

1 **Identifying the provenance of Leach's storm petrels in the North Atlantic using**
2 **polychlorinated biphenyl signatures derived from comprehensive two-**
3 **dimensional gas chromatography with time-of-flight mass spectrometry**

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25 **Abstract**

26 PCB signatures can be used for source identification, exposure studies, age dating and bio-
27 monitoring. This study uses comprehensive two-dimensional gas chromatography with time-
28 of-flight mass spectrometry (GCxGC-ToFMS) to produce a PCB signature comprised of over
29 80 PCBs for individual Leach's storm petrels (*Oceanodroma leucorhoa*). The Leach's storm
30 petrel is a relatively small, elusive, understudied pelagic bird, which only returns to remote
31 islands under darkness during the breeding season. Samples were obtained from 25
32 Leach's storm petrels found dead in Canada and the UK following storm events in 2006 and
33 2009. Tissue samples were extracted and analysed by GCxGC-ToFMS and results showed
34 that 83 PCB congeners were present in >60% of samples. An assessment of the PCB
35 signature in four different tissue types showed that it did not vary greatly in samples obtained
36 from the gut, heart, liver and stomach. Multivariate statistical analysis identified a distinctive
37 PCB signature in birds from Canada and Europe which was used to identify the regional
38 provenance and transatlantic movement of individual birds. The findings showcase the ability
39 of GCxGC-ToFMS to provide the high quality congener specific analysis that is necessary
40 for PCB fingerprinting, as well as highlighting the potential of PCB signatures for use in
41 ecological studies of movement, foraging and behaviour.

42

43 **Key Words**

44 Polychlorinated biphenyl (PCBs); Leach's storm petrel; Comprehensive two-dimensional gas
45 chromatography, Time of flight mass spectrometry; Chemical fingerprinting.

46 **1. Introduction**

47 **1.1 PCB distribution and signatures in animals**

48 Polychlorinated biphenyls (PCBs) are a group of 209 man-made compounds that were first
49 synthesised in the late 1800s and commercially produced in 1929 (Johnson et al., 2006).
50 They were used extensively throughout the 20th century for a variety of industrial uses. PCB
51 production in the United States peaked in 1970 (Durfree et al., 1976). However, production
52 decreased steadily throughout the 1970's due to a better understanding of the health and
53 environmental risks associated with PCBs. Phasing out began in 1976 in the United
54 Kingdom (UK) (Creaser et al., 2007) and in 1977 in Canada (Environment Canada, 2013).
55 Today policy is largely conducted within an international framework, e.g. the Stockholm
56 convention on persistent organic pollutants aims to eliminate PCB production and use and
57 achieve environmentally sound management of PCBs by 2028 (UNEP, 2013). While PCBs
58 have been largely phased out of commercial/industrial use, they are highly persistent in the
59 environment and are still used in some countries in closed system applications, such as
60 dielectric fluids in electrical equipment. Despite the reduction in PCB inventories and
61 implementation of legislative controls on PCB use, releases to the environment still occur.
62 Coupled with the high persistence of PCBs means they remain contaminants of concern
63 which are found in organisms all over the globe. Investigations involving PCBs often focus
64 on determining the concentrations of the most toxic PCBs (the 12 dioxin like congeners
65 (WHO12)) and/or the most commonly detected PCBs (the European Union 7 indicator
66 congeners (EC7)). Whilst this may be appropriate when determining a health risk or
67 performing simple screening exercises, potentially useful data on the PCB signature is lost
68 as only a fraction of the total number of PCBs present are quantified. Through appropriate
69 sample preparation and analysis by comprehensive two-dimensional gas chromatography
70 with time-of-flight mass spectrometry (GCxGC-ToFMS), over 130 PCBs have been detected
71 within tissue samples (whiting liver) and used to create a detailed PCB signature (Megson et
72 al., 2013a).

73 PCBs can enter the environment through intentional discharges, unintentional spillages and
74 leaks and aerial deposition. Once they have been released into the environment they can
75 undergo further cycling and long range transport. However, the global distribution of PCBs is
76 far from homogenous and different regions of the globe have different total PCB
77 concentrations and specific PCB signatures (Jaspers et al., 2013; Meijer et al., 2003).
78 Variations in PCB signatures have been recorded in a wide variety of different animals
79 (Hansen, 1999; Jaspers et al., 2013), which are believed to be primarily linked to the diet.
80 Specific signatures have been recorded for different species of birds that consume various
81 prey, e.g. fish, insects, mammals and other birds (Hansen, 1999; Jaspers et al., 2006).
82 Variations in PCB signatures have also been used to identify different sub-populations of the
83 same species feeding at different trophic levels. This has been demonstrated for Arctic
84 mammals such as seals and walruses as well as seabirds (Hansen, 1999; Muir et al., 1995;
85 Roscales et al., 2011).

86 Most PCBs are present in animals in relatively low concentrations. This has often restricted
87 investigations to techniques involving destructive tissue sampling so that analysis can be
88 undertaken on lipid rich tissue or eggs. Among ornithological research, novel techniques,
89 such as the analysis of feathers, have been used for non-destructive biomonitoring but only
90 the most abundant PCB congeners are commonly detected (Dauwe et al., 2005; Jaspers et
91 al., 2007). Therefore, due to ethical reasons and analytical limitations, few studies have used
92 birds to investigate regional and geographical patterns of PCB contamination. Most studies
93 have focused on non-migratory passerine species such as starlings (Eens et al., 2013), as
94 they are a non-migratory and, as such, are well suited for monitoring local contamination.
95 Contamination profiles are expected to better reflect local contamination because of their
96 relatively small home ranges, territories and foraging areas (Eens et al., 2013). Less
97 research has been undertaken on the PCB signature of hard to study species such as
98 seabirds that operate over very large spatial scales.

99 1.2 Leach's storm petrel

100 The Leach's storm petrel (*Oceanodroma leucorhoa*) is a small (wingspan 450 to 480 mm,
101 weight 35 to 45 g) pelagic bird that breeds on remote islands (Huntingdon et al., 1996).

102 Despite being globally abundant (>10 million breeding pairs), elusive habits such as
103 nocturnal visits to colonies and pelagic foraging mean that aspects of its ecology remain
104 unknown. In the North Atlantic there are breeding colonies in North America and Western
105 Europe (Figure 1). Newfoundland, Canada supports the largest breeding colonies
106 (Huntingdon et al., 1996; Robertson et al., 2006) and the European colonies are
107 predominantly divided between two small island archipelagos in Iceland (Vestmanyjaer) and
108 Scotland (St Kilda) (Mitchell et al., 2004).

109 Although they spend much of their time at sea, large numbers of Leach's storm petrel can be
110 driven onshore during severe storm events. Many of these birds are discovered either dead
111 or moribund, which presents an opportunity to undertake detailed assessments on
112 carcasses and investigate their origin. During 2006 and 2009 a series of storm events in
113 waters around the UK and Canada drove many Leach's storm petrels inland. Twenty five
114 carcasses were obtained from wrecked birds that had been killed by these storms and
115 subsequently recovered by members of the public. Twelve were recovered from
116 Newfoundland and 13 from the UK. It was unclear if the wrecked birds discovered in the UK
117 and Newfoundland were from local colonies, or from a combination of different breeding
118 colonies widely spread across the North Atlantic. Tissue and feather samples were obtained
119 from these wrecked birds and assessments of provenance were undertaken using highly
120 branched isoprenoid (HBI) concentrations and stable isotope ratios ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$). Analysis
121 of HBI concentrations was able to distinguish between birds recovered from the UK and
122 Newfoundland (see Table 1). HBIs provide recent dietary insights and the results indicate
123 that the birds wrecked in the two areas were feeding locally in the weeks preceding the
124 storms (Brown et al., 2013). Stable isotope ratios in a feather are linked to the prey
125 consumed during the growth phase of that feather; therefore ratios are often used for

126 tracking the dispersal of migrant wildlife (Hobson, 2007). However, the results for feathers
127 obtained from the birds used in this study were inconclusive due to the mixture of feathers
128 available from the wrecked birds and similarity in the signatures from the two sub-
129 populations. (see Table 1) (Bicknell, 2011).

130 **1.3 Aim**

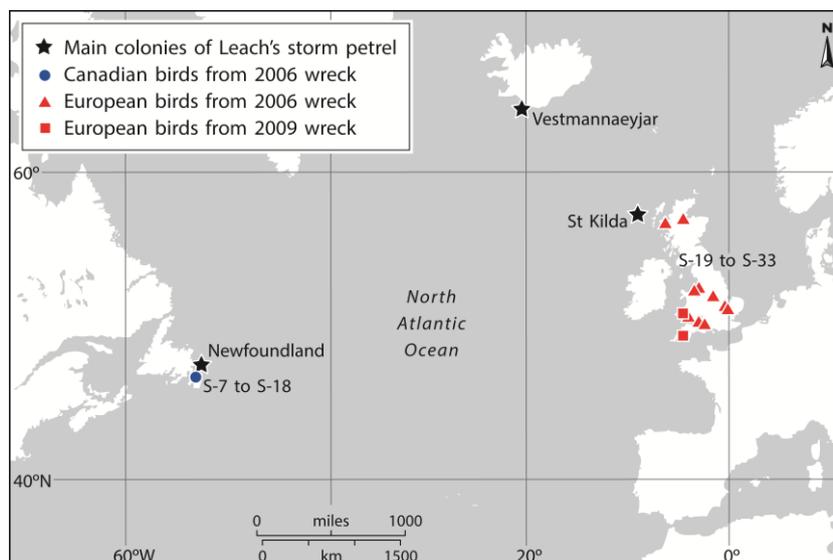
131 Among marine predators, seabirds have been proposed as useful bio-indicators for PCBs
132 and other persistent organic pollutants, mainly because they are positioned at high trophic
133 positions, breed at specific locations and are widely distributed (Burger and Gochfeld, 2004).
134 However, much of this research on seabirds has focused on coastal species due to the
135 ethical and technical limitations of sampling strategies associated with pelagic species
136 (Elliott, 2005; Yamashita et al., 2007). This paper assesses PCBs in the highly pelagic
137 Leach's storm petrel using a recently reported GCxGC-ToFMS method for fingerprinting
138 PCBs in environmental samples (Megson et al., 2013a). This study aims to provide valuable
139 baseline data on PCBs in Leach's storm petrels from the North Atlantic and demonstrate the
140 potential of using PCB signatures as a tool for identifying the provenance of individual birds.

141 **2 Methodology**

142 **2.1 Sample collection and preparation**

143 The same set of carcasses that were analysed in previous studies undertaken by Bicknell
144 (2011) and Brown et al. (2013) were used in this study. This comprised 12 birds from
145 Newfoundland that were driven onshore during a storm event in early September 2006 (S-7
146 to S-18), 11 birds from the UK that were recovered after a storm in December 2006 (S-21 to
147 S-33) and two birds that were recovered from the UK after a storm in December 2009 (S-19
148 and S-20) (Figure 1). All carcasses were discovered by members of the public and stored at -
149 80 °C prior to sampling in 2011. Individual organs could not be removed from the majority of
150 the samples due to partial decomposition. However, for one bird (S-22), decomposition was

151 not as severe and four specific organs were removed for analysis. Two organs were also
152 removed from S-23 and S-24.



153

154 Figure 1. Location of the main Leach's storm petrel colonies and the sites from which
155 wrecked birds were recovered during storms in 2006 and 2009. Twelve birds were recovered
156 from Newfoundland (S-7 to S-18) and 13 birds from the UK in 2006 (S-21 to S-33) & 2009
157 (S-19 and S-20)

158 2.2 Extraction

159 Sample extraction was undertaken following the established method for PCB extraction in
160 tissues reported by Megson et al. (2013a) and outlined in Brown et al. (2013). All samples
161 were freeze-dried (-45 °C; 0.2 mbar; 72 h) and ground into a powder, internal standards
162 were added and an organic extract obtained by adding dichloromethane/methanol (2:1 v/v)
163 and ultrasonication (8 x 10 min). Extracts were filtered, dried and re-suspended in hexane
164 before being separated into a non-polar fraction by column chromatography (SiO₂). Samples
165 were blown down to approximately 50 µL using nitrogen, left overnight in a clean
166 environment to evaporate to dryness and reconstituted with 10 µL of ¹³C₁₂ internal standard
167 (CIL-EC-5370 EN-1948-4 PCB sampling standard, LGC) and 90 µL of hexane prior to
168 analysis by GCxGC-ToFMS.

169 2.3 GCxGC-ToFMS Analysis

170 2.3.1 Analytical procedure

171 Samples were analysed using the method described by Megson et al. (2013a) using a time-
172 of-flight mass spectrometer, (LECO, St. Joseph, MI Pegasus 4D) coupled to a two
173 dimensional gas chromatograph (Agilent Technologies 7890A) equipped with a thermal
174 modulator (LECO, St. Joseph, MI). The gas chromatograph was installed with a Rtx-PCB (60
175 m x 0.18 mm x 0.18 μm) ¹D column and a Rxi-17 (1.5 m x 0.1 mm x 0.1 μm) ²D column. A
176 sample volume of 1 μL was injected in splitless mode. All data files were processed using
177 ChromaTOF software set to identify 10,000 peaks with a signal-to-noise ratio of > 10:1.
178 Throughout this paper PCBs are referred to using the Guitart numbering system (Guitart et
179 al., 1993).

180 2.3.2 Data Quality

181 Analytical blanks were run with each batch of approximately 10 samples. All samples were
182 spiked with a ¹³C₁₂ internal standard (CB-60, CB-127, CB-159) which was used to quantify
183 PCB concentrations by isotope dilution. Concentrations were normalised to dry weight tissue
184 mass and are therefore reported as ng g⁻¹. As samples were originally extracted for the
185 analysis of HBIs, PCB recovery could not be accurately determined for each sample;
186 therefore reported concentrations were not corrected based on sample recovery or lipid
187 corrected. However, application of this method to other tissue samples (such as fat and
188 blood) for the determination of PCBs consistently recorded recoveries in the range 30 – 60%
189 (unpublished data), which meets the recovery requirements of US EPA method 1668C.
190 Experiments using a 50:50 mixture of A1254:A1016 (at 500 $\mu\text{g L}^{-1}$ total PCBs) showed no
191 significant loss from the blow-down procedure for any of the PCBs analysed (recovery of 101
192 \pm 3.4 %; 1 standard deviation). Limits of detection for individual PCBs were in the range 0.1 -
193 5 ng g⁻¹ (dry weight). Accuracy and precision were measured for the sum of the European
194 Union 7 indicator congeners (EC7) (CB-28, CB-52, CB-101, CB-118, CB-138, CB-153, CB-

195 180) by analysing a 10 mg L⁻¹ Aroclor 1248 standard three times. The accuracy of the sum
196 of the EC7 congeners for the three samples was 105 ± 0.9 % (1 standard deviation).

197 **2.4 Statistical analysis**

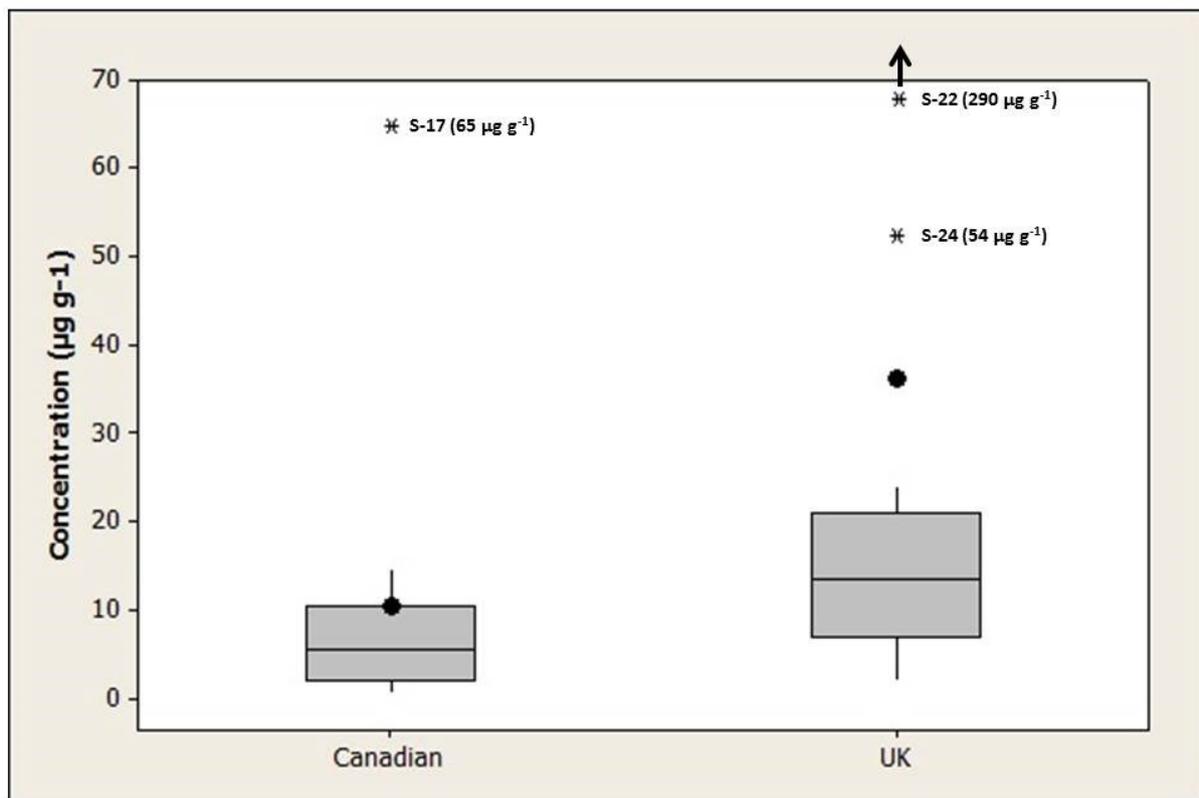
198 The results for the 25 storm petrels were subjected to principal component analysis (PCA).
199 For the birds where individual organs were removed the results from each organ were
200 included in the analysis. The samples denoted with; 'a' were obtained from the liver, 'b' from
201 the stomach, 'c' from the guts and 'd' from the heart. Where a PCB was not detected it was
202 included in the dataset as a '0'. As part of the data quality check, other values were
203 substituted for '0', but these had no observable effect on the data output and so the '0's were
204 retained. To reduce any bias from a high proportion of non-detects for a specific congener,
205 PCBs that were not detected in over 60% of samples (i.e. PCBs present in less than 18 out
206 of the 30 samples) were removed from the analysis following the guidance of Helsel (2006).
207 The resultant data set contained 30 samples and 83 PCBs. Before performing PCA the data
208 were normalised by transformation to a percent metric to remove concentration/dilution
209 effects. The data were then mean centred and scaled using a Z-transform (autoscale
210 transform) to prevent high concentration variables from dominating the analysis (Johnson et
211 al., 2007). The first three principal components explained 65.5% of the variance in the data
212 set. Scatter plots showing goodness of fit on a congener-by-congener basis are shown in
213 Supplementary Information (S1). These justify the use of a three principal component model
214 over a two principal component model as they show the improvement in the goodness of fit
215 for the more chlorinated congeners.

216 **3 Results**

217 **3.1 PCBs in the Leach's storm petrel**

218 The most dominant PCBs encountered in the samples were CB-153, CB-118, CB-138 and
219 CB-180. In each sample these accounted for approximately 30%, 10%, 10% and 10% of the
220 total PCB load respectively. PCB concentrations were calculated for the European Union 7

221 indicator (EC7) congeners and varied from 0.6 $\mu\text{g g}^{-1}$ (S-12) to 290 $\mu\text{g g}^{-1}$ (S-22). Total
222 concentrations of the EC7 congeners appeared to be greater in the birds found in the UK
223 (mean value of 36 $\mu\text{g g}^{-1}$) compared with the birds found in Canada (mean value of 11 $\mu\text{g g}^{-1}$)
224 ¹), although these differences were not statistically significant (Figure 2).



225

226 Figure 2. Box and whisker plots of the sum of European Union 7 indicator (EC7) congeners
227 in Leach's storm petrels found in Canada and the UK. The box shows the interquartile range
228 (25th percentile to 75th percentile), with the median as a horizontal line across each box and
229 the mean displayed as a black dot. Whiskers (lines) represent 1.5 times the interquartile
230 range and samples beyond the interquartile range are plotted as individual points.

231 The PCB concentrations are reported along with the results of the previous investigations
232 undertaken by Bicknell (2011) and Brown et al. (2013), in Table 1. While no correlation
233 between the PCB concentrations and stable isotope data was observed, there was a strong
234 positive correlation between the PCB and HBI concentration data (R^2 value of 0.7 and P

235 value of 0.006). Where possible the sex of the bird was also determined, however this was
 236 not well correlated with the stable isotope, HBI or PCB data. PCB concentrations in birds
 237 wrecked from the 2009 storm were slightly lower than the birds from the 2006 storm;
 238 however there was no observable difference in the PCB signatures (see Section 3.3).

239 Table 1. Concentrations of Σ EC7 PCBs, HBIs (Brown et al., 2013) and stable isotope ratios
 240 from Leach's storm petrels from the identified subgroups (Bicknell, 2011), concentrations
 241 from separate organs of the same bird were averaged.

Sample	Σ EC7 PCBs ($\mu\text{g g}^{-1}$)	Total HBIs (ng g^{-1})	$\delta^{15}\text{N}$ (‰)	$\delta^{13}\text{C}$ (‰)	Sex (M/F)
Recovered from Canada					
S-7	3.1	-	15.3 ^a	-20.0 ^a	F
S-8	2.7	-	13.2 ^a	-17.7 ^a	F
S-9	6.0	0.22	15.2	-18.8	F
S-10	6.4	0.45	14.6	-19.0	F
S-11	1.8	0.21	14.2	-18.8	F
S-12	0.6	0.17	14.9	-19.2	F
S-13	5.0	-	14.9 ^a	-19.9 ^a	F
S-14	15	-	14.6 ^a	-17.9 ^a	F
S-15	1.0	-	-	-	F
S-16	9.7	-	14.7 ^a	-18.9 ^a	F
S-17	65	-	14.3 ^a	-19.1 ^a	F
S-18	11	0.42	13.7	-19.1	M
Average (1 σ)	10.6 (+/- 17.6)	0.29 (+/- 0.13)	14.5 (+/- 0.58)	-19.0 (+/- 0.19)	
Recovered from the UK					
S-22	290	4.7	15.0	-19.2	-
S-23	16	2.2	15.4	-19.1	-
S-24	52	4.2	14.7	-20.7	F

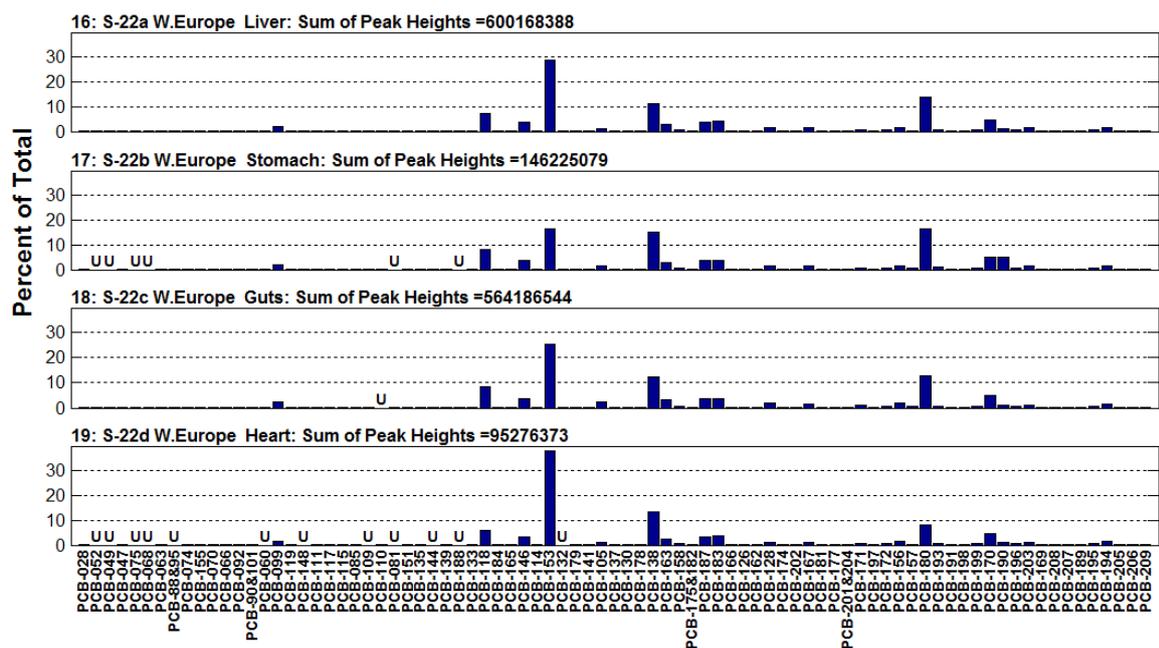
S-25	24	2.4	14.7	-19.1	M
S-26	15	1.2	14.2	-18.9	F
S-29	18	2.9	12.2	-19.1	F
S-19	4.3	-	13.4	-17.6	-
S-20	2.1	-	14.7	-18.3	-
S-21	6.6	2.7	14.2	-18.6	M
S-27	14	4.3	13.8	-19.8	-
S-30	7.4	0.9	14.6	-18.9	M
S-32	8.1	-	14.4	-19.1	F
S-33	13	2.6	12.7	-19.0	M
Average (1 σ)	36.2 (+/- 77.3)	2.8 (+/- 1.3)	14.2 (+/-0.92)	-19.0 (+/- 0.72)	

242 - = not analysed

243 ^a = Sample collection was targeted towards newly grown feathers (produced whilst the birds
244 were at their respective breeding grounds); however these birds had not finished moulting
245 and therefore the sample had to be obtained from an old tail feather rather than new one.

246 3.2 PCB signatures in different organs

247 Individual organs could not be removed from the majority of the samples due to partial
248 decomposition. However, where different organs could be removed the PCB signature
249 appeared to be similar in each organ, although the stomach contained higher proportions of
250 CB-190 and depleted proportions of CB-153 (Figure 3). The covariance in the signature of
251 the different organs can also be observed in the PCA scores plot (see Figure 4).

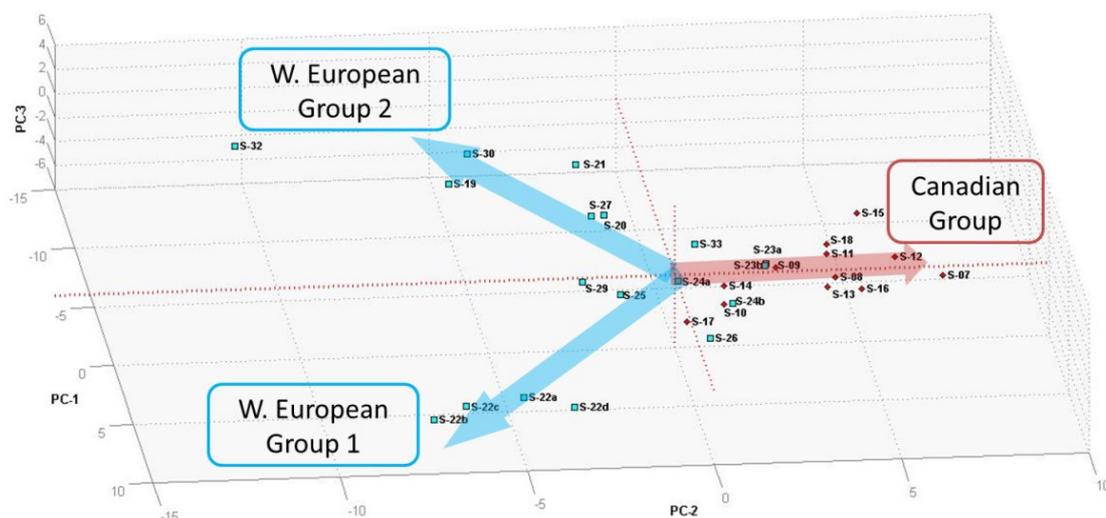


252
 253 Figure 3. Congener plots showing differences in signature in liver, stomach, guts and heart
 254 from the same Leach's storm petrel found wrecked in the UK. 'U' represents a PCB present
 255 at a concentration below the limit of detection.

256 3.3 Identifying Leach's storm petrels wrecked in Western Europe and Canada

257 Three main groups of birds were identified through principal component analysis (Figure 4),
 258 with a gradation/mixing between the groups. The three groups were labelled as; Canadian
 259 group, Western European group 1 and Western European group 2. The Canadian group
 260 were separated by a positive score on principal component 2, whereas birds recovered from
 261 the UK predominantly had a negative score on principal component 2. The birds found in the
 262 UK were further subdivided as Western European group 1, based on a positive score on
 263 principal component 1 and Western European group 2 based on a negative score on
 264 principal component 1. A similar 3 end member system was also produced when the data
 265 were assessed using the unmixing model, polytopic vector analysis. This is a self-training,
 266 receptor modelling technique that can be used to resolve the following in a multivariate
 267 mixed chemical system; the most likely number of end members, the composition of each
 268 end member and the relative proportions of each end member in a sample (Johnson et al.,

269 2007). A ternary diagram of the mixing model results from the polytopic vector analysis is
270 presented in Supplementary Information (S2).

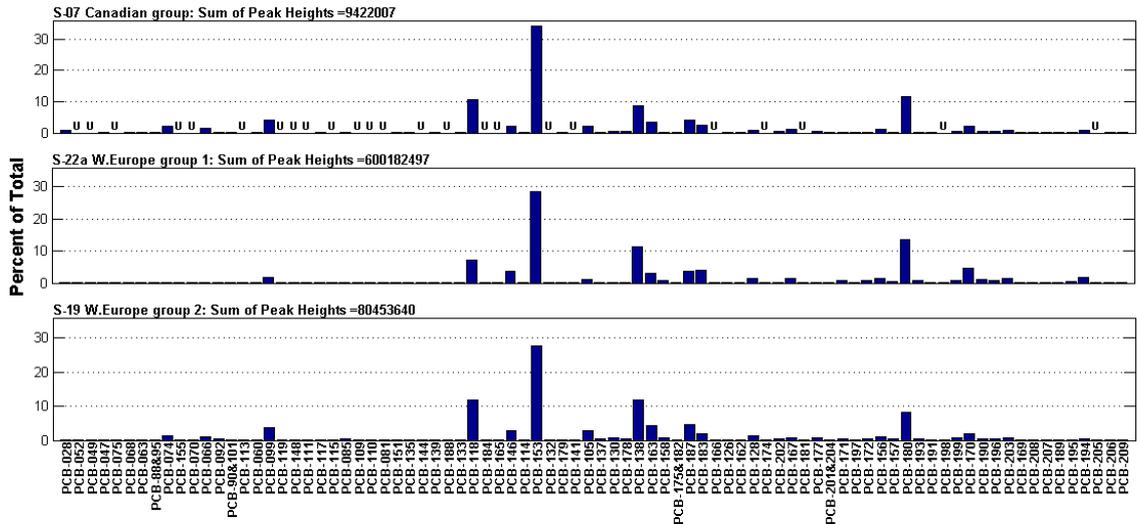


271

272 Figure 4. PCA scores plot showing three main groups of birds. Red dots represent birds
273 found in Canada and blue dots represent birds found in the UK.

274

275 Although overlap between the different groups is apparent, the results reveal differences in
276 the PCB signature in the birds recovered from Canada and the UK. The birds recovered
277 from the UK had higher proportions of CB-138, CB-163, CB-187 and CB-184, whereas the
278 birds recovered from Canada tended to have higher proportions of CB-153. The results also
279 split the birds recovered from the UK into two sub-groups. Western European group 1
280 generally contained higher proportions of the more chlorinated congeners; CB-170, CB-180,
281 CB-183 and CB-194 whereas the Western European group 2 birds generally contained
282 higher proportions of the less chlorinated congeners; CB-66, CB-74, CB-99 and CB-105
283 (Figure 5).



284

285 Figure 5. Bar charts of representative PCB signatures from each of the three groups; S-07
 286 from the Canadian group, S-22a from the W. European group 1 and S-19 from the W.
 287 European group 2.

288 **4 Discussion**

289 **4.1 PCB signatures in different organs**

290 Analysis of individual organs obtained from the same Leach's storm-petrel showed that the
 291 PCB signature did not vary greatly between the liver, stomach, guts and heart (Figure 3).
 292 This is consistent with previous research which has shown that a similar PCB signature was
 293 present in a variety of different tissue samples analysed from the same bird (Boumphrey et
 294 al., 1993). The main difference in the signature of organs analysed in this study was
 295 observed in the stomach which contained higher proportions of CB-190 and depleted
 296 proportions of CB-153. As the contents of the stomach were not completely removed prior to
 297 extraction, this difference could be associated with the PCB signature of undigested food
 298 within the stomach.

299 Although the PCB signature remained relatively constant between different organs the total
300 concentrations of the EC7 PCBs were more variable. The highest concentration was
301 recorded in the liver ($580 \mu\text{g g}^{-1}$), which was higher than concentrations recorded in the
302 stomach ($150 \mu\text{g g}^{-1}$), guts ($240 \mu\text{g g}^{-1}$) and heart ($210 \mu\text{g g}^{-1}$). The relative similarity of the
303 PCB signature in different organs (Figure 3) demonstrates that comparisons between birds
304 can be made irrespective of the tissue type sampled. Nonetheless it is preferable to use the
305 same tissue type if direct comparison of PCB concentrations between samples is required.
306 In this study the highest total PCB concentrations were obtained from the liver, which
307 suggests that it is a good tissue type for future studies. Using the liver should provide a
308 higher number of PCBs to be detected compared to other tissue types, leading to a more
309 informative PCB signature.

310 **4.2 Distinguishing differences among individual Leach's storm petrels**

311 Previous attempts to distinguish differences among the 25 wrecked Leach's storm petrels
312 have involved the analysis of stable isotopes and HBIs. While stable isotopes showed no
313 clear differentiation in feather signatures (Bicknell, 2011), interpretation of HBIs provided a
314 division of the samples obtained from Canada and the UK (Brown et al., 2013). In the current
315 study, the application of principal component analysis to PCB data obtained by GCxGC-
316 ToFMS provided additional information. Firstly, principal component analysis of PCB
317 signatures not only distinguished between birds that were collected from Canada and the UK
318 but also identified a further sub-division in the European birds (Figure 4). The HBI analysis
319 showed that the total concentrations and relative distributions of HBI isomers were similar for
320 all Leach's storm petrels recovered from the UK. HBIs provide dietary insights for relatively
321 short periods of time (e.g. <1 month (Brown and Belt, 2012)) and so this relative similarity
322 was interpreted as being consistent with dietary contributions in the weeks prior to the birds
323 being wrecked in December (Brown et al., 2013). This period coincides with the breeding /
324 fledging period (between late May and November) and indicates that during this period the
325 birds were all feeding in a similar area. In contrast to HBIs, PCBs are known to be highly

326 persistent in animals and so the signature is representative of many years exposure
327 throughout the animal's lifetime (Jaspers et al., 2013). The main exposure pathway for
328 animals is usually linked to their diet; therefore variations in feeding patterns could explain
329 the differences in the PCB signatures observed in Figure 4.

330 The diet of the Leach's storm petrel predominantly comprises small fish and zooplankton,
331 although feeding preference has been shown to vary slightly throughout the fledging period
332 and is based on the availability of different food sources throughout the year (Hedd et al.,
333 2009). In Newfoundland fish were identified as the preferential food source and comprised
334 60 - 90% of a storm petrel's diet. Results from Hedd et al. (2009) showed that mature
335 lanternfish (myctophids) which vertically migrate from the mesopelagic during the night were
336 the most consumed food source (78% of identified fish). The higher concentrations of PCBs
337 and higher proportions of the more chlorinated congeners are both indicators that the
338 Western European group 1 birds were feeding on prey from higher tropic levels (Bentzen et
339 al., 2008; Muir et al., 1995), and/or prey from the mesopelagic and deep sea rather than
340 surface water species (Roscales et al., 2011). Analysis of stable isotopes in the blood of
341 breeding Leach's storm petrels revealed regional differences in $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, attributed to
342 potential differences in food-web structure/length in each region (Bicknell et al., 2013). The
343 principal component analysis of the PCB signatures supports this hypothesis by indicating
344 the three groups of birds are representative of sub-populations consuming prey from
345 different trophic levels and/or from different regional ecosystems.

346 Biometric variability may also influence the PCB signature of birds. Whilst this study was
347 able to demonstrate that there was no gender based variation, any age related variation
348 could not be investigated because the age of the birds was not known. PCB signatures in
349 long-lived animals, such as humans, have been shown to vary over time due to variable
350 exposure and the subsequent biotransformation and elimination of less persistent congeners
351 (Megson et al., 2013b; Quinn and Wania, 2012). As a result, older individuals tend to have
352 higher total PCB concentrations which are dominated by the more chlorinated congeners.

353 Whilst this could indicate that the observed sub-grouping of the European birds was a
354 function of age, the absence of a similar division in the Canadian birds, which might also be
355 expected to contain age variation, suggests that this is unlikely.

356 **4.3 Evidence for ocean-wide movement of Leach's storm petrels**

357 There was overlap between the two groups of birds recovered from Newfoundland and the
358 UK. Although S-17 was collected in Newfoundland, PCA revealed it had a PCB signature
359 representative of European birds. In addition to the PCA assignment, this sample also
360 contained higher total PCB concentrations than the other Canadian birds, which was more
361 representative of European birds. This suggests that S-17 was originally from Europe but
362 had migrated across the Atlantic.

363 There were also four birds (S-23, S-24, S-26 and S-33) collected in the UK that have a
364 similar PCB signature to the Canadian birds. This would suggest these birds originated from
365 the Newfoundland region of Canada. However, as the HBI data for these four birds is
366 consistent with the other birds recovered from the UK it also suggests they were feeding
367 around Western Europe in the weeks leading up to the storm. It is therefore likely that these
368 birds had already migrated to Europe more than a month prior to the storm that
369 subsequently killed them.

370 These findings provide further evidence of regular movement of individual Leach's storm
371 petrels across the North Atlantic, and the high level of connectivity between regions and
372 colonies as indicated by previous stable isotope and genetic studies (Bicknell et al., 2012;
373 Bicknell et al., 2013).

374 **5 Conclusions**

375 Analysis of tissue obtained from 25 wrecked storm petrels by GCxGC-ToFMS was used to
376 produce a comprehensive data set with 83 specific PCB congeners present in >60% of
377 samples. Analysis of different organs from the same bird showed that the PCB signature did

378 not vary greatly in samples obtained from the gut, heart, liver and stomach. The data set was
379 interrogated by multivariate statistical analysis which identified different PCB signatures in
380 birds recovered from Canada and the UK. The differences in PCB signatures are believed to
381 be representative of sub-populations consuming prey taxa from different trophic levels
382 and/or utilising different feeding locations although possible influences due to age could not
383 be discounted. There was some overlap in the PCB signatures of birds recovered from
384 Canada and the UK, thereby providing further evidence of regular movement of individual
385 Leach's storm petrels across the North Atlantic and a high level of connectivity between
386 regions and colonies.

387 The results of this study show how PCB fingerprinting can be a useful tool to study the
388 provenance, geographical movement, and feeding habits of animals such as the Leach's
389 storm petrel. As with any fingerprinting exercise, the most reliable conclusions are drawn
390 from multiple lines of evidence. Previous investigations have shown how HBI and stable
391 isotope analysis of blood and feathers can be used to assess recent movement of
392 individuals. In this study PCBs have been used to identify differences over longer time
393 scales.

394 The findings highlight the ability of GCxGC-ToFMS to provide the high quality congener
395 specific analysis that is necessary when comparing PCB signatures. This work builds on
396 previous studies using PCB signatures in birds by successfully applying the technique to an
397 understudied pelagic species utilising a large territory and foraging area.

398 **Acknowledgements**

399 The authors would like to thank Ann Tanderitispole and Chris Gallagher (University of
400 Strathclyde) for their assistance with sample analysis and the Scottish Funding Council and
401 EPSRC Grant EP/D013739/2 for funding associated with the University of Strathclyde
402 Laboratory. Alec Kettle (Leco) for his help and support with the research. Thanks to Greg
403 Robertson at Canada Wildlife Services, Newfoundland for supplying the samples from the

404 Canadian wreck and members of the public that kindly sent in corpses found in the UK.
405 Finally, David Megson would also like to thank Plymouth University for funding this project as
406 part of his PhD research.

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