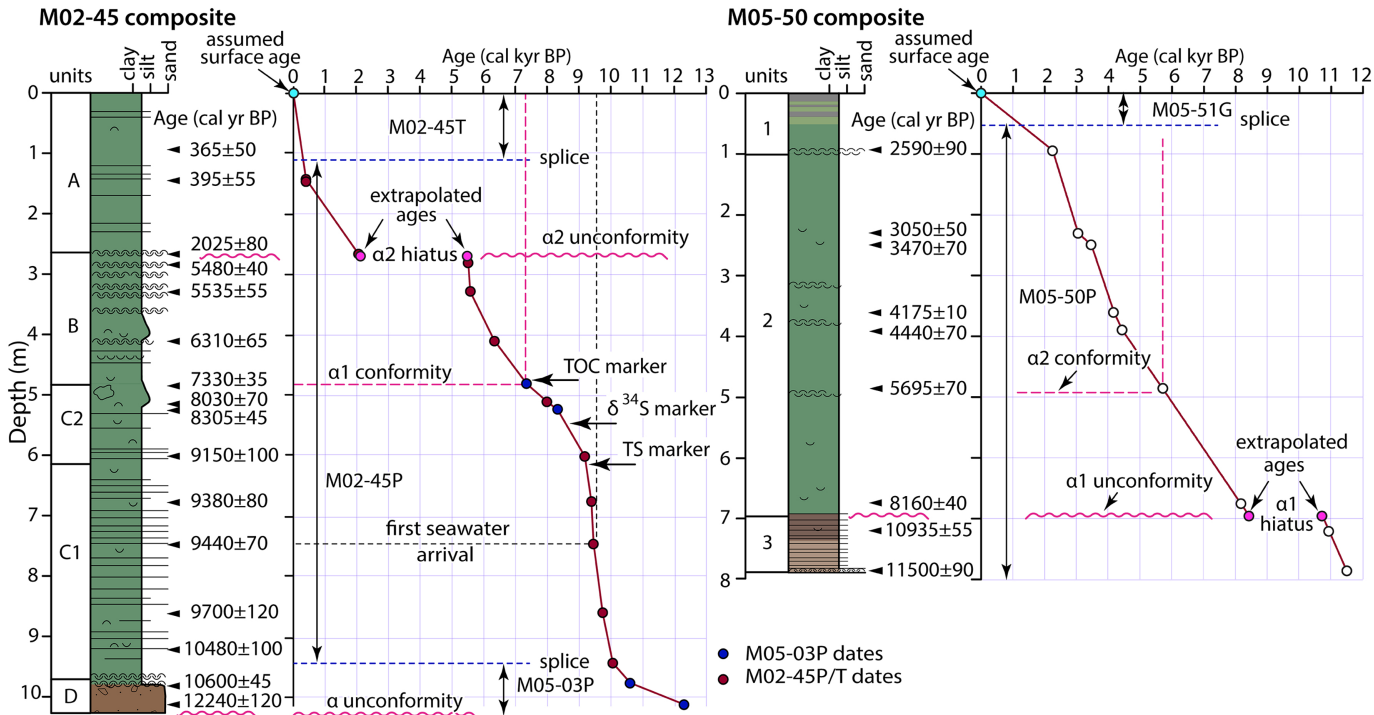


Figure 6

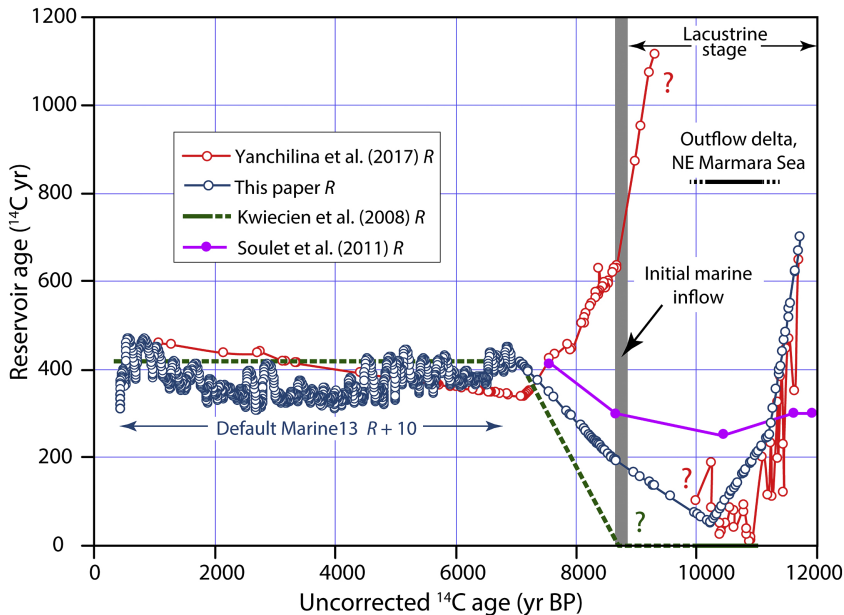


Figure 7

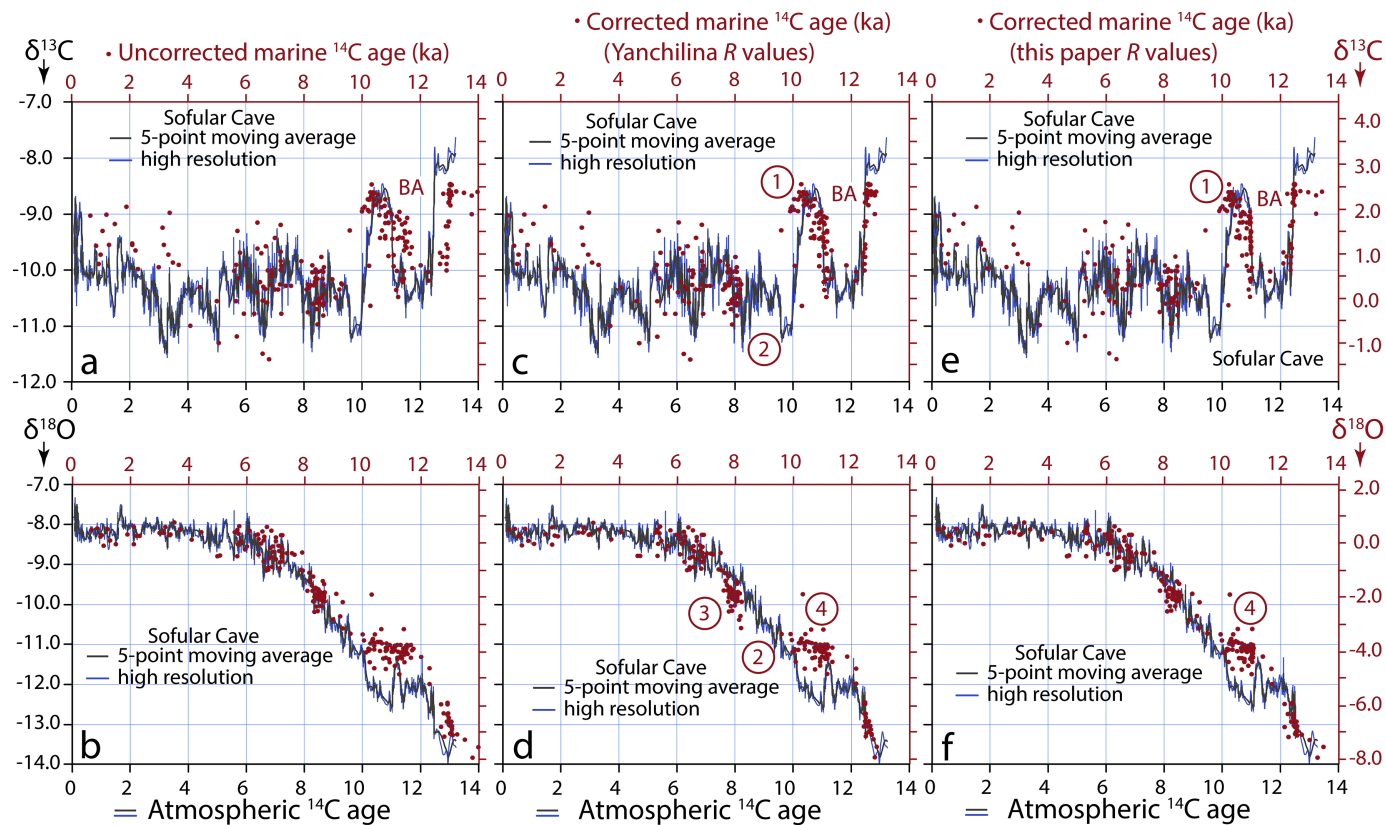


Figure 8

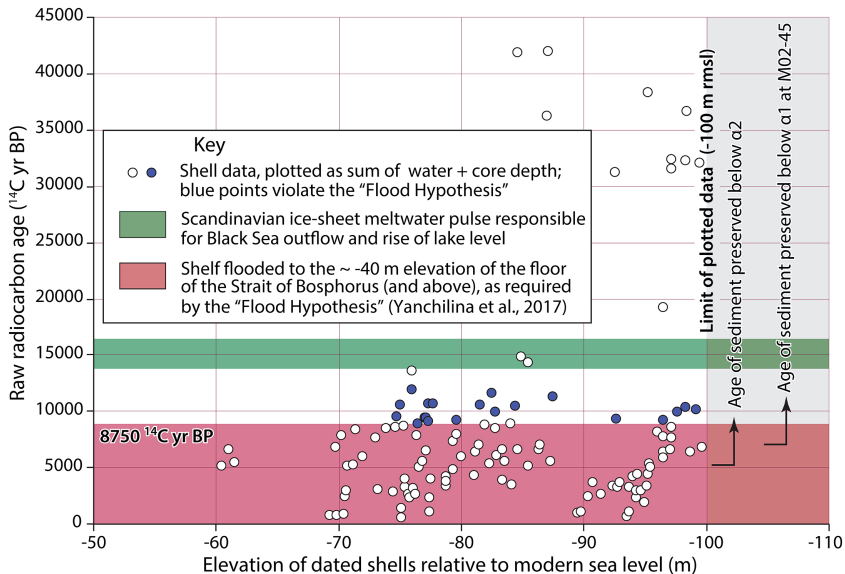


Figure 9

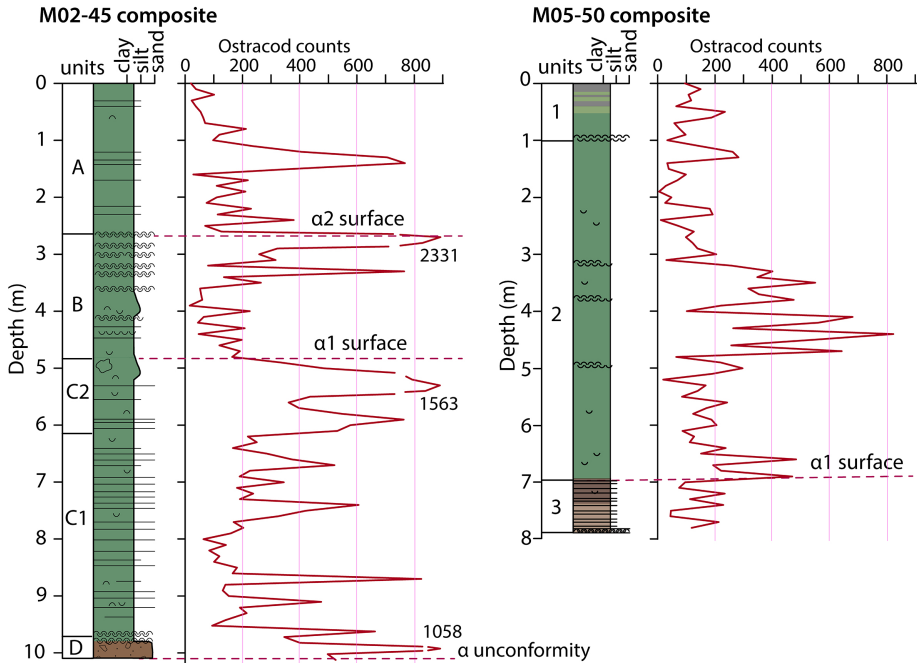


Figure 10

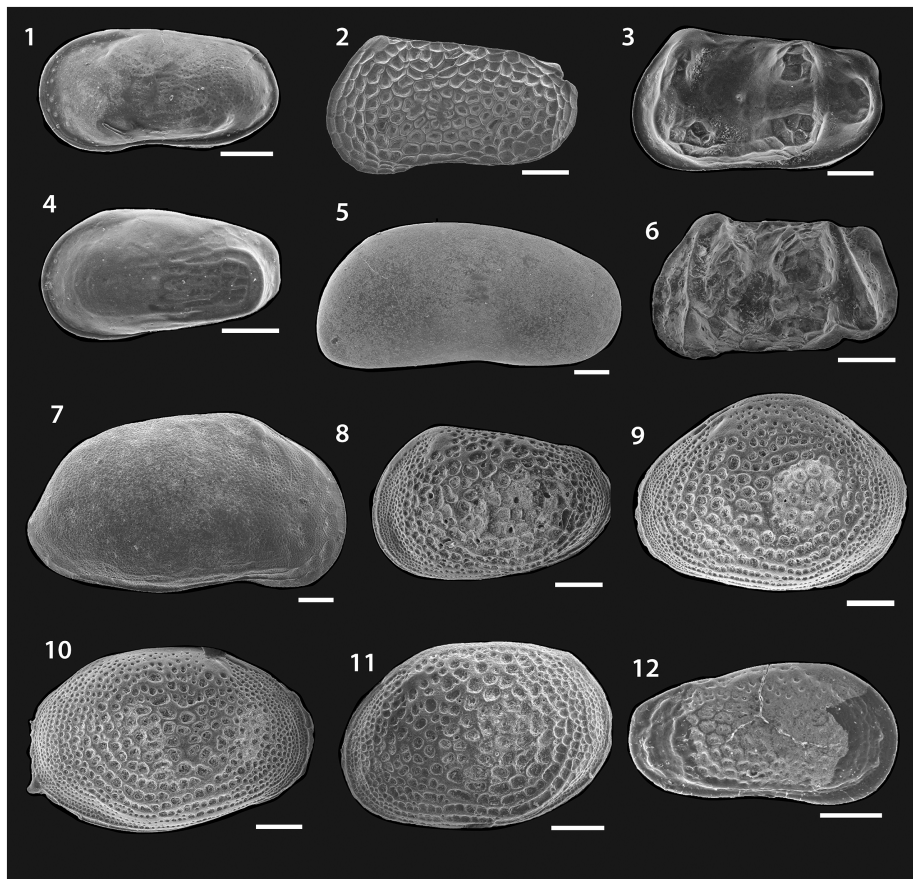


Figure 11

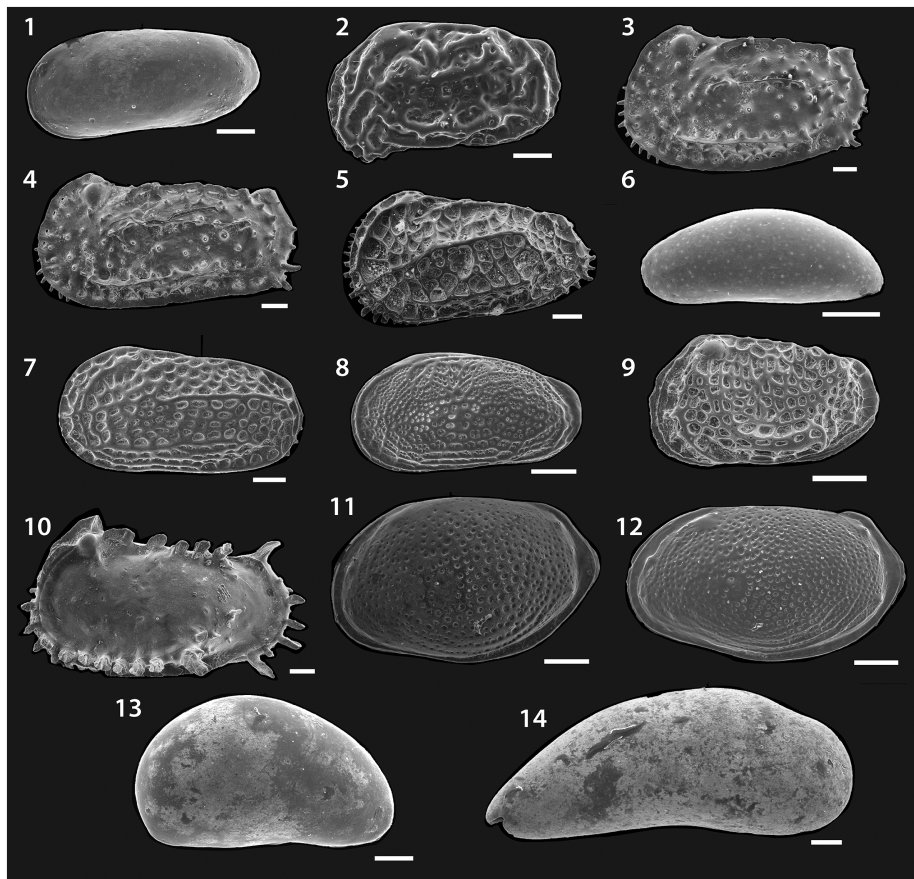


Figure 12

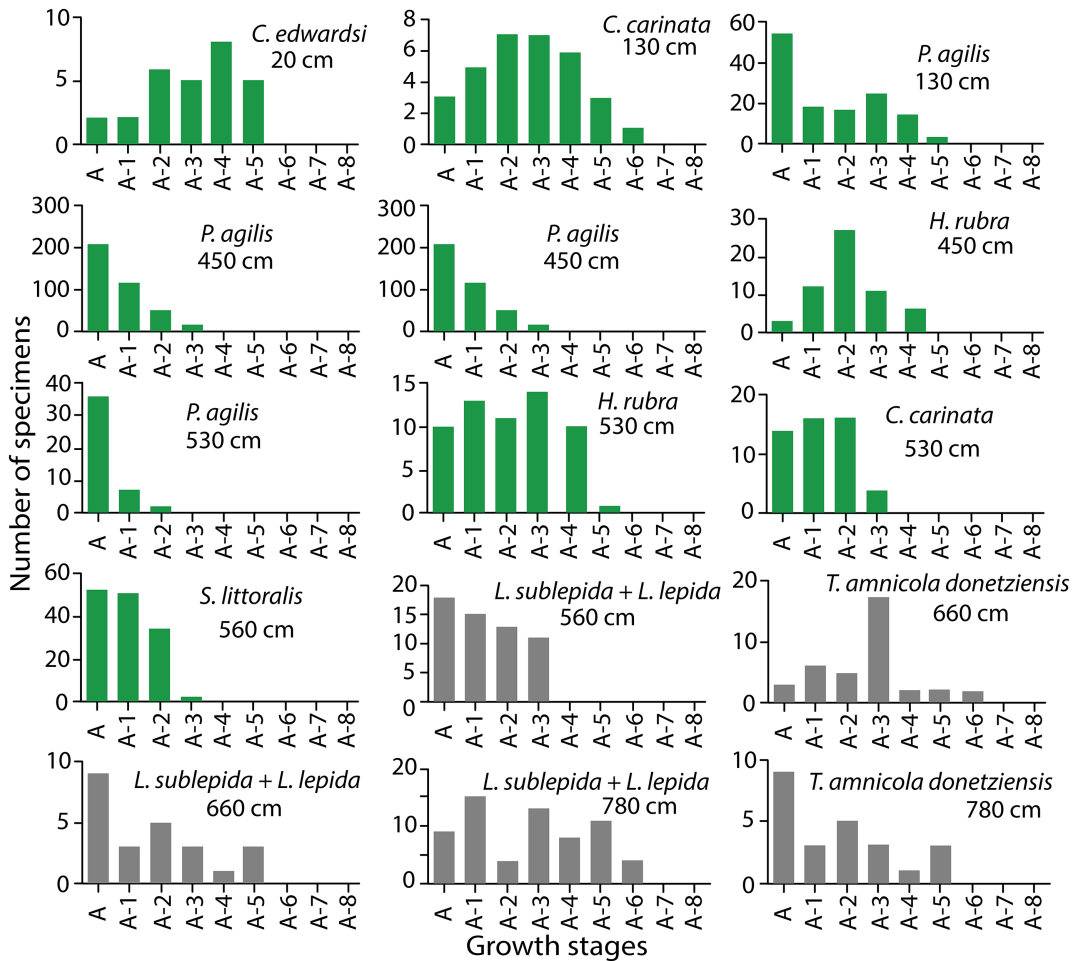


Figure 13

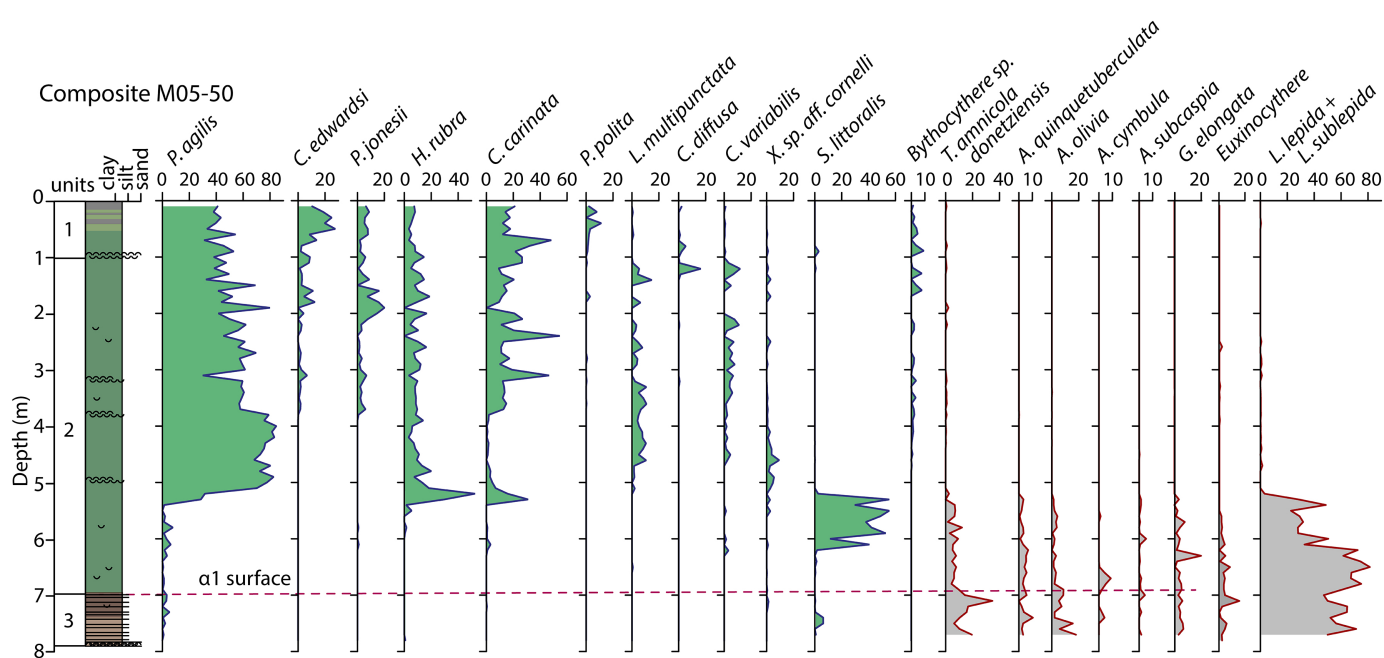


Figure 14

Composite M02-45

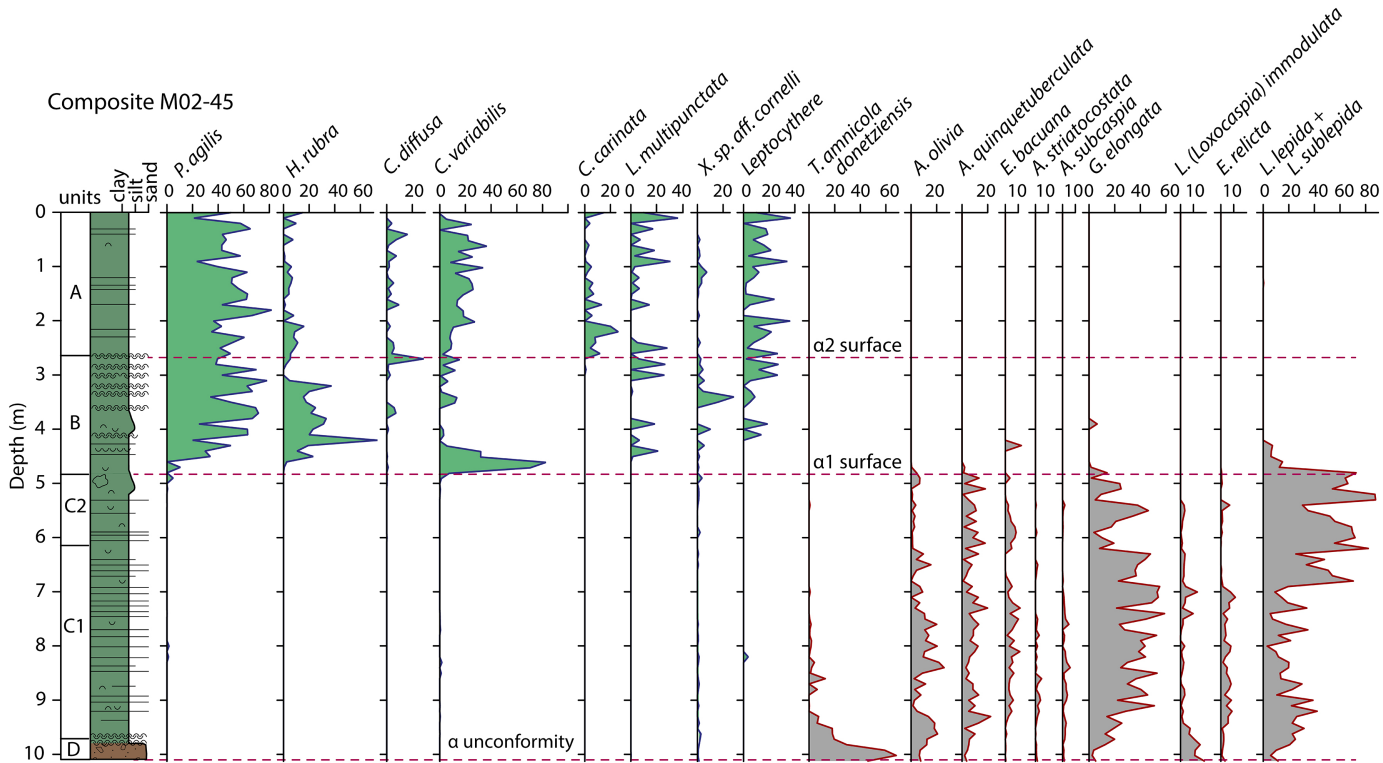


Figure 15

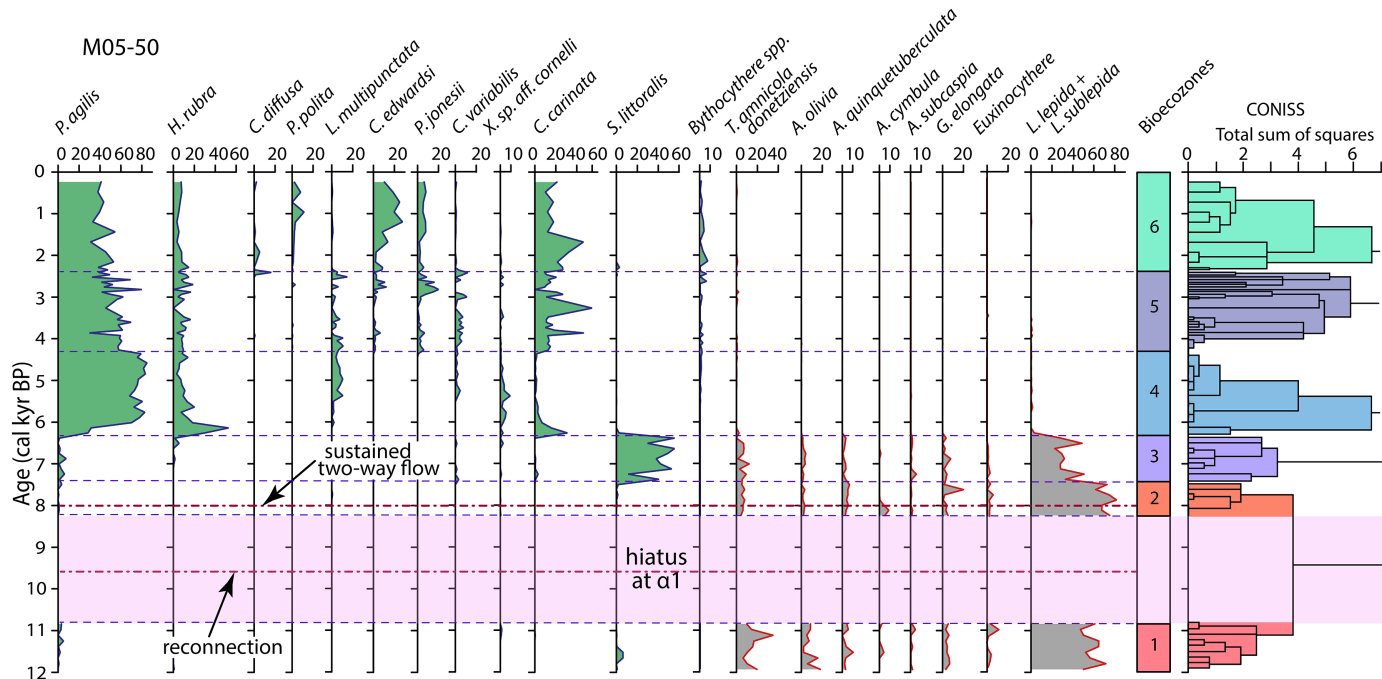


Figure 16

M02-45

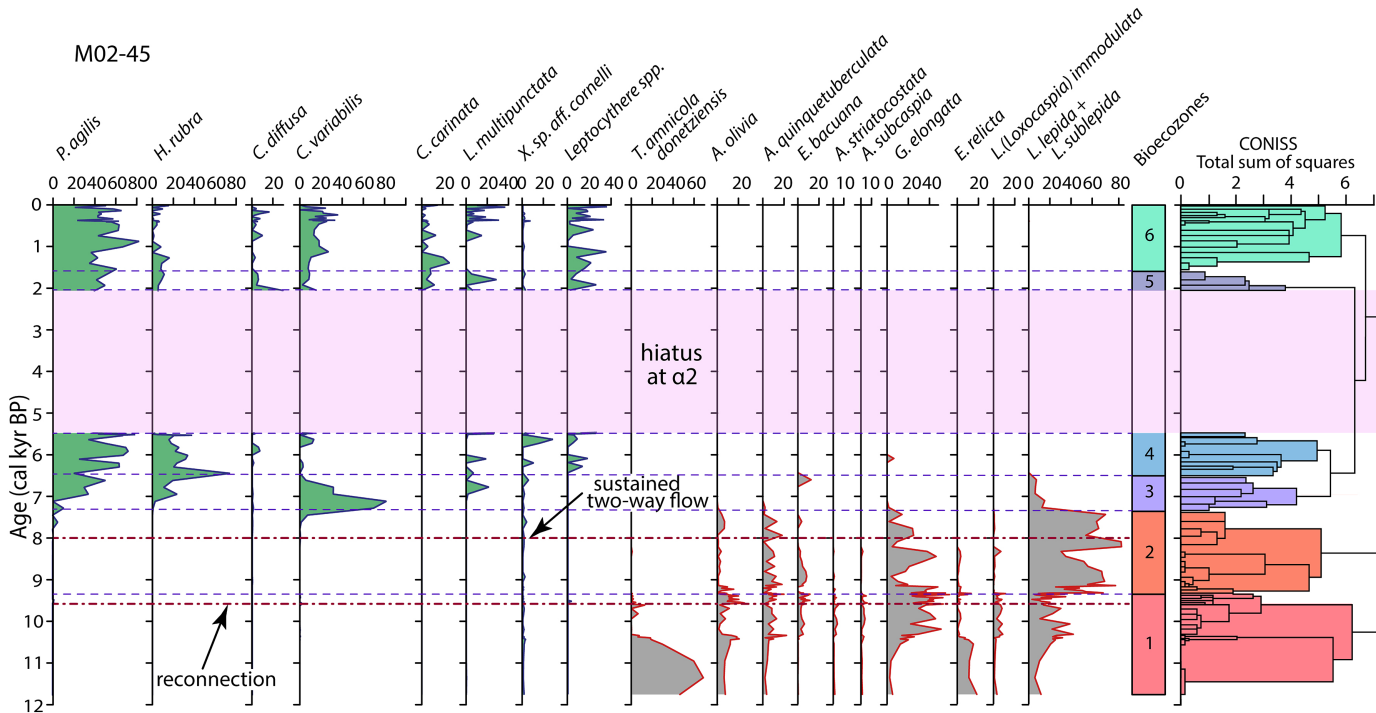


Figure 17

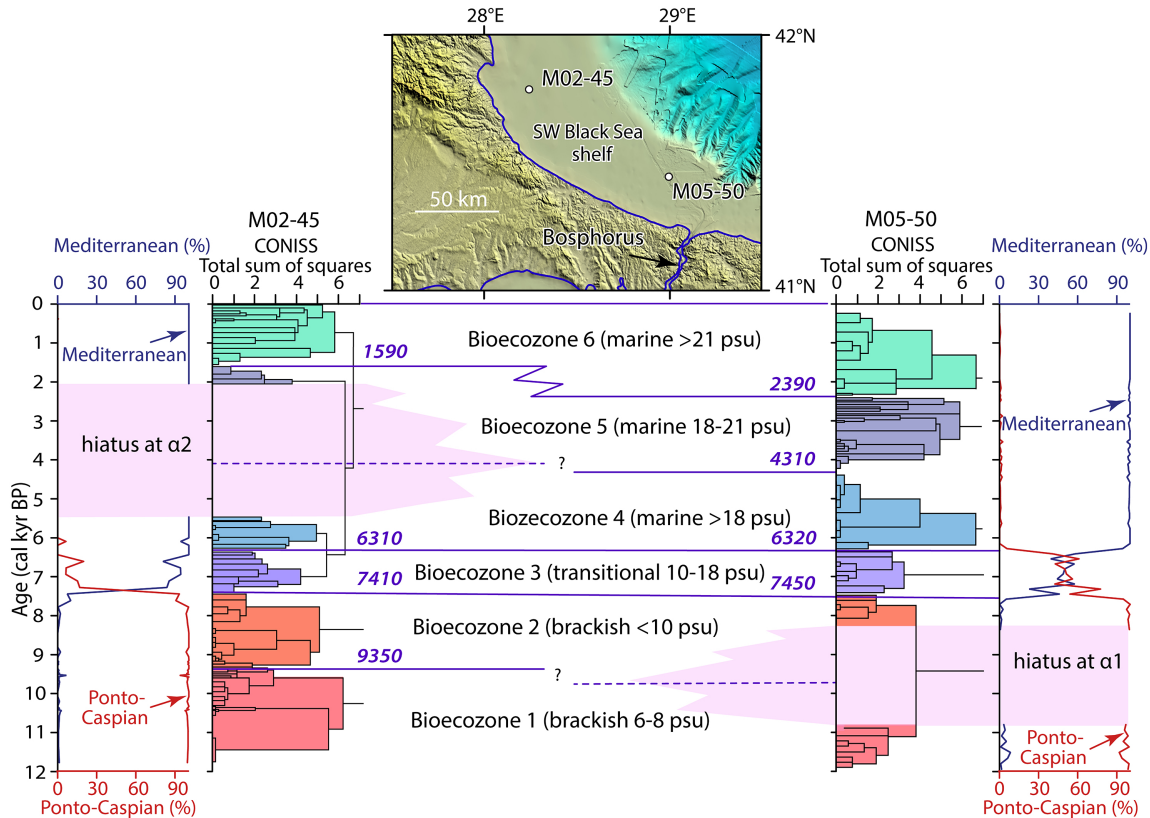


Figure 18